

Université de Montréal

Lipoprotein-Associated Phospholipase A₂ (Lp-PLA₂) in Acute Coronary Syndrome

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Abstract

Lipoprotein associated phospholipase A₂ (Lp-PLA₂) is a biomarker of several inflammatory diseases and syndromes. An elevated Lp-PLA₂ level is associated with unstable atherosclerotic plaques. Bound to plasma lipoproteins (LDL and HDL), Lp-PLA₂ prevents the formation of biologically active oxidized phospholipids on their surface such as oxidized phosphatidylcholine (oxPC). Nevertheless, the products of Lp-PLA₂ action, lysophosphatidylcholine (LPC) and non-esterified fatty acids (NEFA) are both known to aggravate inflammation.

Thus, understanding the metabolism of Lp-PLA₂ could help us better understand its role in plaque formation, as studies have shown high expression of Lp-PLA₂ and LPCs in unstable plaques. Moreover, studies showed correlation between increased Lp-PLA₂ mass and activity and increased risk of coronary artery disease, stroke, and death. The inhibition of Lp-PLA₂ with a small molecule, Darapladib, has not demonstrated benefit in reduction of cardiovascular events in two clinical studies.

Here, the first chapter will focus on Lp-PLA₂ and cardiovascular disease in man, highlighting the latest updates in the literature. The second and third chapters will introduce experimental work on Lp-PLA₂ in the setting of acute coronary syndrome.

Keywords : Lp-PLA₂, Lipoprotein-associated phospholipase A₂, PAF-AH, platelet activating factor acetyl hydrolase, acute coronary syndrome, ACS, cardiovascular diseases, HDL, LDL

Résumé

La phospholipase A₂ liée aux lipoprotéines (Lp-PLA₂) est un biomarqueur de plusieurs maladies inflammatoires et un niveau sérique élevé est associé à l'instabilité de la plaque artérioscléreuse. Comme son nom l'indique, la Lp-PLA₂ est liée aux lipoprotéines plasmatiques (LDL et HDL) et son rôle est de prévenir l'accumulation de phospholipides oxydés à la surface des lipoprotéines. Toutefois, les produits de dégradation des phospholipides oxydés par la Lp-PLA₂ - le lysophosphatidyl choline par les acides gras oxydés peuvent aussi promouvoir l'inflammation.

Mieux comprendre le métabolisme de la Lp-PLA₂ pourrait nous permettre de mieux apprécier son rôle dans la formation d'une plaque artérioscléreuse instable, car des études antérieures ont démontré une forte expression de la Lp-PLA₂ dans la plaque. De plus, il existe une forte corrélation entre les niveaux et l'activité plasmatiques de la Lp-PLA₂ et la maladie coronarienne, les accidents cérébraux-vasculaires et la mortalité cardiaque. L'inhibition de la Lp-PLA₂ avec une petite molécule, le darapladib, n'a pas démontré de bénéfice sur les événements cardiovasculaires dans deux études cliniques.

Cette thèse présentera d'abord une revue de la littérature sur la Lp-PLA₂ et les maladies cardiovasculaires et les deuxième et troisième chapitres, une étude clinique réalisée sur des patients avec un syndrome coronarien aigu.

Mots-clés : Phospholipase A₂ liée aux lipoprotéines (Lp-PLA₂); Facteur d'activation des plaquettes (PAF); acyl hydrolase (AH); syndrome coronarien aigu (ACS); lipoprotéines de haute densité (HDL); lipoprotéines de basse densité (LDL).

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List of abbreviations

ACS: acute coronary syndrome

Apo A1: apolipoprotein A1

Apo B: apolipoprotein B

CAD: stable coronary artery diseases

CVD: cardiovascular diseases

HDL: high density lipoprotein

HDL-C: high-density lipoprotein cholesterol

HPLC: high-performance liquid chromatography

hsCRP: high-sensitivity C-reactive protein

LDL: low density lipoprotein

LDL-C: low-density lipoprotein cholesterol

LPA: lysophosphatidic acid

LpB: apoB-containing lipoprotein fraction

LPC: lysophosphatidylcholine

Lp-PLA₂: Lipoprotein-associated phospholipase A₂

NEFA = non-esterified fatty acids

oxPC = oxidized phosphatidylcholine

PAF = platelet activating factor (1-O-alkyl-2-acetyl-sn-glycero-3-phosphocholine)

PAF-AH = platelet activating factor acetyl hydrolase

PLA₂ = Phospholipase A₂

UTC: ultracentrifugation

Chapter I: Introduction

Lipoprotein-Associated Phospholipase A₂ (Lp-PLA₂): a brief
introduction

1.1 Introduction

Cardiovascular diseases are the leading cause of death worldwide. Over decades, studies aimed to identify and evaluate cardiovascular risk in individuals. The Framingham Heart Study is probably the frontrunner in this field. Launched in 1948, Framingham's first cohort included more than 5,000 participants, and followed in recruiting around 9,000 subjects from their offspring in a second and a third cohorts (1971 and 2002) for a total of more than 14,000 participants. Today, our clinical guidelines include "traditional" modifiable risk factors for cardiovascular diseases: smoking, high cholesterol levels, diabetes, hypertension, obesity, low physical activity and unhealthy diet. Targeting those risk factors by clinicians has resulted in a 50% reduction in age adjusted mortality rates in the US over a 50-year period. In Canada, we see a glimpse of progress as cardiovascular diseases (for the first time) ranked second instead of first as a leading cause of death in 2008. This has largely been confirmed in many observational studies, especially the INTERHEART study.¹⁻⁴ However, an observational study by the National Registry of Myocardial Infarction (NRFMI) showed that out of a total of 542,008 patients presented with a first acute myocardial infarction between 1994 and 2006, an overall 78,103 (14.4%) had none of the risk factors mentioned above, hence the need for additional biomarkers of cardiovascular risk.⁵

Coronary artery disease (CAD) is overwhelmingly caused by atherosclerosis, a process that involves long-term chronic inflammation of the arteries. Thus, the relationship between inflammation, atherosclerosis and the high incidence of cardiovascular diseases (CVD) has been extensively studied. The inflammatory process is triggered by a combination of lipid accumulation in the arterial wall that might undergo oxidation over time (oxLDL), and

activation of several types of inflammatory cells (macrophages, T-cells, mast cells). That results in the formation of arterial plaques that may or may not be stable. Foam cells are lipid-laden macrophages that undergo apoptosis and form the necrotic core of the arterial plaque. In susceptible plaques, the production of certain cytokines e.g. interferon-g (IFN-g) diminishes collagen synthesis and results in plaques with thin fibrous caps. Moreover, the production of proteases by inflammatory cells can disrupt the plaque's fibrous cap leading to plaque rupture. A series of events characterized by platelet activation and thrombus formation follow, and may lead to arterial occlusion and a subsequent event of acute coronary syndrome (ACS) (such as myocardial infarction). On the other hand, stable plaques (less prone to rupture) continue to grow and cause reduction in the arterial blood flow, and can manifest as stable ischemic heart disease.⁶⁻⁸

With the need for new biomarkers for CVD and atherosclerosis, research has yielded new potential biomarkers such as high-sensitivity C-reactive protein (hsCRP), Brain natriuretic peptide (BNP), anti-phosphorylcholine (Anti-PC), and Lp-PLA₂.⁹

Lp-PLA₂, also known as platelet activating factor acetyl hydrolase (PAF-AH), is a member of the phospholipase A₂ (PLA₂) family, group VIIA. According to the biochemical structure and properties of its members, Phospholipase A₂ family is classified into 5 categories (secreted PLA₂, cytosolic PLA₂, Ca⁺ independent PLA₂, lysosomal PLA₂, and PAF-AH PLA₂) divided in 15 groups. PLA₂ enzymes hydrolyze the ester bond at the sn-2 position of phospholipids producing free fatty acids (FFA) and lysophospholipids.¹⁰

In the arterial wall, several phospholipids on LDL undergo oxidation altering their structure to yield oxLDL, which can be taken up by macrophages. OxLDL molecules contain different bioactive compounds that may include phospholipid products, sphingolipid products, free fatty

acid products, oxysterols, cholesteryl ester products, hydroxynonenal and malondialdehyde, and products of apolipoprotein B (ApoB) modification.¹¹

During the oxidation process, reactive oxygen species attack fatty acyl chains and cause them to undergo peroxidation to generate peroxides that provoke different signalling pathways that end up aggravating the inflammation process.¹² Unsaturated chains are more prone to oxidation than saturated fatty acids. Moreover, short-medium chained fatty acids are more prone to oxidation than long chained fatty acids.^{13, 14} Therefore, polyunsaturated fatty acids (PUFA) commonly located on the sn2 position of the fatty acyl chain of the phospholipid are easier targets for oxidation due to the weak hydrogen-carbon bonds in methylene groups, and are believed to be the main source of various oxidized phospholipids.^{12, 15, 16}

While PLA1 enzymes cleave fatty acyl chains at the sn1 position, oxidized phospholipids at the sn2 position trigger Lp-PLA₂, yielding two main products: lysophosphatidylcholine (LPC) and nonesterified fatty acids (NEFA).¹⁴

1.2 Biochemistry of Lp-PLA₂

1.2.1 Structure of Lp-PLA₂

Lp-PLA₂ is a 45 KDa protein that consists of 441 amino acids and doesn't require Ca⁺ to perform its function. The crystal structure of Lp-PLA₂ has been identified.¹⁷ The catalytic site consists of a triad of serine, aspartate, and histidine, located on residues 273, 296, and 351 respectively, characterized Lp-PLA₂ in a class separate from other PLA₂ enzymes that lack the serine.¹⁰

Various cells secrete Lp-PLA₂ such as macrophages, Kupffer cells, platelets, and mast cells, although it is likely that macrophages have the biggest role. Once Lp-PLA₂ is secreted into the plasma, it binds to lipoproteins (mainly HDL and LDL) and circulates as an active enzyme. Some studies reported Lp-PLA₂ distribution as approximately 80% on LDL and 20% on HDL, while others reported that it is around two-thirds on LDL vs. one third on HDL, and that Lp-PLA₂ mass in the plasma is more dependent on LDL-C levels.^{18, 19} The interaction between Lp-PLA₂ and LDL occurs between the N-terminus part of Lp-PLA₂ and the C-terminus part of ApoB.¹⁶ A previous study has shown that residue 205 on Lp-PLA₂ is involved in that binding. On the other hand, it seems that Lp-PLA₂ binds to HDL at three different sites, residues 113–120, 192–204, 360–368, with 192–204 being the region where Lp-PLA₂ binds to apolipoprotein A1 (ApoA1).²⁰

Previous work has shown that Lp-PLA₂ is associated with Lipoprotein a (Lp(a)) in the plasma,^{21, 22} while other studies have demonstrated the presence of a higher tendency of binding between Lp-PLA₂ and small dense electronegative LDL subfraction (LDL-5) and very high-density lipoprotein-1 subfraction (VHDL-1) in human plasma.^{16, 23} A genome-wide association study by *Thanassoulis et al* recently discovered an association between Lp(a)

levels and aortic valve calcification and stenosis, suggesting a causal role for Lp(a) in valvular calcification. However, the relationship between Lp-PLA₂ and Lp (a) is interesting.²⁴

1.2.2 Lp-PLA₂ and PAF

When it was first discovered, and before joining the PLA₂ family, Lp-PLA₂ was initially known as Platelet activating factor acetylhydrolase (PAF-AH). PAF-AH was shown to hydrolyze the ester bond at the sn-2 position of PAF, producing LysoPAF and acetate. PAF-AH was known for its potency in deactivating PAF. Today, it is classified as group VIIA, and is more commonly known as lipoprotein-associated phospholipase A₂ (Lp-PLA₂) due to its larger substrate specificity including PAF and oxLDL.¹⁷

First described in 1972, platelet-activating factor (PAF) is a phospholipid that is produced by various cell types (platelets, neutrophils, endothelial cells, and others) and acts as a communication mediator that transmits signals between cells. Similar to Lp-PLA₂, the name PAF is somewhat misleading because it does not fully describe PAF biological activity. While it activates platelets, PAF has several physiological and pathological effects; it has shown to cause uterine contractions, bronchoconstriction, affect the vascular system (lowering blood pressure, reducing cardiac output, increasing vascular permeability), with a possible effect on the endocrine, gastrointestinal, and other systems.^{25, 26} PAF has been linked to the inflammatory process as a proinflammatory molecule that binds to the PAF membrane receptor expressed on various cells, activates multiple signalling pathways (platelets, neutrophils, macrophages) that could trigger an inflammatory response that may include platelet aggregation, apoptosis, and vascular remodelling.

