Using AB Designs with Nonoverlap Effect Size Measures to Support Clinical Decision Making: A Monte Carlo Validation

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Abstract

Single-case experimental designs often require extended baselines or the withdrawal of treatment, which may not be feasible or ethical in some practical settings. The quasi-experimental AB design is a potential alternative, but more research is needed on its validity. The purpose of our study was to examine the validity of using nonoverlap measures of effect size to detect changes in AB designs using simulated data. In our analyses, we determined thresholds for three effect size measures beyond which the type I error rate would remain below .05, and then examined if using these thresholds would provide sufficient power. Overall, our analyses show that some effect size measures may provide adequate control over type I error rate and sufficient power when analyzing data from AB designs. In sum, our results suggest that practitioners may use quasi-experimental AB designs in combination with effect size to rigorously assess progress in practice.

Keywords: AB design, effect size, power, single-case design, type I error rate, validity.
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Researchers from diverse fields of psychology have recommended that practitioners adopt single-case experimental designs to assess and monitor individual progress (Foster, Watson, Meeks, & Young, 2002; Lundervold & Belwood, 2011; Machalicek & Horner, 2018; Odom et al., 2005). While single-case experimental designs are suitable for use in research, such arrangements are not always feasible or ethical in practical settings (Engel & Schutt, 2017; Janosky, Leininger, Hoerger, & Libkuman, 2009). For example, the demonstration of experimental control in reversal and alternating-treatment designs is done through the repeated application and removal of treatment. In the case of aggressive behaviors or self-injury, this reversal may neither be desired or ethical (Hayes, 1981). The multiple baseline design, on the other hand, typically involves collecting a high number of baseline (i.e., no treatment) sessions prior to implementing treatment, which may be inadvisable for high-risk behavior. Moreover, agencies may not have the time or the funding to continue data collection for an extended period of time (Lanovaz, Turgeon, Cardinal, & Wheatley, 2018).

From a research standpoint, AB designs are limited by their inability to demonstrate a functional relationship and experimental control. From a practice standpoint, however, identifying the precise mechanisms responsible for a behavior change is secondary to demonstrating if a significant change in behavior has occurred from phase A to phase B. Therefore, when conducting clinical evaluations one option is for practitioners to use the quasi-experimental AB design to guide their clinical decision making (Lanovaz et al., 2018). Contrary to its experimental counterparts, the AB design never requires the withdrawal of treatment, nor the completion of additional baseline sessions beyond achieving data stability. That said, not conducting a replication may increase the probability of reaching incorrect
conclusions regarding the data. Some methods of analysis have been developed to control for error rates when analyzing reversal or multiple baseline designs, which can also be readily applied to AB designs (e.g., Bloom, Fischer, & Orme, 1999; Fisher et al., 2003; Manolov, Sierra, Solanas, & Botella, 2014; Nugent, 2001). Although a step in the right direction, these methods do not allow researchers to determine whether within-subject replications would be warranted in practice.

In a recent study addressing these issues, Lanovaz et al. (2018) extracted data from 501 reversal ABAB graphs from theses and dissertations and examined to what extent the results observed following the first phase change matched the results observed following the subsequent phase changes. In approximately 5 of 6 cases, the results of the first phase change was consistent with those of the subsequent phase changes, suggesting that AB designs would lead to erroneous clinical decisions approximately 15% of the time if a replication had not been conducted. To further reduce this error rate, Lanovaz et al. (2018) conducted an analysis, which showed that nonoverlap effect size measures may function as predictors of replication and be used as thresholds to bring type I error rates down to around 5%.

An effect size provides information on the size, magnitude, or meaningfulness of a change associated with an intervention. Although there are several accepted and tested statistics commonly used in group designs (e.g., standardized mean difference indices; Cohen, 1992), these effect size measures are not appropriate for use with single-case designs. Single-case design data typically contain a small number of data points, data are often autocorrelated, and normal distribution of the data cannot be assumed (Hersen & Barlow, 1976; Parker, Vannest, & Brown, 2009). For these reasons, researchers and practitioners typically rely on a group of indices called nonoverlap measures. These indices do not require data to have a specific distribution or scale type (Vannest & Ninci, 2015). Instead,
nonoverlap indices yield an effect size based on the presence or absence of overlap in the data points from contrasted phases.