Later, scientists discovered other phospholipids that have a resembling structure to PAF and function in a similar pattern as signalling molecules. The biologic effects of “PAF-like” molecules include platelet activation, PPAR activation, and stimulation of mitosis. Normal cells deactivate PAF and PAF-like molecules by intracellular PAF-AH, while extracellular PAF-AH (Lp-PLA₂) also deactivates oxLDL in the arterial wall.^{11, 27}

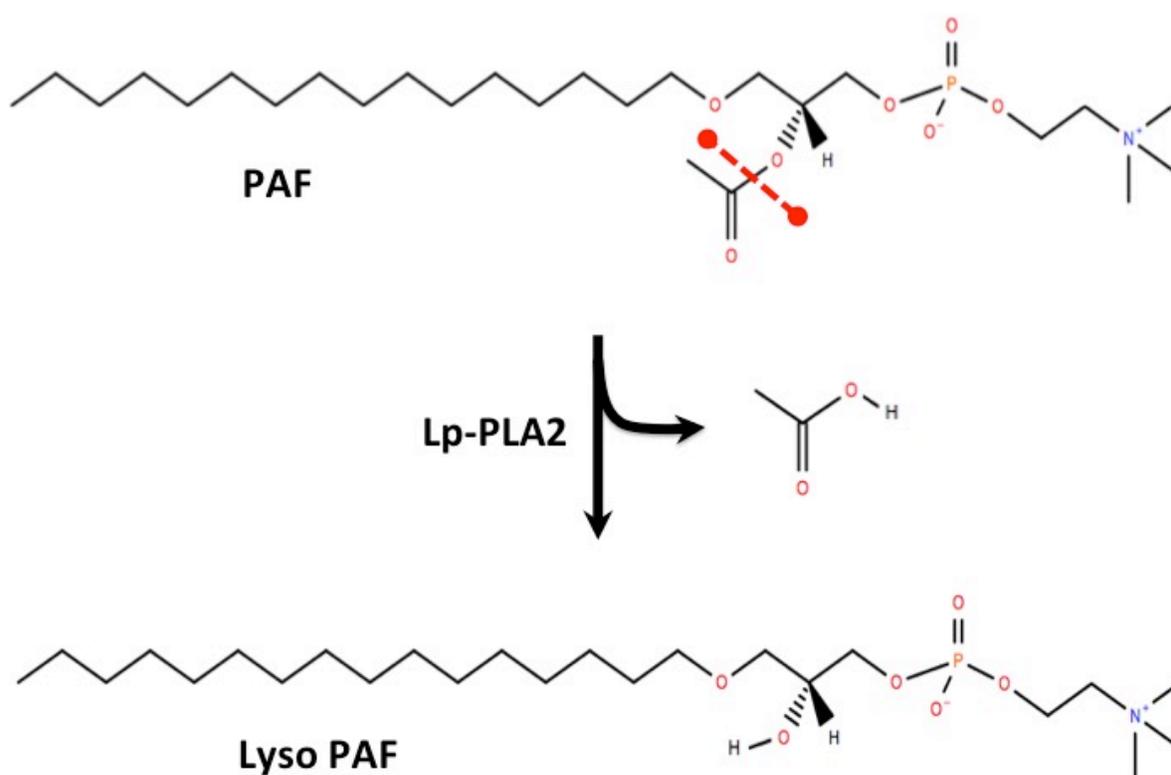


Figure 1. Hydrolysis of PAF by Lp-PLA₂ (PAF-AH). Chemical structure for PAF was obtained from Lipid maps.²⁸

1.2.3 Substrate specificity

When it comes to substrates, it seems that Lp-PLA₂ has broad specificity. An exact substrate for Lp-PLA₂ is not yet well identified despite several studies. Although PAF-AH, by definition, has higher affinity towards PAF, it is now known that Lp-PLA₂ /PAF-AH substrates are not limited to PAF, but also include PAF-like molecules which are also phospholipids such as oxidized phospholipids, esterified isoprostanes, PEIPC and PC-hydroperoxides.¹⁵

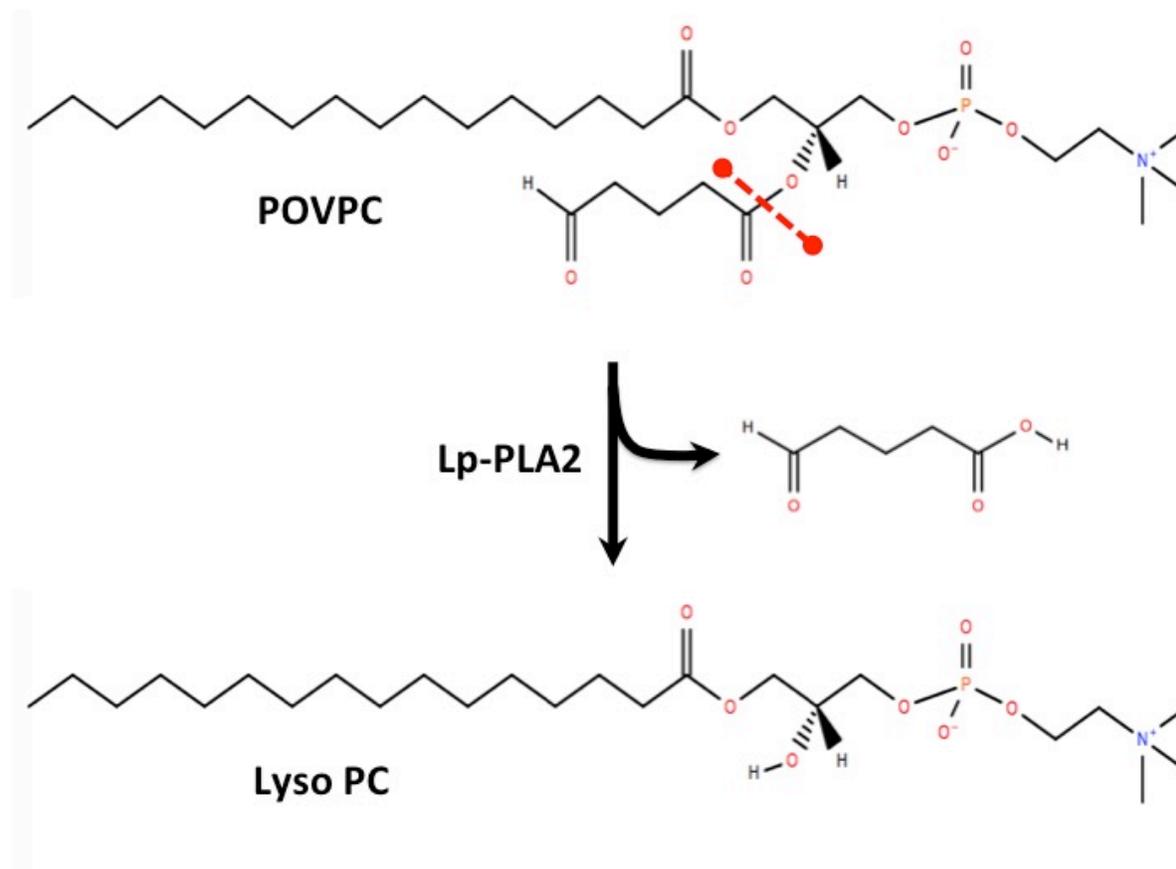


Figure 2. Hydrolysis of an oxidized phospholipid (POVPC) by Lp-PLA₂. Chemical structure for POVPC was obtained from Lipid maps.²⁸

Min et al reported that diglycerides, triglycerides and acetylated alcohols are also potential substrates for Lp-PLA₂. With an extensive possible structure of an oxLDL molecule, Lp-PLA₂ recognizes substrates that have a structure of a glyceride derivative with a hydrophobic chain at the sn-1 position and an ester at the sn-2 position.²⁹ It has also been suggested that Lp-PLA₂ residues 189-239 are responsible for substrate specificity.²¹

Despite the large number of potential substrates that could be targeted by Lp-PLA₂, it seems that it targets certain phospholipids. Previous work has identified several truncated PC oxidation products as main substrates of Lp-PLA₂.¹⁴

The specificity of the enzyme's reaction can be linked to length of the fatty acyl on the sn2 position of the phospholipid. Action of Lp-PLA₂ seems to be more specific to fatty acyl chains of up to nine methylene groups. Phospholipids with longer chains (containing more than nine methylene groups) seem to be exempted from the action of PAF-AH, unless they are oxidized.

14, 30

1.2.4 Products of Lp-PLA₂ and their biologic effects

In order to neutralize PAF, Lp-PLA₂ cleaves at the sn2 position of PAF to produce lysoPAF, a biologically inactive product, and acetate.

A main product of Lp-PLA₂ action on oxidized phospholipids is lysophosphatidylcholine (LPC), in the form of saturated and monounsaturated LPCs. LPC is produced in the plasma by 2 other pathways: LCAT, and another phospholipase A₂: sPLA₂ which is classified as group IIa. With the production of LPC, a series of proinflammatory consequences escalate the inflammatory process and involves different cells. Those can be described by attracting

monocytes, stimulating arachidonic acid release, upregulating cytokines and adhesion molecules, and induction of apoptosis.³¹

With the additional effect of certain enzymes that target LPC, lysophosphatidic acid (LPA) is produced, consequently leading to platelet activation.¹¹ LPC also acts as an intermediate product that could lead to the reproduction of PAF through the enzyme lysophosphatidylcholine acyltransferase (LPCAT), amplifying the inflammatory process.²⁷ The other product of Lp-PLA₂ action on oxidized phospholipids is free fatty acids (FFA). Those are also have been shown to be proinflammatory as they act as monocyte chemoattractants.³²

1.2.5 Regulation of Lp-PLA₂

The *PLA2G7* gene located on chromosome 6p21.1-p12 encodes Lp-PLA₂. *PLA2G7* is expressed in different tissues in the following decreasing order: thyroid, leukocytes, prostate, ovaries, lung, and others.^{33, 34} Lp-PLA₂ has been associated with the pathophysiology of several diseases and syndromes such asthma, atopy, dementia, coronary artery disease (CAD), heart failure, and stroke.²¹

To date, we do not have a complete understanding of Lp-PLA₂ regulation. Taking in consideration its broad substrate specificity and controversial function, it seems that the regulation of Lp-PLA₂ is multifactorial. Several studies concluded that Lp-PLA₂ expression is dependent on cell differentiation and inflammatory mediators.^{35, 36} A recent study demonstrated that serum amyloid A (SAA) significantly increases Lp-PLA₂ expression in THP-1 cells and ApoE knock out mice.³⁷ Lp-PLA₂ activity in the plasma can also be estrogen regulated.³⁸

As the main producers of Lp-PLA₂, studies targeted Lp-PLA₂ expression by macrophages in atherosclerotic plaques as well as the plasma. At the transcriptional level, *Ferguson et al* showed that Lp-PLA₂ mRNA levels were elevated in macrophages (especially M1 macrophages) but not in their precursors, the monocytes, and that expression increases post differentiation. Moreover, the study found Lp-PLA₂ mRNA and protein levels to be higher in foam cells as compared to macrophages³⁶. Another study looked at the regulation of Lp-PLA₂ in macrophages and found that interferon gamma (IFN- γ) - a proinflammatory molecule - and lipopolysaccharide repressed Lp-PLA₂ synthesis³⁹.

Previous studies have shown a higher expression of Lp-PLA₂ in aortic plaques⁴⁰. A recent study by *Mahmut et al* has shown a several fold increase in mRNA expression for Lp-PLA₂ in patients with calcific aortic valve disease (CAVD) as compared to subjects with non-calcified aortic valves. The same study reported a higher plasma Lp-PLA₂ activity in CAVD patients as compared to controls, and a higher Lp-PLA₂ activity in stenotic valves than in non-mineralized valves⁴¹. We do not know much about its regulation in the plasma, although oxidized phospholipids have been shown to play a role in its regulation through several pathways.

OxLDL appears to be a main player in the regulation of Lp-PLA₂ expression. A study by *Wang et al* suggested that Lp-PLA₂ upregulation was due to the presence of its substrates (oxidized PC species), but not its products. In the same study, the authors noticed that PAF didn't exert a similar effect on Lp-PLA₂ expression³⁵, while other studies showed that PAF.

Finally, it is worthwhile mentioning that several studies have shown that despite Lp-PLA₂ being regulated by several pathways, it does not act as an acute phase reactant.^{19, 36, 42}

1.3 Lp-PLA₂ animal models

Because atherosclerosis is a complex process, choosing the suitable animal model depends on the specific aspect under study.⁴³ The mouse is probably not the best model for Lp-PLA₂ and atherosclerosis studies, because their lipid profile is different from humans, and as a result the distribution of Lp-PLA₂ is also different. Despite that fact, Lp-PLA₂ and a specific Lp-PLA₂ inhibitor, Darapladib were studied in LDLR-deficient mice and homozygous ApoE-deficient mice. Both studies showed that Lp-PLA₂ inhibition by Darapladib reduces inflammation and is associated with a decrease in plaque formation.^{44, 45}

A study by *Wilensky et al* preferred a pig model to study Lp-PLA₂⁴⁶, where the authors mention that the reasons for choosing the pig model rather than the mouse model were the similarity of plasma lipoprotein profile in pigs to humans, the similarity of Lp-PLA₂ distribution in pigs to humans, the better ability to study coronary lesions in those models compared to mice. In addition to those points, pig models had other advantages: anatomical resemblance of their cardiovascular system to humans, the resemblance of their atherosclerotic lesions to human lesions, their bigger size (bigger coronaries, bigger plaques), less need for genetic manipulation, and they can have atherosclerotic lesions even with chow diet (although slower) without the need for induction with high fat diet. In preclinical studies, the use of large animals (pigs, dogs, monkeys) is preferred for testing the efficacy of new agents such as Darapladib. The pig is often selected because of its similarities in terms of metabolism and cardiovascular system to that of man. Thus, the pig model appears a more suitable model for atherosclerotic studies compared to mice or rabbits.^{43, 47}

1.4 Lp-PLA₂ and statins

Statins have shown to reduce both the risk of cardiovascular events (by around 30%) and mortality. Those agents appear to play a role in limiting the atherosclerotic process by decreasing the levels of circulating LDL-cholesterol in the plasma, and possibly through other mechanisms, although not yet confirmed. Several studies show that Lp-PLA₂ levels are significantly decreased after taking lipid-lowering medications.^{22, 48} It is noticed that patients who were on high doses of statins (HMG-CoA reductase inhibitors) had reduced Lp-PLA₂ mass and activity of about 20% to 30%, with Atorvastatin being more effective than other statins.^{19, 49, 50} In one of the mentioned studies, Atorvastatin did not affect Lp-PLA₂ activity on HDL-C which confirms that statins do not influence Lp-PLA₂ production, but rather increased LDL-C clearance.⁵¹ Statins lower Lp-PLA₂ levels. The role of the specific Lp-PLA₂ inhibitor, Darapladib, has been questioned in clinical trials.

1.5 Controversy of Lp-PLA₂ function

The role that Lp-PLA₂ plays in CVD is still not very clear. To date, two main opposite roles for the enzyme in atherosclerosis have been suggested: anti-inflammatory and proinflammatory. One view holds that the substrates for Lp-PLA₂ action, such as PAF and oxPC are known to be proinflammatory molecules, thereby emphasizing the antiatherogenic properties of Lp-PLA₂. However, the by-products of Lp-PLA₂ action are also proinflammatory molecules.⁵² Several studies have shown high expression of Lp-PLA₂ and LPC in unstable plaques. Moreover, they have shown correlation between increased Lp-PLA₂ mass and activity and increased risk of CAD, stroke, and death.^{40, 53}

Overexpression of Lp-PLA₂ and its products in atherosclerotic plaques could be to the fact that *PLA2G7* is upregulated by several inflammatory mediators that are abundant in an inflammatory milieu. Moreover, in vivo studies have shown reduced inflammation with recombinant Lp-PLA₂, which supports an anti-inflammatory role ²¹. A study conducted by Heart Protection Study Collaborative Group concluded that lipoproteins have high influence on the association between Lp-PLA₂ and risk of coronary events. With adjustment for apoB levels, the association between the two became trivial ⁵⁴.

While elevated Lp-PLA₂ plasma levels are associated with increased cardiovascular risk, HDL-associated Lp-PLA₂ is associated with a lower cardiovascular risk, as observed in 524 stable CAD patients in 3 year follow up study. That supports a more recent interpretation of those observations suggesting that Lp-PLA₂ on HDL might participate in the antioxidant function of HDL and has an overall anti-inflammatory role, while Lp-PLA₂ on LDL has a proinflammatory function ⁵².

Taking different points in consideration, it is not clear whether the inhibition of Lp-PLA₂ is beneficial in decreasing cardiac risk and mortality, as causality cannot be justified according to those associations. To reconcile this controversy, it is conceivable that the actions of Lp-PLA₂ maybe beneficial in one environment, but deleterious in another. Thus, we can postulate that the effects of Lp-PLA₂ maybe beneficial in specific tissues e.g. plasma, but maybe deleterious in the plaque, where the molecules resulting from its action may then trigger secondary messengers, and further aggravate inflammation.

1.6 Epidemiologic studies of Lp-PLA₂

During the past decades, studies repeatedly explored the association between Lp-PLA₂ and risk of CVD. In 2010, the Lp-PLA₂ collaboration studies, a meta-analysis, evaluated the records of 79,036 subjects of 32 prospective studies and showed positive correlation between Lp-PLA₂ and the risk of CAD, stroke, and death from vascular and nonvascular causes.⁵⁵

A more recent study by *Gonçalves et al* demonstrated significant correlations between Lp-PLA₂, LPC species and LPA with several inflammatory mediators (IL-1 β , IL-6, MCP-1, macrophage inflammatory protein-1 β) and anti-inflammatory mediator (IL-10) in the atherosclerotic plaques in humans.³¹ A previous study showed a higher carotid plaque expression of Lp-PLA₂ and a higher concentration on LPC in symptomatic patients as compared to non-symptomatic patients undergoing carotid endarterectomy.⁴⁰ *Vicker's et al* showed that Lp-PLA₂ and oxLDL were highly correlated and expressed in areas with greater potential of plaque rupture artery (bifurcation and internal segments of the carotid artery).⁵⁶

Two recent studies investigated Lp-PLA₂ in the ACS setting and noticed strong positive correlations between Lp-PLA₂ and LDL-C, ApoB with no association with hs-CRP and troponins. Meanwhile, studies have shown that the correlation between Lp-PLA₂ and HDL-C was either weak or inverse.^{19, 40, 57}

In the setting of familial hypercholesterolemia (FH), a recent study by *Tellis et al* that included 53 patients and detected significant elevation in Lp-PLA₂ mass and activity.²² In addition, few case studies reported high plasma Lp-PLA₂ activity in patients with Tangier's Disease (TD), which is manifested by low levels of circulating HDL and defective cholesterol efflux caused by mutations in the *ABCA1* gene, leading to higher risk of CVD⁵⁸. In a study that included almost 500 subjects, increased Lp-PLA₂ levels were reported in heart failure and aortic

aneurysm patients.⁵⁹ In 48 patients with severe aortic stenosis without overt atherosclerosis, inverse correlation between Lp-PLA₂ and aortic valve areas was reported, in addition to significant elevations in Lp-PLA₂ levels.⁶⁰

The controversy of Lp-PLA₂ function extends to genetic studies. A relatively common *PLA2G7* mutation of (V279), has been identified in several populations, and has shown to be more prevalent in East Asian populations (especially in Japan) than other Asian or European countries such as Turkey. This mutation results in partial or complete loss of function of Lp-PLA₂. Studies evaluating this mutation reported conflicting results, with more studies emphasizing a proinflammatory role for Lp-PLA₂ in the carriers of this mutation.⁶¹⁻⁶³

Finally, Brilakis reported that race and sex influence Lp-PLA₂ levels, suggesting taking those factors in consideration when measuring Lp-PLA₂ levels.⁶⁴ It also seems that diet and lifestyle measures such as smoking, weight and other factors might affect Lp-PLA₂ activity.⁶⁵ Another recent study by *Da Silva et al* reported that Lp-PLA₂ activity is influenced by glucose levels, HDL size and Apo B/Apo AI ratio in obese adolescents.⁶⁶

1.7 Lp-PLA₂ inhibition: the past, the present, and the future

At time of discovery, Lp-PLA₂ or better known at that time as PAF-AH was thought to have exclusive anti-inflammatory effects. PAF-AH studies showed significant reduction of severe inflammation in vivo and in vitro. It is ironic that what is known today as a proinflammatory enzyme was not very long ago known as an anti-inflammatory enzyme and was suggested as a treatment for inflammation. Studies suggesting a rather anti-inflammatory character of Lp-PLA₂ emphasize its role on degrading PAF and oxidized phospholipids on lipoproteins. At the

present time, the epidemiological and experimental data does not allow to solve the controversy on the protective or the proatherogenic role of Lp-PLA₂.

An argument can be made that despite the positive correlation between Lp-PLA₂ and increased risk of CVD, the information present is not enough to imply causality.⁵⁵ The only way to show causality is by undergoing clinical trials.