Despite the findings of Lanovaz et al. (2018), additional research is needed to provide guidance for using effect size as a predictor of the need for replication. First, Lanovaz et al. (2018) based their analyses on nonsimulated data so that the patterns would mimic those of behavior typically observed in practice. Using nonsimulated data limited the number of datasets (i.e., 501) on which the analysis could be based, which precluded an in-depth analysis of the effects of trend, autocorrelation, and number of points. Second, the dual-criteria method of analysis (see Fisher et al., 2003) applied to each graph had limited power, that may have biased some of the results. Third, the use of nonsimulated data prevented the analysis of power of such an approach without leading to a logical fallacy. As such, the extent to which using AB designs with effect size would lead to false negatives remains unknown.

While several studies have evaluated the performance of nonoverlap measures for calculating effect size in published datasets (e.g., Ma, 2006; Parker et al. 2009; Tarlow, 2007), additional research is needed to examine how effect size may be used to analyze AB designs. The purpose of our study was to extend the study conducted by Lanovaz et al. (2018) by completing a more in-depth analysis of using effect size to detect changes in AB designs using simulated data. Specifically, we (a) established thresholds of effect size beyond which a replication would be deemed unnecessary for different values of autocorrelation and trend, and for varying phase lengths, and (b) assessed if using these thresholds produced sufficient power to detect a true effect.

**Method**

**Data Generation**
We used the arima.sim function of R code to generate raw datasets representative of AB designs for our Monte Carlo simulation of a first-order autoregressive model\(^1\).

Autoregressive integrated moving average (ARIMA) models are statistical models typically used to analyze time series data. ARIMA models can generate datasets that contain a variety of characteristics including trend and autocorrelation, both of which are often present in single-case design data. The malleability of this model makes it ideal for producing simulated datasets. More specifically, we generated the times series using the equation

\[ x_t = a x_{t-1} + \epsilon_t \]  

where \( x \) is equal to a univariate time series, \( t \) is an integer index of time, \( a \) is the autoregressive coefficient (i.e., autocorrelation), and \( \epsilon \) is the error term. The error term had a mean of zero and a standard deviation of 1. We systematically manipulated four parameters; a) number of data points in each phase, b) autocorrelation, c) trend, and d) standardized mean difference between Phase A and Phase B.

Equation 1 only allows the manipulation of number of data points and autocorrelations. Thus, we transformed our initial values using the following formula to add trend, a constant, and a simulated effect.

\[ y^A_t = x^A_t - \beta(t-1) + c \]  

\[ y^B_t = x^B_t + \beta(t - m - 1) - \beta(m - 1) + c + SMD \]  

where \( SMD = \frac{\mu_B - \mu_A}{\sigma} \)

In Equations 2 and 3, the \( y_t \) represents each data point at time \( t \), \( x_t \) the values obtained using Equation 1, and the superscripts A and B the phases to which each point belongs. \( \beta \) adds a reversing trend to the dataset, which decreases during baseline (Phase A) and increases

\(^1\) To facilitate replication, we provide the R code used to generate the data, to measure effect sizes, and to conduct the analyses at: https://osf.io/9br7g/?view_only=8ecddb1554b64e2388c62d792153995b
following treatment (Phase B). We used this reversing pattern because (a) the implementation of treatment may produce changes in trend that should be considered in single-case designs (Kratochwill et al., 2010), (b) researchers and practitioners are unlikely to start a treatment when the trend is already increasing (if the purpose is to increase the target), and (c) it mimics cyclical baseline patterns of behavior that may be observed in clinical practice. The $c$ represents a constant of 10 that we added to each value in our time series to avoid negative values. To simulate an effect of treatment in phase B only (i.e., treatment), we added the standardized mean difference (SMD), which is computed by subtracting the mean of all points in phase A ($\mu_A$) by the mean of all points in phase B ($\mu_B$) and by dividing the result by the standard deviation ($\sigma$) of the error term (i.e., 1).