A normal level of Lp-PLA₂ is <200 ng/mL, while a level of ≥223 ng/mL is considered high and current guidelines do not warrant Lp-PLA₂ measurement as part of routine testing in patients with low risk for CAD.⁶⁷

Phospholipase A₂ inhibition presents an appealing therapeutic target. Two experimental phospholipase A₂ inhibitors exist: Darapladib (Lp-PLA₂ inhibitor) and Varespladib (sPLA₂ inhibitor). When we compare the two agents, we notice that they inhibit different PLA₂ isoforms. Darapladib uniquely inhibits Lp-PLA₂; Varespladib inhibits several secreted PLA₂ isoforms (Group IIa, Group V and Group X), which have shown to have proinflammatory effects in atherosclerotic plaque formation. Similar to Lp-PLA₂, they produce LPC and NEFA.¹⁸ In addition to Lp-PLA₂ inhibition, Darapladib lowered the expression of 24 genes, several of which are inflammatory markers.⁴⁶

The two agents (Darapladib, Varespladib) have been evaluated in several clinical trials, and in different in vitro and in vivo studies. Both have shown efficacy in inhibiting the enzymes in vitro, but lacked efficacy in humans.^{44, 45, 68} In a phase II randomized clinical trial that sought to look at the effects of Darapladib in patients with stable coronary heart disease or coronary heart disease risk equivalent, positive correlation was present between LDL-C and Lp-PLA₂ activity, while a negative correlation between HDL-C and Lp-PLA₂ was noticed.⁴⁹ In the phase II (**I**ntegrated **B**iomarker **A**nd **I**maging **S**tudy– **2**) IBIS-2 trial, more than 300 patients

with angiographically documented coronary heart disease and another risk factor were randomized to darapladib 160 mg/d or placebo for 12 months to examine the effects of Darapladib, where the results showed reduction in plaque areas and necrotic core in the treatment group as compared to the control group.

IBIS-2 paved the road for Darapladib to undergo two large randomized phase III clinical trials.

The STABILITY (STabilisation of Atherosclerotic plaque By Initiation of darapLadibTherapY) trial studied the effects of chronic treatment with Darapladib on the incidence of major adverse cardiovascular events (MACE) in high-risk patients with chronic CHD in 15,828 men or women with chronic coronary heart disease. STABILITY failed to meet the primary end points (relative risk reduction of 6%), despite some reductions in some of the secondary endpoints.^{68, 69} The SOLID-TIMI 52 trial (The Stabilization Of pLaques usIng Darapladib-Thrombolysis In Myocardial Infarction 52) evaluated if Darapladib can lower the chances of having a cardiovascular event when treatment is started within 30 days after ACS. This study involved 13,000 men or women hospitalized for ACS.⁷⁰ Overall, Darapladib appears to be well tolerated by patients, and has not been reported to have adverse effects, except for minor effects like diarrhea and odour.⁷¹ Nevertheless, SOLID-TIMI 52 suffered the same fate as STABILITY, and failed to meet its primary end point.⁷²

The other agent, Varespladib, did not have fruitful results. The phase III trial (Vista-16), which evaluated the safety and efficacy of Varespladib treatment on morbidity and mortality when added to Atorvastatin in 5,189 subjects with ACS, was unexpectedly terminated in March 2012. Varespladib results were consistent in all the previous trials it underwent, but it stumbled in Vista-16 due to unknown reasons at present.^{73, 74}

Lp-PLA₂ signifies a relation between oxidized lipoproteins (oxLDL) and inflammation and plaque susceptibility to rupture. It is highly possible that Lp-PLA₂ is a physiological responder rather than a cause of inflammation. Will Lp-PLA₂ have the same fate as hsCRP? The new results of the randomized clinical trials urge us to believe that Darapladib and other Lp-PLA₂ inhibitors may not see the light at the end of the tunnel. Until proven otherwise, Lp-PLA₂ is of limited benefit in a clinical setting.

Chapter II: Clinical Research

Hypothesis, aims and objectives

Acute coronary syndromes are underlined by a long-term inflammatory state. Lp-PLA₂ functions by oxidizing modified phospholipids on the surface of the lipoprotein, an action that results in producing and accumulation of molecules (namely lysophosphatidylcholine and non-esterified fatty acids) within the vessel wall, contributing to unstable plaque formation and aggravating inflammation. Multiple studies have observed an association between the role of Lp-PLA₂ in plaque formation and increased risk of coronary artery disease, suggesting an inflammatory role for Lp-PLA₂.

The hypothesis was that the inflammatory milieu accompanying Acute Coronary Syndromes alters Lp-PLA₂ levels and distribution (HDL and LDL). In the present study, we examined Lp-PLA₂ mass in patients with ACS acutely (within 48 hours) and again at ACS recovery (12 weeks after their first presentation). Stable CAD and healthy control subjects were used for comparison.

In the plasma, Lp-PLA₂ is mainly distributed between LDL-C and HDL-C, with a greater percentage on LDL-C. A secondary objective was to compare the distribution of Lp-PLA₂ in the plasma and among lipid fractions (HDL and LDL) in ACS acute and ACS recovery groups. To answer our research questions, we used two different separation techniques including ultracentrifugation (UTC) and high-performance liquid chromatography (HPLC).

Chapter III: Clinical Research

Lipoprotein-Associated Phospholipase A₂ (Lp-PLA₂) in Acute
Coronary Syndrome: Relationship with low-density lipoprotein
cholesterol

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Abstract

Background: Lipoprotein-associated phospholipase A₂ (Lp-PLA₂) might play a role in the formation of vulnerable atherosclerotic plaques. Its plasma distribution and mass in subjects with acute coronary syndrome (ACS) has yet to be characterized.

Methods: We compared plasma levels of Lp-PLA₂ in 24 patients within 48 hours of an ACS (acute) and 12 weeks after (recovery), in 26 patients with stable coronary artery disease and in 10 normal healthy control subjects. Lp-PLA₂ mass was determined using enzyme-linked immunosorbent assay.

Results: The ACS patients (mean age 57 ± 8.7 years) had high-sensitivity C-reactive protein (hsCRP) levels of 30.46 ± 57.57 mg/L (ACS acute) vs. 1.69 ± 1.32 mg/L (ACS recovery). Plasma Lp-PLA₂ levels were significantly higher in ACS acute subjects than in ACS recovery subjects (143.13 ± 60.88 ng/mL vs. 88.74 ± 39.12 ng/mL; $P < 0.0001$). Interestingly, stable coronary artery disease patients had higher Lp-PLA₂ levels than ACS recovery patients (121.72 ± 31.11 ng/mL vs. 88.74 ± 39.12 ng/mL; $P = 0.0018$). There was a strong correlation between Lp-PLA₂ and low-density lipoprotein (LDL) cholesterol (LDL-C) ($r = 0.709$; $P < 0.0001$) or changes in LDL ($r = 0.449$; $P = 0.027$), suggesting that the major determinant of plasma Lp-PLA₂ is LDL-C. No significant correlations were observed between Lp-PLA₂ and hsCRP or high-density lipoprotein (HDL) cholesterol. When separated using high-performance liquid chromatography, $> 65\%$ - 70% of Lp-PLA₂ mass was within the apolipoprotein B-containing lipoprotein fraction, with approximately 30% - 35% on HDL fraction, with no significant change in distribution between ACS acute and recovery.

Conclusions: Subjects with an ACS have markedly increased Lp-PLA₂ levels acutely related to LDL-C levels.

Résumé

Introduction: La phospholipase A₂ associée aux lipoprotéines (Lp-PLA₂) pourrait jouer un rôle dans la formation de plaques athéroscléreuses vulnérables. Sa distribution plasmatique et sa masse chez les sujets ayant un syndrome coronarien aigu (SCA) a encore à être déterminée.

Méthodes: Nous avons comparé les concentrations plasmatiques de Lp-PLA₂ chez 24 patients dans les 48 heures d'un SCA (aigu) et dans les 12 semaines après (rétablissement), chez 26 patients ayant une coronaropathie stable et chez 10 témoins en bonne santé. La masse de la Lp-PLA₂ a été déterminée à l'aide de l'immunoabsorption par enzyme liée.

Résultats: Les patients ayant un SCA (âge moyen de $57 \pm 8,7$ ans) ont eu des concentrations de protéine C réactive élevées de $30,46 \pm 57,57$ mg/l (SCA aigu) vs $1,69 \pm 1,32$ mg/l (rétablissement du SCA). Les concentrations plasmatiques de la Lp-PLA₂ ont été significativement plus élevées chez les sujets ayant un SCA aigu que chez les sujets rétablis d'un SCA ($143,13 \pm 60,88$ ng/ml vs $88,74 \pm 39,12$ ng/ml; $P < 0,0001$). Fait intéressant, les patients ayant une coronaropathie stable ont eu des concentrations de la Lp-PLA₂ plus élevées que les patients rétablis d'un SCA ($121,72 \pm 31,11$ ng/ml vs $88,74 \pm 39,12$ ng/ml; $P = 0,0018$). Il existait une forte corrélation entre la Lp-PLA₂ et le cholestérol à lipoprotéines de faible densité (C-LDL; $r = 0,709$; $P < 0,0001$) ou les changements dans le LDL ($r = 0,449$; $P = 0,027$), ce qui suggère que le déterminant majeur de la Lp-PLA₂ plasmatique est le C-LDL. Aucune corrélation significative n'a été observée entre la Lp-PLA₂ et la protéine C réactive hautement sensible ou le cholestérol à lipoprotéines de haute densité (HDL). À la séparation par la chromatographie à haute performance, $> 65\%$ - 70% de la masse de la Lp-PLA₂ a été dans la fraction de la lipoprotéine contenant l'apolipoprotéine B, dont approximativement 30%

-35% sur la fraction de HDL, et aucun changement significatif dans la distribution entre le SCA aigu et le rétablissement du SCA.

Conclusions: Les sujets ayant un SCA ont en phase aigue une augmentation significative des concentrations de la Lp-PLA₂ liés aux taux de C-LDL.

Lipoprotein-associated phospholipase A₂ (Lp-PLA₂) is a novel biomarker that is postulated to play a role in the formation of atherosclerotic plaques and might lead to plaque instability. Human Lp-PLA₂ is encoded by the *PLA2G7* gene.⁷⁵ It is produced by many cells, inflammatory (monocytes/macrophages, mast cells) and noninflammatory cells (liver Kupffer cells, platelets, erythrocytes), but it appears that macrophages are the main producers.²³ Similar to other phospholipase A₂ enzymes, Lp-PLA₂ catalyzes the hydrolysis of the ester bond at the sn-2 position of phospholipids. Lp-PLA₂ is bound to lipoproteins, predominantly to low-density lipoprotein (LDL) (approximately 80%), with a significantly smaller portion on high-density lipoprotein (HDL) (approximately 20%),⁷⁶ hence the name, Lipoprotein-associated phospholipase A₂. Originally described as platelet activating factor acetyl hydrolase, Lp-PLA₂ cleaves the oxidized glycerophospholipid acetyl-glyceryl-ether-phosphorylcholine in the sn-2 position to yield lysophosphatidylcholine (LPC) and an acetyl group. However, the substrate specificity of Lp-PLA₂ is not well defined and it is thought to inactivate highly reactive oxidized phospholipids on lipoproteins. The products of the Lp-PLA₂ action, LPC and nonesterified fatty acids, are also known to be proinflammatory and to be involved in cell signalling.³¹ Lp-PLA₂ thus represents a link between oxidized lipoproteins, inflammation, and plaque instability. In the arterial wall, LDL is modified into oxidized lipoproteins under oxidative stress, modifying some of the phospholipids on the LDL particle. The production of oxidized phospholipids activates LDL-bound Lp-PLA₂. Lp-PLA₂ hydrolyzes those oxidized phospholipids (mainly oxidized phosphatidylcholine) and generates 2 different molecules: LPC and nonesterified fatty acids.²³

Lp-PLA₂ is found in atherosclerotic plaques using immunohistology and its presence correlates with inflammatory cells and histological characteristics of plaque instability.⁷⁷ A

meta-analysis of Lp-PLA₂ in cardiovascular disease (CVD) reveals that Lp-PLA₂ levels (mass and activity) are significantly associated with each other and with proatherogenic lipid markers. Lp-PLA₂ levels are significantly related to CVD risk in a continuous and graded manner.⁵⁵ Because Lp-PLA₂ levels predict future cardiovascular events, the measurement of Lp-PLA₂ might be considered (Class IIb) in intermediate-risk asymptomatic adults.⁷⁸

The effects of lipid-lowering therapies in Lp-PLA₂ levels have been previously examined,^{48, 79} and the data show that statins significantly lower Lp-PLA₂ levels. *Albert et al* showed that approximately 6% of the variability in the response of Lp-PLA₂ to pravastatin was accounted for by a reduction in LDL cholesterol (LDL-C) levels.⁸⁰ In most reports, the effect of statins on Lp-PLA₂ levels was significantly attenuated after adjusting for changes in LDL-C levels, reflecting the strong affiliation between these 2 substances in plasma. A recent study of Lp-PLA₂ levels in the first 3 days after an acute coronary syndrome (ACS) reveals that Lp-PLA₂ levels probably increase acutely, but that the subsequent change in levels correlate with the change in LDL-C.⁵⁷ Previous data have not shown any significant correlations between reductions in Lp-PLA₂ levels and changes in HDL cholesterol (HDL-C) concentrations.

The purpose of the present study was to examine plasma levels of Lp-PLA₂ in ACS and its relationship to high-sensitivity C-reactive protein (hsCRP) and lipoprotein levels. Unlike C-reactive protein, Lp-PLA₂ does not seem to behave as an acute-phase reactant.⁴² We hypothesized that the inflammatory milieu that is associated with an ACS might alter plasma levels of Lp-PLA₂. The main objective of this study was to determine plasma levels and distribution of Lp-PLA₂ at the time of an ACS (within 48 hours of onset of symptoms) and again, 12 weeks later, compared with patients with stable, chronic coronary artery disease

(CAD) (> 1 year after the event). We also determined the concentration of Lp-PLA₂ in lipoprotein fractions.

Methods

Study subjects.

Patients, men and women aged between 18 and 80 years, were selected from the McGill University Health Center, Canada. Subjects within the ACS acute group were sampled within 48 hours of symptom onset (n = 24) according to current diagnostic criteria.⁸¹ The diagnosis of ACS was confirmed in a clinical assessment by a cardiologist (J.G.), including electrocardiographic changes (non-ST and ST segment elevation acute myocardial infarction), troponin I elevation, and the presence of CAD on coronary angiography. The same patients were resampled 12 weeks later (ACS recovery; n = 24). The 12-week time point was selected because most of the metabolic, lipoprotein, and inflammatory abnormalities have resolved by then.⁸² As comparison groups, we also examined subjects with CAD and normal healthy control subjects (normals). The CAD group consisted of patients with chronic, stable CAD who had been followed in the lipid clinic for at least 11 year after a cardiovascular event, and the normals consisted of age-matched healthy subjects without CAD. These subjects had no CAD, documented using coronary angiography; all were awaiting elective aortic or mitral valve replacement or repair, and were not taking statin therapy. All subjects with stable CAD continued taking statins at the time of sampling, according to current guidelines and ethics requirements.⁸³⁻⁸⁶ Exclusion criteria in the ACS patients included refusal to participate, inability to return for a 12-week follow-up visit, hemodynamic instability requiring vasopressor support, cardiac support devices, the need for urgent coronary artery bypass surgery, and the lack of documented atherosclerotic CAD, uncontrolled hypertension, triglycerides ≥ 5 mmol/L, severe obesity (body mass index ≥ 35), alcohol intake > 21 drinks per week, and presence of thyroid, hepatic, or renal disease. Subjects were excluded if they

had autoimmune disease or any chronic or acute infectious or inflammatory illness. The research protocol was reviewed and approved by the Research Ethics Board of the McGill University Health Center; all subjects gave written informed consent to participate.

Plasma lipids, lipoprotein lipids, lipoprotein isolation, apolipoprotein B, hsCRP, and Lp-PLA₂ mass assays.

Blood was collected from study subjects and control subjects using venipuncture; plasma was separated (2000g, 4 °C for 20 minutes) and multiple aliquots were frozen at -80 °C in 0.5-mL aliquots. Total cholesterol, triglycerides, and HDL-C were determined enzymatically and LDL-C was determined using the Friedewald formula. A sample of plasma (9 mL) was immediately processed for isolation of lipoproteins using ultracentrifugation (UTC). We isolated apolipoprotein B (apoB)-containing lipoproteins (LpB) in the < 1.09 g/mL density fraction and HDL using sequential UTC at a density between 1.09 g/mL and 1.21 g/mL using KBr as a salt. Isolated LpB and HDL fractions were then extensively dialyzed against phosphate-buffered saline, and total protein concentrations in these fractions were measured using a modified Bradford procedure (BioRad, Mississauga, Canada). Apolipoprotein A-1, apoB, and hsCRP concentrations were determined using nephelometry (Behring Nephelometer 100 Analyzer). Lp-PLA₂ concentrations (mass) were measured using a commercial Lp-PLA₂ enzyme-linked immunosorbent assay (Quantikine ELISA, R&D Systems).

Pools of plasma (125 uL) from the ACS acute and the ACS recovery group (n = 24) were separated into lipoprotein fractions using high-performance liquid chromatography (HPLC) with a Superose 6 10/300 GL column (GE Healthcare). A 150-mM NaCl mobile phase with a

flow rate of 0.4 mL/mL was used for the separation of the sample into seventy-two 400-uL HPLC fractions that were collected. Total cholesterol was determined enzymatically using the Infinity Cholesterol Liquid Stable Reagent (Thermo Electron Corporation) following the manufacturer's instructions.

Statistical analyses.

Discrete variables are described as mean \pm SD. For comparisons between ACS acute and ACS recovery, 2-tailed paired t tests with equal variance were selected. A *P* value of < 0.05 was considered significant. Pairwise comparisons between ACS acute, CAD, and Normals were considered significant at the 0.05 level. Pearson correlation coefficients were used to examine the relationship between Lp-PLA₂ and LDL-C, apoB, hsCRP, HDL, and troponins. Linear regression was used to determine the effect of LDL-C-lowering on Lp-PLA₂ levels. All statistical analyses were performed using the GraphPad Prism software V5.0 (La Jolla, CA).

Results

Patient characteristics are summarized in Table 1. There were 24 patients in the ACS groups (19 men, 5 women, mean age 57 ± 9 years), 26 with chronic, stable CAD (24 men, 2 women, mean age 62 ± 9 years) and 10 Normals (10 men aged 57 ± 17 years). Mean hsCRP level in ACS acute subjects was 30.46 ± 57.57 mg/L and decreased to 1.69 ± 1.32 mg/L in ACS recovery, not significantly different from CAD (1.30 ± 1.22 mg/L) and Normals (1.85 ± 1.50 mg/L). Peak troponin I level was 24.54 ± 36.29 ug/L in ACS acute (reference range, < 0.06 mg/L), indicating acute myocardial injury. Plasma levels of LDL-C differed markedly between groups. Approximately half of the patients in ACS acute were taking various doses of statins at the time of sampling; the LDL-C in ACS acute was 2.35 ± 0.76 mmol/L. As per clinical practice guidelines,⁸³⁻⁸⁶ ACS patients were put on a high-dose statin regimen (predominantly atorvastatin 80 mg or rosuvastatin 40 mg) and the LDL-C decreased to 1.52 ± 0.43 mmol/L at 12 weeks ($P < 0.0001$ ACS acute vs. ACS recovery) (Table 1). CAD patients had a mean LDL-C of 2.03 ± 0.43 mmol/L. Normals were not using statins by design and the mean LDL-C was 3.19 ± 0.86 mmol/L. HDL-C levels were similar between ACS acute, ACS recovery, CAD, and Normals (Fig. 1).

Table 1. Study subject characteristics

Characteristic	ACS acute	ACS recovery	CAD	Normals
n	24	24	26	10
Age, y	57 ± 9	57 ± 9	62 ± 9	57 ± 17
Sex	M 19/F 5	M 19/F 5	M 24/F 2	M 10/F 0
hsCRP, mg/L	30.46 ± 57.57	1.69 ± 1.32	1.30 ± 1.22	1.85 ± 1.50
Total cholesterol, mmol/L	4.19 ± 0.89	3.09 ± 0.68	3.80 ± 0.60	5.11 ± 1.04
Triglycerides, mmol/L	1.82 ± 1.07	1.23 ± 0.60	1.48 ± 0.91	1.99 ± 1.22
HDL-C, mmol/L	1.02 ± 0.24	1 ± 0.29	1.10 ± 0.28	1.14 ± 0.17
LDL-C, mmol/L	2.35 ± 0.76	1.52 ± 0.43	2.03 ± 0.43	3.19 ± 0.86
ApoB, g/L	0.85 ± 0.20	0.61 ± 0.12	0.75 ± 0.14	0.99 ± 0.20
Lp-PLA ₂ in plasma, ng/ml	143.13 ± 60.88	88.74 ± 39.12	121.72 ± 31.11	188.70 ± 61.53

Relevant P values are shown in Figures 1 and 2.

ACS, acute coronary syndrome; ApoB, apolipoprotein B; CAD, coronary artery disease; F, female; HDL-C, high-density lipoprotein cholesterol; hsCRP, high-sensitivity C-reactive protein; LDL-C, low-density lipoprotein cholesterol; Lp-PLA₂, Lipoprotein-associated phospholipase A₂; M, male.