**Effect Size Measures**

While at least nine nonoverlap measures exist, we focused on three nonoverlap measures: Tau-U for nonoverlap with baseline trend control (Tau-U; Parker, Vannest, & Davis, 2011), the percentage of data points exceeding the median (PEM; Ma 2006), and the robust improvement rate difference (R-IRD; Parker et al., 2009). We selected these measures as each is an improvement on other commonly used nonoverlap measures (i.e., percentage of nonoverlapping data, nonoverlap of all pairs, and improvement rate difference; see Ma, 2006; Parker et al. 2009; Parker & Vannest, 2009 for details), and each measure has been widely adopted by researchers. In addition, the calculation of each of these measures differs considerably from each other so as to make them representative of the available methods. We refer readers seeking more information on other measures to reviews by Parker et al. (2011) and Vannest and Ninci (2015).

**Tau-U.** In the current paper, Tau-U refers to the revised version described by Parker et al. (2011) that corrects for baseline trend. Tau-U compares baseline and treatment and adjusts for trend present in baseline (i.e., $\text{Tau-U}_{A \text{ vs. B- Trend A}}$). To correct for baseline trend, a
Kendall’s rank correlation is calculated between the A phase data and the session numbers and used to adjust Tau. The formula for calculating Tau-U is

\[ \text{Tau-U} = \frac{S_p - S_A}{mn} \]

(4)

where \( S_p = \sum_{i=1}^{m} \sum_{j=1}^{n} \left[ I(y_j^b > y_i^A) - I(y_j^b < y_i^A) \right] \) and \( S_A = \sum_{i=1}^{m-1} \sum_{j=i+1}^{n} \left[ I(y_j^A > y_i^A) - I(y_j^A < y_i^A) \right] \).

In previous and subsequent equations, the variables \( m \) and \( n \) represent the number of points in the baseline and treatment phases, respectively. The \( I \) represents the indicator function, which equates to 1 when the function is true and 0 when the function is false. The range of Tau-U index varies as a function of the values of \( m \) and \( n \) and can be expressed as

\[ -(2n + m - 1)/(2n) \text{ to } (2n + m - 1)/(2n). \]

Although often grouped with non-overlap measures, Tau-U technically belongs to a class of non-parametric rank correlation coefficients. That is, it measures the strength of the association between contrasted phases, rather than the amount of overlapping data points (Tarlow, 2017).

**Percentage Exceeding the Median.** If the anticipated direction of treatment is an increase, the PEM (Ma, 2006) is the percentage of data points in the B phase that exceed the median of the A phase. In the event phase B data points are equal to the median of the A phase they are counted as half a data point. To compute PEM, we used the original calculation method described by Ma (2006)

\[ PEM = \frac{1}{n} \sum_{i=1}^{n} \left[ I(y_i^b > y_A) \right] \]

(5)

where \( y_A \) is equal to the median of the A phase Percentage exceeding the median can range from 0 to 100, however to make the measure comparable to the scales of the other measures we converted it to a proportion, resulting in a range of 0 to 1. A value of .5 or less indicates that no treatment effect is present.

**Robust Improvement Rate Difference.** The R-IRD (Parker et al., 2009) is the improvement rate of the treatment phase minus the improvement rate of the baseline phase.
The improvement rate for each phase is calculated by determining the minimal number of overlapping data points that must be removed from either baseline or treatment to eliminate all overlap between the phases. For datasets where the anticipated outcome is an increase in treatment, the baseline improvement rate is the number of data points removed from the A phase divided by the length of the A phase. The treatment improvement rate is the number of data points left in the B phase after all overlap is removed divided by the length of the B phase. Pustejovsky (2017, 2018) provided a method for computing R-IRD from the percentage of all nonoverlapping data (PAND; Parker, Hagan-Burke, & Vannest, 2007). The exact algebraic relation of R-IRD to PAND is

\[
R\text{-IRD} = \frac{1}{2mn} \left( \frac{(m+n)^2 PAND}{100^2} - m^2 - n^2 \right)
\]

where \( PAND = 100 \% \left( \frac{x}{m+n} \right) \) and \( x = \max \{(i+j)I(y^{i+1}_{(m+n)} > y^j_{(m+n)})\} \) \( = = + \.ReLU \)

The R-IRD can range from 0 to 1 with a score of 1 indicating that there is no overlap between the phases. Parker et al. (2009) suggest that values less than .5 indicate “small” effect, values of .5 to .7 indicate a “medium” effect, and .7 to 1 indicate a “large” effect.