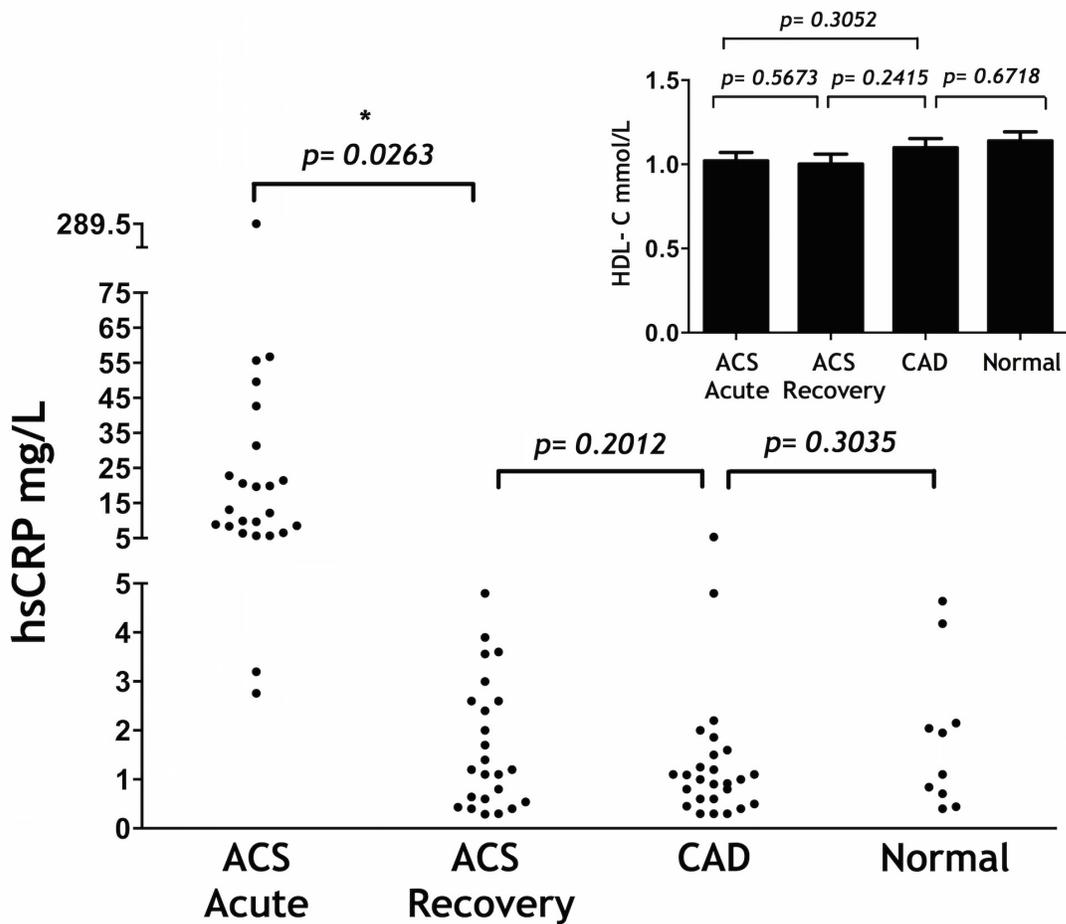


Figure 1. hsCRP and HDL-C (inset) levels in study subjects. Patients with an ACS (ACS acute) were sampled within 48 hours of presentation. The same patients were sampled 12 weeks later (ACS recovery). Note the increase in hsCRP in the ACS acute group, compared with the recovery, CAD, and normal subjects. ACS, acute coronary syndrome; CAD, coronary artery disease; HDL-C, high-density lipoprotein cholesterol; hsCRP, high-sensitivity C-reactive protein.

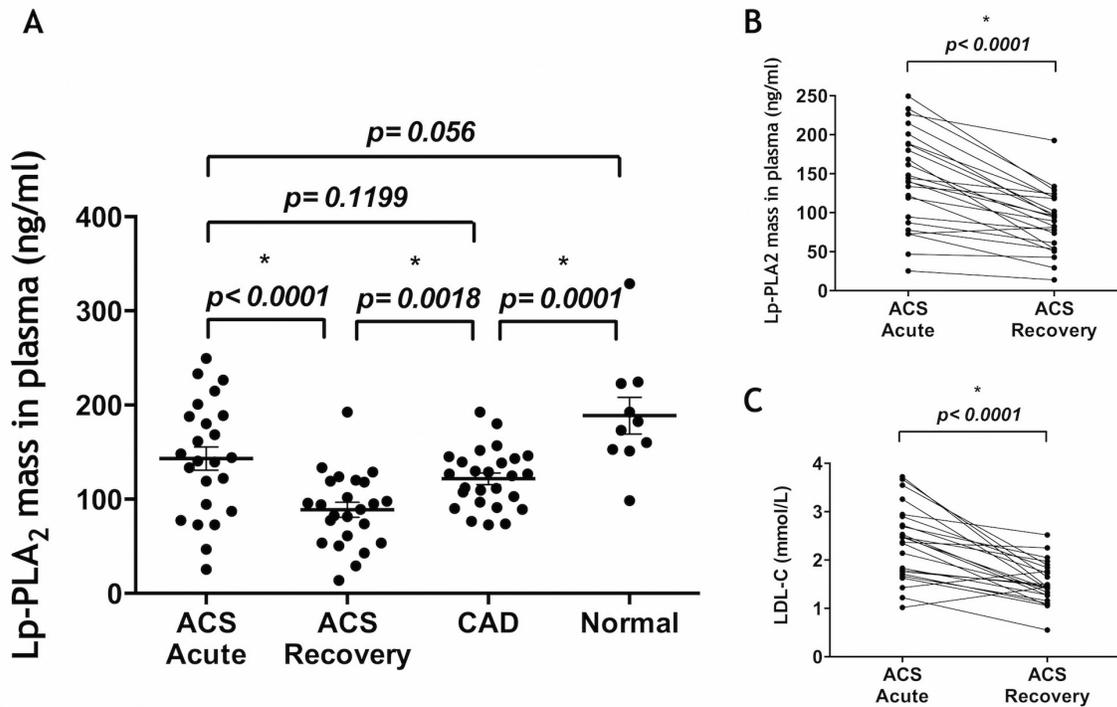


Figure 2. (A) Lp-PLA₂ levels in the plasma of study subjects: ACS acute (n = 24), ACS recovery (n = 24), CAD (n = 26), and normal healthy subjects (n = 10). (B) Difference in Lp-PLA₂ mass between ACS acute and recovery. (C) Difference in LDL-C between the same groups. ACS, acute coronary syndrome; CAD, coronary artery disease; LDL-C, low-density lipoprotein cholesterol; Lp-PLA₂, lipoprotein-associated phospholipase A₂.

Plasma Lp-PLA₂ levels were significantly increased in ACS acute (143.13 ± 60.88 ng/mL) compared with ACS recovery (88.74 ± 39.12 ng/mL; Fig. 2). CAD subjects had slightly higher levels than ACS recovery (121.72 ± 31.11 ng/mL) and Normals had Lp-PLA₂ levels in the normal range (188.70 ± 61.53 ng/mL), and not significantly higher than the ACS acute subjects (reference range, < 200 ng/mL).⁶⁷ In all patients, Lp-PLA₂ levels decreased at 12 weeks (Fig. 2B), and paralleled the decrease in LDL-C (Fig. 2C).

UTC was used to separate LpB at a density of 1.006 g/L $<$ LpB < 1.09 g/L and HDL $d = 1.09$ g/L $<$ HDL < 1.21 g/L. Only approximately 50% of Lp-PLA₂ mass was recovered in the LpB fraction (Supplementary Figure 1A) (43% of total) and 3% in the HDL fraction (data not shown), suggesting that Lp-PLA₂ is only loosely bound to lipoproteins and can be sheared off during UTC.

We then separated lipoproteins using HPLC as shown in Figure 3A. The Lp-PLA₂ distribution closely matched that of the LDL-C peak. A representative control HPLC analysis from a patient with high triglycerides (Fig. 3B) and high HDL-C (Fig. 3C) confirmed that Lp-PLA₂ resides predominantly within LDL. HPLC analysis of serum did not reveal a shift in the distribution of Lp-PLA₂ between HDL and LDL. UTC, as previously stated, caused a significant loss of LDL- and HDL- associated Lp-PLA₂, making quantification unreliable.

When examining the correlations of Lp-PLA₂ in all subjects, there was a strong and highly significant correlation between LDL-C and Lp-PLA₂ ($r = 0.709$; $r^2 = 0.503$; $P < 0.0001$), suggesting that half the variance in Lp-PLA₂ can be explained by LDL-C levels (Fig. 4A and B). Similar data were obtained for total plasma apoB ($r = 0.775$; $r^2 = 0.6009$; $P < 0.0001$). The change in Lp-PLA₂ also correlated strongly with change in LDL-C between ACS acute and ACS recovery patients ($n = 24$; $r = 0.449$; $P = 0.027$) (Fig. 4B). These data suggest that LDL is

the major determinant of Lp-PLA₂. In the 4 groups combined, the correlation between triglycerides and plasma Lp-PLA₂ levels was $r = 0.3868$ ($n = 84$; $P = 0.0003$) (Fig. 4D). No significant correlation was observed between HDL-C and Lp-PLA₂ ($r = 0.181$; $P = 0.099$) (Fig. 4E), between hsCRP and Lp-PLA₂ ($r = -0.023$; $P = 0.839$) (Fig. 4F) or between troponin I and Lp-PLA₂ in ACS acute (data not shown). When we examined groups separately, there was no correlation between hsCRP and Lp-PLA₂ levels. The correlations for all variables are shown in Table 2.

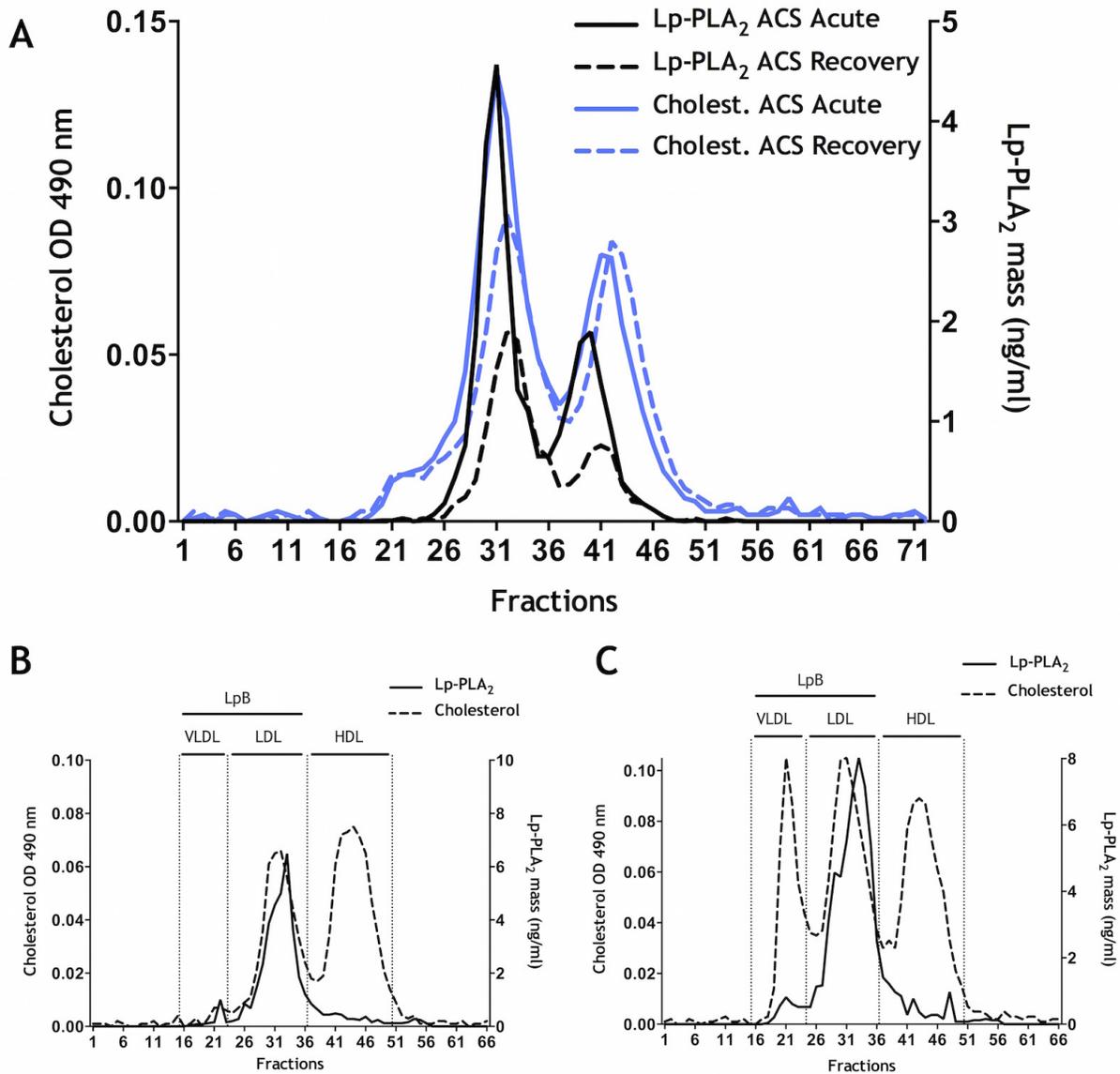


Figure 3. Distribution of Lp-PLA₂ mass. (A) Lp-PLA₂ and cholesterol mass assays performed after HPLC separation of pooled plasma samples from ACS acute (n = 24) and ACS recovery (n = 24). Note that the distribution of Lp-PLA₂ parallels that of LDL predominantly. (B) and

(C) show the distribution of Lp-PLA₂ in 2 control subjects, a normo-lipidemic subject (B) and 1 with elevated triglyceride levels (C), showing that Lp-PLA₂ remains predominantly within the LDL-size fraction. ACS, acute coronary syndrome; HPLC, high-performance liquid chromatography; LDL, low-density lipoprotein; Lp-PLA₂, lipoprotein-associated phospholipase A₂; OD, optical density.

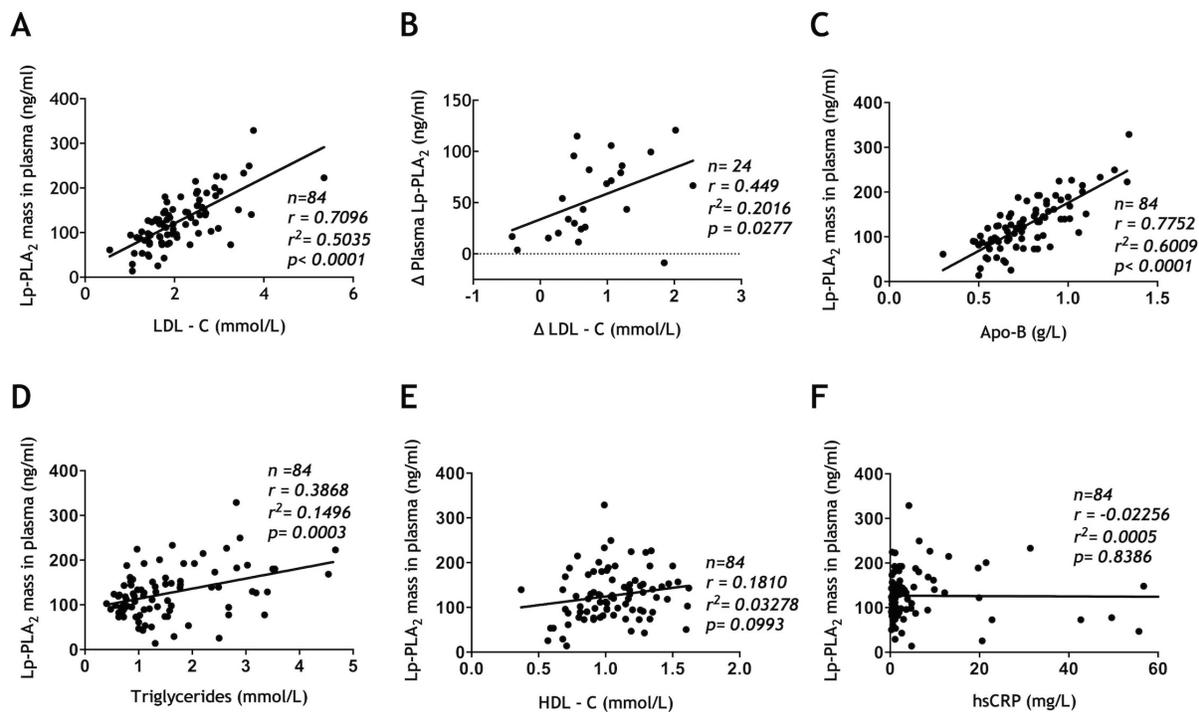


Figure 4. Correlations between Lp-PLA₂ and LDL-C (A); change in LDL-C (B); Apo-B (C); HDL-C (D); triglycerides (E), and hsCRP (F). Apo-B, apolipoprotein B; HDL-C, high-density lipoprotein cholesterol; hsCRP, high-sensitivity C-reactive protein; LDL-C, low-density lipoprotein cholesterol; Lp-PLA₂, lipoprotein-associated phospholipase A₂.

Table 2. Pearson correlation coefficients of Lp-PLA₂ mass with cardiovascular risk factors in the study subjects.

Factor	Lp-PLA ₂ mass	
	<i>r</i>	<i>p</i>
Age, y	0.02612	0.8136
BMI	-0.2438	0.0351
hsCRP, mg/L	-0.02256	0.8386
Troponins *	0.1044	0.6355
Total Cholesterol, mmol/L	0.7508	< 0.0001
Triglycerides, mmol/L	0.3868	0.0003
LDL-C, g/L	0.7096	< 0.0001
Δ LDL-C, g/L **	0.449	0.0277
HDL-C, mmol/L	0.181	0.0993
ApoB, g/L	0.7752	< 0.0001
ApoA1, g/L	0.2643	0.0158

ACS, acute coronary syndrome; ApoA1, apolipoprotein A1; ApoB, apolipoprotein B; BMI, body mass index; HDL-C, high-density lipoprotein cholesterol; hsCRP, high-sensitivity C-reactive protein; LDL-C, low-density lipoprotein cholesterol; Lp-PLA₂, lipoprotein-associated phospholipase A₂.

* For ACS acute only (n = 24).

** Between ACS acute and ACS recovery (n = 24).

Discussion

In the present study, we showed that Lp-PLA₂ in human is not an acute phase reactant and does not increase in acute inflammation. We also showed that Lp-PLA₂ mass is closely related to LDL and LpB, with only a minor fraction within HDL. This has important implications if Lp-PLA₂ is considered a therapeutic target. Lowering LDL-C with statins has a marked and proportional effect on Lp-PLA₂ mass.

A meta-analysis of the association between Lp-PLA₂ mass or activity in plasma or serum shows that Lp-PLA₂ levels predict recurrent cardiovascular risk. A small molecule, darapladib, is a specific inhibitor of Lp-PLA₂ function and is currently undergoing clinical trials in patients with CAD. In the **Stabilization of Atherosclerotic Plaque by Initiation of Darapladib Therapy (STABILITY)** trial, more than 15,000 patients with established CAD and another risk factor were randomized to darapladib 160 mg/d or placebo and a background of appropriate lipid-lowering therapy with statins.⁶⁸ In the **Stabilization of Plaques using Darapladib - Thrombolysis in Myocardial Infarction 52 (SOLID-TIMI 52)** trial, 13,000 patients with an ACS have been randomized to darapladib or placebo.⁷⁰ Both trials are expected to be complete in 2013 and 2014 respectively. Importantly, both trials will be performed in patients treated to strict LDL-C goals. Considering the redundancy of phospholipases A₂ - there are 15 known phospholipase A₂ groups comprising as many isoforms in humans⁸⁷; it is not certain that this approach will prevent CVD.

A similar approach was used with varespladib, an inhibitor of the secretory phospholipase A₂ isoforms IIa, V, and X. In the **Vascular Inflammation Suppression to Treat Acute Coronary Syndrome-16 Weeks (VISTA-16)** trial, varespladib failed to meet its primary end point of CVD reduction.^{70, 73, 74, 88}

Previous groups have examined Lp-PLA₂ levels in the early phase of an ACS. *Dullaart et al* reported that Lp-PLA₂ levels in ACS did not differ from those found in patients with noncardiac chest pains. Interestingly, Lp-PLA₂ levels were greater in a subset of patients with ST segment elevation myocardial infarction. The study did not control for baseline lipid values.⁸⁹ *Ostadal et al* recently reported that Lp-PLA₂ levels were increased in the initial hours of an ACS, compared with in the following days. They reported an observation similar to ours, showing a relationship between LDL-C and Lp-PLA₂ and a lack of correlation with C-reactive protein and troponin I.⁵⁷

The distribution of Lp-PLA₂ on lipoproteins was demonstrated many years ago,^{76, 90} and revealed that a minor percentage (approximately 20%) was associated with HDL. In a prospective study, Lp-PLA₂ in plasma and within HDL was determined in more than 500 patients who were followed for 3 years. Although Lp-PLA₂ in plasma levels were associated with worse outcomes, HDL-associated Lp-PLA₂ mass was associated with a lower risk (hazard ratio, 0.972; 95% confidence interval, 0.952-0.993; $P = 0.010$). An even more impressive effect of HDL-associated Lp-PLA₂ activity on cardiac death (hazard ratio, 0.689; 95% confidence interval, 0.496-0.957; $P = 0.026$) was observed.⁵²

The biology of Lp-PLA₂ is only partly understood. Initially described as a platelet-activating factor (or 1-O-alkyl-2-acetyl-sn-glycerol-3-phosphorylcholine) acetylhydrolase, its physiological role remains uncertain. It is possible that on LDL, Lp-PLA₂ hydrolyzes oxidized phospholipids, generating LPC and oxidized lipids that are deleterious within an atherosclerotic plaque, whereas when present on HDL, it neutralizes platelet-activating factor and prevents platelet activation. Lp-PLA₂ is present in atherosclerotic plaques, as shown using immunohistochemistry and its presence correlates with histological features of plaque

instability.⁷⁷ However, a causal relationship between Lp-PLA₂ presence and plaque rupture has not been yet demonstrated. Thus, for the present time, the measurement of Lp-PLA₂ mass should be considered a biomarker of atherosclerosis. Its pathological role and potential as a therapeutic target must be further studied.

The main conclusion of the present study is the very close correlation between LDL-C and Lp-PLA₂ levels, suggesting that statin therapy might contribute to the reduction in cardiovascular events in a pleiotropic fashion. Our study has several limitations. The sample size was relatively small. We were careful to compare the same patients in ACS acute and ACS recovery groups and to match CAD patients and Normals for age and HDL-C levels. We did not determine Lp-PLA₂ activity; however, previous studies have shown a strong correlation between mass and activity.^{55,90} Importantly, the quantification of Lp-PLA₂ within lipoprotein fractions is limited by the technique used. We have shown that UTC causes loss of Lp-PLA₂ mass, most likely because of the shear stress engendered. More problematic is the quantification of Lp-PLA₂ within the HDL range. It is extremely difficult to separate HDL particles from plasma proteins using HPLC. Thus, “lipoprotein-free” Lp-PLA₂ (i.e. not bound to lipoproteins is very difficult to quantify). In our previous study of HDL proteomics in ACS, Lp-PLA₂ was not identified as 1 of the 70 proteins within HDL.⁹¹

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Chapter IV:

Discussion, conclusion and future work

4.1 Discussion and conclusion

The purpose of the study was to investigate the role of Lp-PLA₂ in patients with acute coronary syndromes. Acute coronary syndromes are characterized by systemic inflammation, as evidenced by hsCRP levels and serum amyloid levels. Because of the importance of Lp-PLA₂ in the pathogenesis of atherosclerosis, and the development of agents that block Lp-PLA₂, we undertook this study to examine whether Lp-PLA₂ levels vary in patients with an acute inflammatory state. To further explore the relationship between Lp-PLA₂ and ACS events, we hypothesized that Lp-PLA₂ levels would be elevated in acute coronary syndromes acutely, reflecting an overall inflammatory state.