**Analysis 1: Establishing Threshold Values for Effect Size**

Our first analysis was conducted to determine if we could identify thresholds above which it would be unlikely for a practitioner to erroneously identify a behavior change.

We manipulated the number of data points in the baseline and treatment phases to produce six variations of phase length combinations (i.e., 5:5, 5:10, 5:15, 10:5 10:10, 10:15). For example, 5:10 represented a dataset with 5 baseline points and 10 treatment points and 15:10 a dataset with 15 baseline points and 10 treatment points. For each of six phase length variations, we generated a frequency distribution of each effect size measure (i.e., Tau-U, PEM, and R-IRD) in the absence of a true effect (i.e., SMD = 0), with or without autocorrelation (i.e., \( a = 0 \) or 0.2), and in the presence or absence of trend (i.e., \( \beta = 0 \) or 0.1).
We set the autocorrelation parameter at 0.2 because Shadish and Sullivan (2011) reported this value as the meta-analytic mean in their review of single-case research, whereas, we set the trend at 0.1, which was the median trend that we found when we reanalyzed 295 nonsimulated baseline datasets from a previous study (see Lanovaz et al., 2018). For each combination of parameters (72 in total), we generated a distribution of 100,000 datasets. We programmed our R script to select the effect size value below which at least 95% of all scores fell, which would generate threshold effect size values that produce a type I error rate of .05 or less.

**Analysis 2: Determining Power**

We repeated our first analysis with Tau-U and R-IRD, but we added the effect size parameter, SMD, to the points from phase B to simulate the effect of a treatment. We did not include PEM in our second analysis because we were unable to identify thresholds that adequately controlled for type I error rate for multiple sets of parameters (see results for the first analysis). The SMD value varied from 0.5 to 3.0 in 0.5 increments, which produced 288 combinations of parameters. Then, we examined the proportion of datasets in the distribution that produced effect size values equal to or higher than the threshold values for the same set of parameters as identified in the first analysis. Autocorrelation and trend vary widely in single-case designs, and the true value may remain unknown because datasets contain few data points. To remain conservative, practitioners may choose to use the highest threshold values (typically those established for datasets containing autocorrelation and trend) for all their datasets. We therefore also repeated our analyses using the most conservative threshold value (i.e., the highest value) for each of the six phase length variations.

**Results**

Table 1 shows the threshold values obtained for Tau-U, PEM, and R-IRD in our first analysis. The results of our analysis indicate that it is possible to set thresholds for effect size
above which it is unlikely that the observed change is due to chance. Across all phase length variations, the established threshold values were the lowest when no autocorrelation or trend was present in the data. The highest threshold values were nearly always associated with datasets containing both autocorrelation and trend. For example, when datasets contained 5 data points in baseline and 10 in treatment with no autocorrelation or trend (i.e., $a = 0, \beta = 0$), the obtained threshold values for Tau-U and R-IRD were 0.60 and 0.70, respectively. With the addition of autocorrelation and trend (i.e., $a = 0.2, \beta = 0.1$), those threshold values increased to 0.80 for Tau-U and 0.85 for R-IRD. For PEM, threshold values that produced a type I error rate of .05 or less did not exist for multiple sets of parameters.

Table 2 presents the power of Tau-U and R-IRD to detect a SMD of 2.5 for the exact and most conservative threshold values, which we obtained as part of our second analysis. We chose to present the data for a SMD of 2.5 as researchers have shown that SMDs for single-case data are typically 3.0 or higher (e.g., Gierut, Morissette, & Dickinson, 2015; Lanovaz et al., 2018; Rogers & Graham, 2008). However, the power at a SMD of 3.0 often reached the ceiling of 1, which masked the contribution of the individual parameters; we thus present the values at 2.5 to facilitate comparisons. For effect sizes of 2.5, power was typically near or above 0.8 except when both phases contained only five points. Whether Tau-U or R-IRD was more powerful mainly depended on the number of points in each phase, and expectedly, the conservative values were less powerful than the exact values. Both effect size measures performed best when the number of points increased in either phase. For example, applying the most conservative threshold values to datasets containing 5 baseline and 10 treatment data points the power of Tau-U to detect a SMD of 2.5 was .87 (autocorrelation and trend absent). However, when the number of data points in each phase was increased to 20, there was a corresponding increase in the power (i.e., .99) to detect an effect.
Figure 1 summarizes the power of Tau-U and R-IRD across SMDs for 5 points in phase A and 10 points in phase B. We selected these phase lengths for presentation because practitioners typically want to minimize the number of baseline sessions while conducting sufficient treatment sessions to ensure repeated exposure to treatment. At these phase lengths, Tau-U was more powerful than R-IRD and typically had sufficient power to detect SMDs of 2.0 or more. To allow the reader to consider the effects of different sets of parameters, we provided the tables for the other values of SMD and the figures for the other phase length variations as supplementary material\(^2\).

**Discussion**

Overall, our analyses show that Tau-U and R-IRD may provide adequate control over type I error rate and sufficient power when detecting changes in quasi-experimental AB designs. In addition to independently validating the properties of these measures with simulated data, our results show that the Tau-U and R-IRD can support clinical decision making. In contrast, PEM seems inadequate as the measure is prone to ceiling effects, which inflates the type I error when the threshold is at 1.0. Taken together with the results obtained by Lanovaz et al. (2018), our study suggests that practitioners may adopt AB quasi-experimental design with Tau-U or R-IRD to assess individual progress while minimizing interpretation errors.

For example, a practitioner who decides to collect at least five data points during baseline and at least ten data points during treatment may use 0.80 and 0.85 as thresholds for Tau-U and R-IRD, respectively (see Table 1). Observing an effect size equal to or higher than these values should produce a false positive interpretation in less than 1 in 20 cases, which is on par with the type I error rate threshold used in experimental designs. If the effect size is below the threshold, the practitioner may rely on visual analysis to provide further support. If

\(^2\) See [https://osf.io/9br7g/?view_only=8ecdb1554b64e2388c62d792153995b](https://osf.io/9br7g/?view_only=8ecdb1554b64e2388c62d792153995b)
the visual analysis shows differentiation between phase A and phase B, the practitioner may conduct a within-subject replication using an experimental design to confirm the results (if possible to withdraw the treatment). Otherwise (i.e., visual analysis does not show differentiation), the practitioner may conclude that there was no behavior change and assess the effects of an alternative treatment. Thus, only a small subset of individuals would require an experimental design, considerably reducing the costs and resources necessary to assess and monitor progress.

Practitioners should take into consideration the limitations of our effect size measures. First, nonoverlap measures may produce ceiling effects. When no overlap is present, the measure will indicate a large effect size (i.e., 1.0). However, this effect size only indicates the absence of overlap and does not provide information on the magnitude of the change between baseline and treatment (Ma, 2006). Second, the version of Tau-U used in this study is affected by the ratio of baseline to treatment data points. As such, the effect size may be artificially inflated by collecting additional treatment data points until Tau-U is statistically significant (Brossart, Laird, & Armstrong, 2018; Tarlow, 2017). If a practitioner finds a baseline trend is reducing their effect size, a within-subject replication may be warranted to confirm the effect.

Although our results are consistent with those obtained with nonsimulated datasets (Lanovaz et al., 2018) and we relied on prior research to set the values of our parameters, our simulations may not have perfectly mimicked patterns observed in practice, which is a limitation. Future research should apply the current thresholds to nonsimulated data to examine to what extent the results match those of other types of analyses (e.g., visual analyses, randomization tests). An additional limitation is that we only used nonoverlap effect size measures as part of our analyses. We could not evaluate mean-based magnitude measures such as the log response ratio (Pustejovsky, 2018) because SMD directly
manipulated the mean. Therefore, inclusion of mean-based effect size measures would have produced circular outcomes.