We measured the levels of Lp-PLA₂ in patients with acute coronary syndromes within 48 hours of their first presentation, and three months later once the inflammation has subsided. For comparison, we used control patients who were not on medications, and have no evidence of coronary artery disease on coronary angiography (those patients were selected for elective valvular replacement but no bypass surgery). A second control group of stable chronic coronary artery disease patients at least one year post cardiac event were selected.

Subjects with ACS in the acute state exhibited a significant increase in Lp-PLA₂ levels compared to ACS recovery. Lp-PLA₂ levels were higher in stable chronic coronary artery disease patients than in subject with ACS in the recovery phase. A strong correlation was noted between Lp-PLA₂ and LDL-C, and the difference in Lp-PLA₂ levels strongly correlated with the difference in LDL-C levels in ACS acute and ACS recovery groups. Those correlations suggest that LDL-C levels have a major influence on Lp-PLA₂ levels, and might

partly explain the reduction of Lp-PLA₂ levels in the ACS recovery group, as Lp-PLA₂ resided mostly within LDL. When we compared the distribution of Lp-PLA₂ among the plasma and different lipid fractions (HDL, LDL) in ACS acute and ACS recovery, we did not notice any difference in Lp-PLA₂ distribution between HDL and LDL. It is important to mention that there was lack of correlation between Lp-PLA₂ and hsCRP and troponin I.

The hypothesis was not confirmed as Lp-PLA₂ levels went down dramatically along with LDL-C levels, and this is probably due to the fact the Lp-PLA₂ travels predominantly with LDL. The main conclusion of the study was that the significant increase in Lp-PLA₂ levels in subjects with an acute ACS, an increase that was mostly attributed to increased LDL-C in those subjects.

The role that Lp-PLA₂ plays in cardiovascular disease is still unclear. However, there have been new insights on the role of Lp-PLA₂ in plaque formation. To better understand that role, *Fanning et al* induced Diabetes mellitus (DM) and hypercholesterolemia (HC) 37 pigs. Abdominal aortic plaques exhibited more atherosclerosis, greater plaque area (4.4 fold), more calcifications and higher risk for intraplaque haemorrhage as compared to plaques in the coronary arteries. When Lp-PLA₂ inhibition was induced in 20 pigs, Darapladib significantly reduced intraplaque haemorrhage in the abdominal aortic plaques, with no significant effect on the coronaries. However, the study reported greater plaque reduction in the coronaries as compared to the abdominal aortic plaques (2.9 fold) with Darapladib treatment. The reduced effect of Darapladib on plaque reduction in the abdominal aorta compared to coronaries suggests site-specific effect of Darapladib on plaque reduction. This finding also suggests that Lp-PLA₂ was more involved in the inflammatory pathways that led to plaque formation in the

coronaries. Site-specific results of Darapladib might reduce the favourability of Darapladib use as potential treatment for atherosclerosis.⁹²

While studies demonstrated encouraging results for Lp-PLA₂ inhibition in animal models, clinical trials had disappointing outcomes. The results from two large clinical trials (STABILITY and SOLID-TIMI 52) show that Lp-PLA₂ inhibition by Darapladib did not change patient outcome, questioning the potential benefit of Lp-PLA₂ inhibition.^{68,70}

The enthusiasm for measuring Lp-PLA₂ as a biomarker of risk has dropped markedly due to two reasons. The first reason would be the finding that Lp-PLA₂ is strongly associated with LDL-C, and that statins can bring Lp-PLA₂ levels down markedly. The second reason would be that two trials of agents that specifically block Lp-PLA₂ have had neutral results. While those agents were well tolerated, they did not change cardiovascular outcomes.

In other cardiovascular diseases such as stroke, Lp-PLA₂ did not encounter much success. Although regarded as a predictor of risk in vascular diseases including acute coronary syndrome and stroke, Lp-PLA₂ has not shown to have an additional prediction value. A study by *Riba-Llena et al* included 921 stroke-free subjects to assess whether novel biomarkers such as Lp-PLA₂ could be a valuable addition to standard biomarkers such as troponin I and brain natriuretic peptide (BNP) in predicting silent brain infarcts. Lp-PLA₂ activity was associated with silent brain infarcts in women in an independent fashion, but unfortunately Lp-PLA₂ failed to add any value in disease prediction in that specific population. In men, Lp-PLA₂ activity did not show significant association with silent brain infarcts, despite adjusting for several factors including age, blood pressure, cholesterol levels and statin therapy.⁹³

Another study by *Tai et al* studied Lp-PLA₂ in the context of brain ischemia. Lp-PLA₂ mass and activity were measured in 100 subjects, where 58 subjects of the total population suffered ischemic events (transient ischemic attack and stroke). Lp-PLA₂ did not show statistical significance in subjects with ischemia compared to non-ischemic subjects, and did not exhibit diagnostic value in acute brain ischemia.⁹⁴

There are 15 phospholipase A₂ (PLA₂) isoforms, and the by-products from these reactions, which include highly oxidized fatty acyl chains may themselves be toxic to the vascular endothelial cells, and the resulting product (lyso phospholipids) may be involved in signalling. It is therefore not clear whether the activity of this enzyme is beneficial in terms of cardiovascular disease or not, and the body of knowledge does not allow us to make a firm commitment. Another interesting point to mention is that the biological system seems highly redundant because of the multiplicity of lipases with various specificities in plasma.

A Mendelian randomization meta-analysis by *Holmes et al* analysed 19 general population studies and 10 acute coronary syndrome studies in an attempt to investigate a possible causal relationship between secreted PLA₂ (sPLA₂-IIA mass) and major vascular events. *Holmes et al* could not establish a causal relationship, and concluded that secretory PLA₂ inhibition may not be of clinical benefit in preventing cardiovascular events.⁹⁵

Another genetic study by *Polfus et al* performed exome sequencing for 6325 subjects to evaluate a possible causal effect of Lp-PLA₂ on the incidence of coronary disease. Lp-PLA₂ activity was strongly associated to one variant (rs34159425), which was described by the authors as a variance of low frequency that has no significant association with coronary events. The researchers concluded the unlikely presence of a casual relationship between Lp-

PLA₂ and the incidence of coronary disease.⁹⁶ Based on the two previous studies, Lp-PLA₂ was not found to increase cardiovascular disease. Both studies reduce the possibility that Lp-PLA₂ has a causal link to cardiovascular disease.

To conclude, current evidence indicates that Lp-PLA₂ is involved in the pathophysiology of acute coronary syndrome and cardiovascular diseases in general, with no causal relationship identified. Therefore, current role of Lp-PLA₂ in clinical therapy and as a standard biomarker is incomplete with lack of supporting evidence.

4.2 Future work

Although the present study is limited by a relatively small number of patients, the results obtained are important. Thus, we will aim to increase sample size in future work. The loss of Lp-PLA₂ mass in Ultracentrifugation due to high speed involved in the nature of this technique is another point to be addressed. Therefore, we might include additional techniques for sample preparation in our future work, such as isolating HDL by polyethyleneglycol (PEG), thus avoiding losing Lp-PLA₂ mass. Moreover, additional experiments assessing Lp-PLA₂ role could be added, ideally measuring Lp-PLA₂ activity as an indicator of Lp-PLA₂ function.

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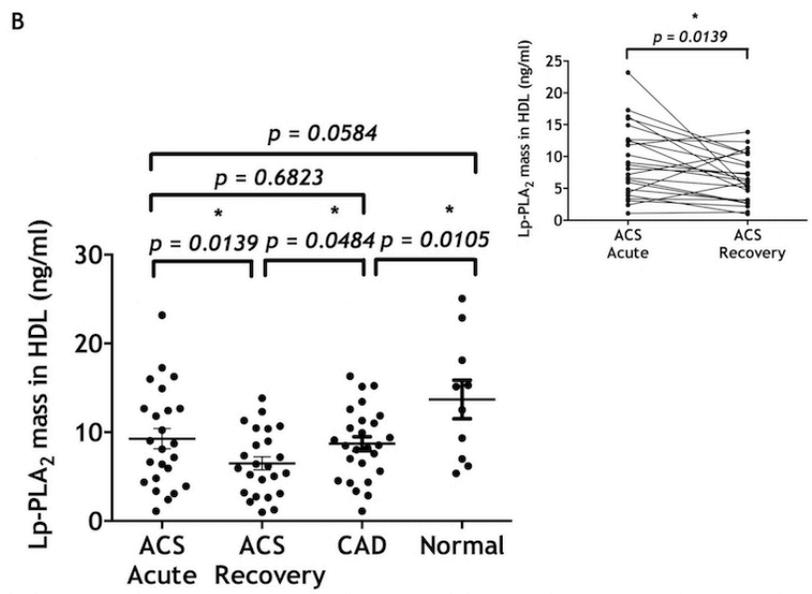
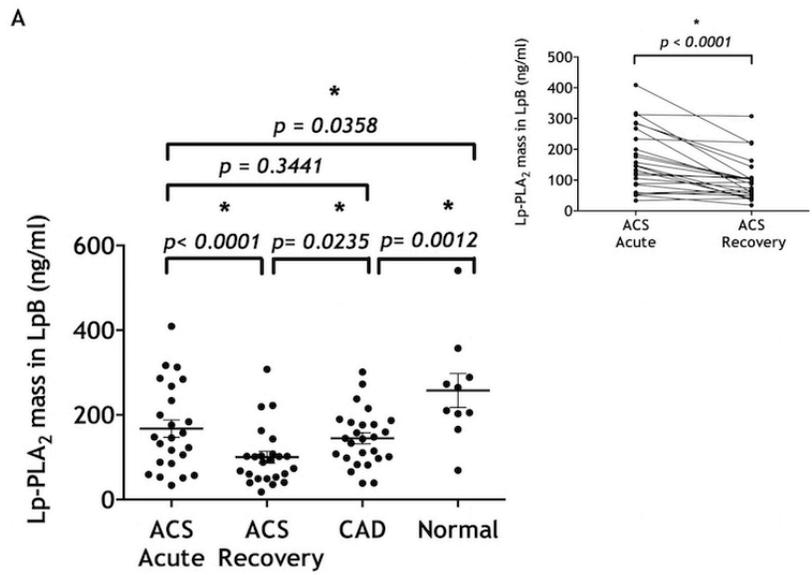
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Annex 1



Supplementary Figure 1. (A) Lp-PLA₂ mass in the apoB-containing lipoprotein fraction (LpB) obtained by UTC. Note the results closely parallel those obtained in **Figure 2**. (B) Lp-PLA₂ mass in high-density lipoprotein (HDL) fraction. Measurements performed in ACS acute (n=24), ACS recovery (n=24), Stable CAD (n=26), and normal healthy subjects (n=10). Note the different units for both the LpB and HDL fractions reflecting the volumes and concentrations obtained after UTC and sample dialysis.