Finally, the primary concern against the use of quasi-experimental AB designs is their inability to demonstrate functional relations (or experimental control). An experimenter has demonstrated a functional relation when an observed change was shown to be the product of an independent variable (e.g., treatment) while ruling out extraneous variables. The AB design does not rule out the effects of maturation or history (Christ, 2007), which prevents the identification of the variable responsible for an observed change. However, the AB design can determine whether the behavior (or target) has changed significantly from phase A to phase B, which is arguably more important for practitioners than identifying the exact mechanism responsible for this change. In sum, our results suggest that practitioners may use quasi-experimental AB designs in combination with Tau-U or R-IRD to assess and monitor progress in practice.
References


### Table 1

**Summary of Threshold Values for Tau-U, PEM, and R-IRD for Each Phase Length Variation**

<table>
<thead>
<tr>
<th>Data Characteristics</th>
<th>5:5</th>
<th>5:10</th>
<th>5:15</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Tau-U</td>
<td>PEM</td>
<td>R-IRD</td>
</tr>
<tr>
<td>$a = 0, \beta = 0$</td>
<td>0.76</td>
<td>NA</td>
<td>0.80</td>
</tr>
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<tr>
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<td>1.00</td>
</tr>
<tr>
<td>$a = 0.2, \beta = 0.1$</td>
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<td>NA</td>
<td>1.00</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Data Characteristics</th>
<th>10:5</th>
<th>10:10</th>
<th>10:15</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Tau-U</td>
<td>PEM</td>
<td>R-IRD</td>
</tr>
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<td>0.85</td>
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<td>0.78</td>
<td>1.00</td>
<td>0.70</td>
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</table>

**Note.** PEM: percentage of data points exceeding the median, R-IRD: robust improvement rate difference, $a$: autocorrelation, $\beta$: trend parameter. NA indicates it was not possible to identify a threshold value that produced a type I error rate of .05 or less.
Table 2

Summary of Power for Tau-U and R-IRD for Each Phase Length Variation for a Standardized Mean Difference (SMD) of 2.5

<table>
<thead>
<tr>
<th>Data Characteristics</th>
<th>Tau-U</th>
<th>R-IRD</th>
<th>Tau-U</th>
<th>R-IRD</th>
<th>Tau-U</th>
<th>R-IRD</th>
</tr>
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<tbody>
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<td>5:5</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>a = 0, β = 0</td>
<td>0.85</td>
<td>0.93</td>
<td>0.98</td>
<td>0.97</td>
<td>0.99</td>
<td>0.94</td>
</tr>
<tr>
<td>a = 0, β = 0.1</td>
<td>0.89</td>
<td>0.92</td>
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<td>0.88</td>
<td>0.99</td>
<td>0.87</td>
</tr>
<tr>
<td>a = 0.2, β = 0</td>
<td>0.73</td>
<td>0.61</td>
<td>0.94</td>
<td>0.81</td>
<td>0.97</td>
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<tr>
<td>a = 0.2, β = 0.1</td>
<td>0.79</td>
<td>0.60</td>
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<td>0.85</td>
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<tr>
<td>5:10</td>
<td></td>
<td></td>
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<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>a = 0, β = 0</td>
<td>0.84</td>
<td>0.97</td>
<td>1.00</td>
<td>1.00</td>
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<td>1.00</td>
<td>0.98</td>
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<td></td>
</tr>
<tr>
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<td>0.87</td>
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<td>0.88</td>
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</tr>
<tr>
<td>a = 0, β = 0.1</td>
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<td>0.57</td>
<td>0.94</td>
<td>0.88</td>
<td>0.97</td>
<td>0.49</td>
</tr>
<tr>
<td>a = 0.2, β = 0</td>
<td>0.72</td>
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<tr>
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<td>0.60</td>
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Note. R-IRD: robust improvement rate difference, a: autocorrelation, β: trend parameter.
**Figure 1.** Power for the exact and most conservative threshold values for Tau-U and robust improvement rate difference (R-IRD) for 5 points in phase A and 10 points in phase B. $a$ represents the autocorrelation and $\beta$ the trend parameter.