

Université de Montréal

Modulation of the 5-HT₃ Receptor as a Novel Anti-Dyskinetic Target in Parkinson's Disease

par Cynthia Kwan

Département de pharmacologie et physiologie

Faculté de Médecine

Mémoire présenté

en vue de l'obtention du grade de maîtrise

en pharmacologie

option neuropharmacologie

Décembre, 2017

© Cynthia Kwan, 2017

Résumé

La L-3,4-dihydroxyphénylalanine (L-DOPA) est le traitement le plus efficace de la maladie de Parkinson. Cependant, avec une administration chronique de L-DOPA, les patients développent des complications motrices telles que les dyskinésies. Des études antérieures ont montré que le blocage des récepteurs type 3 de la sérotonine (5-HT₃) réduit les niveaux de dopamine dans les ganglions de la base, suggérant qu'il pourrait atténuer la libération de dopamine qui caractérise l'état dyskinétique. Ici, nous avons étudié les effets de l'ondansétron, un antagoniste hautement sélectif du récepteur 5-HT₃ à diminuer et à prévenir le développement des dyskinésies induites par L-DOPA chez le rat lésé à la 6-hydroxydopamine. Dans la première expérience, les rats sensibilisés avec L-DOPA pour induire des mouvements involontaires anormaux (AIMs), ont reçu L-DOPA en combinaison avec l'ondansétron ou un véhicule. Dans la seconde expérience, les doses efficaces d'ondansétron ont été administrées simultanément avec L-DOPA pendant 22 jours, et la sévérité des dyskinésies a été évaluée. Après 3 jours d'élimination, L-DOPA a été administré en aigu et la sévérité des dyskinésies évaluée. Nous avons trouvé que l'ondansétron 0,0001 mg/kg en combinaison avec L-DOPA, a significativement diminué la sévérité des dyskinésies par rapport à L-DOPA seul. Ondansétron 0,0001 mg/kg, administré en même temps que L-DOPA, a retardé le développement des dyskinésies. L'action anti-dyskinétique de l'ondansétron n'a pas compromis le bénéfice thérapeutique conféré par la L-DOPA. Ces résultats suggèrent que l'antagonisme des récepteurs 5-HT₃ est une stratégie thérapeutique potentiellement nouvelle et efficace pour soulager la sévérité et prévenir le développement des dyskinésies.

Mots-clés : maladie de Parkinson, dyskinésie, sérotonine, récepteur 5-HT₃, L-DOPA, 6-OHDA, rat

Abstract

L-3,4-dihydroxyphenylalanine (L-DOPA) is the most effective treatment for Parkinson's disease. However, with chronic administration of L-DOPA, patients develop motor complications such as dyskinesia. Previous studies have shown that 5-HT₃ receptor blockade reduces dopamine levels within the basal ganglia, suggesting that it could mitigate the aberrant dopamine release that characterises the dyskinetic state. Here, we investigated the effects of the highly-selective 5-HT₃ antagonist ondansetron at diminishing the expression of established, and preventing the development of L-DOPA-induced dyskinesia in the 6-hydroxydopamine-lesioned rat. In the first set of experiments, rats were primed with L-DOPA to induce abnormal involuntary movements (AIMs), after which L-DOPA was administered, in combination with ondansetron or vehicle. The effect of ondansetron on L-DOPA anti-parkinsonian action was subsequently determined by the cylinder test. In the second set of experiments, rats were administered effective doses of ondansetron, started concurrently with L-DOPA for 22 days, during which dyskinesia severity was monitored. After a 3-day washout period, an acute challenge of L-DOPA was administered and AIMs severity was assessed. We found that acute challenges of ondansetron 0.0001 mg/kg in combination with L-DOPA, significantly diminished the severity of AIMs compared to L-DOPA alone. Ondansetron 0.0001 mg/kg, when started concurrently with L-DOPA, attenuated the priming process leading to the development of dyskinesia. The anti-dyskinetic action of ondansetron did not compromise the therapeutic benefit conferred by L-DOPA. These results suggest that 5-HT₃ receptor antagonism is a potentially new and effective therapeutic strategy to alleviate the severity, and prevent the development of dyskinesia.

Keywords: Parkinson's disease, dyskinesia, serotonin, 5-HT₃ receptor, L-DOPA, 6-OHDA, rat

Table of content

Résumé.....	i
Abstract.....	ii
Table of content	iii
List of tables.....	vii
List of figures.....	viii
List of abbreviations	ix
Acknowledgements.....	xiii

I. Introduction..... 1

1. General Introduction.....	2
2. Parkinson's Disease	3
2.1. Epidemiology of Parkinson's Disease	3
2.2. Aetiology of Parkinson's Disease.....	4
2.3. Risk factors of Parkinson's disease.....	5
2.4. Pathophysiology of Parkinson's disease.....	14
2.5. Dopaminergic system in Parkinson's disease.....	17
2.6. Clinical features of Parkinson's disease	20
2.7. Current pharmacotherapy for Parkinson's disease	22
2.8. Surgical interventions for Parkinson's disease	25
3. L-DOPA induced dyskinesia	25
3.1. Clinical characteristics of dyskinesia.....	25
3.2. Timing of dyskinesia.....	26
3.3. Risk factors for the induction of dyskinesia	28
3.4. Risk factors for developing dyskinesia.....	30
3.5. Dyskinesia rating scales in Parkinson's disease	33
3.6. Pharmacological management of dyskinesia	34
3.7. Surgical options for dyskinesia.....	34
3.8. Basal ganglia circuitry in dyskinesia	35

3.9.	Dopaminergic system in dyskinesia.....	45
3.10.	Serotonergic system in dyskinesia.....	47
4.	5-HT ₃ receptor	52
4.1.	Localization of 5-HT ₃ receptors.....	53
4.2.	5-HT ₃ receptor subtypes and properties.....	54
4.3.	Physiology and pharmacology of 5-HT ₃ receptors.....	55
4.4.	5-HT ₃ receptors in Parkinson's Disease and L-DOPA-induced dyskinesia	57
5.	Animal models of Parkinson's disease	59
5.1.	The 6-OHDA-lesioned rat.....	60
5.1.1.	Injection of 6-OHDA into the MFB.....	61
5.1.2.	Compensation	61
6.	Behavioural testing	62
6.1.	Cylinder test.....	62
6.2.	ALO AIMS.....	63
7.	Objectives and hypotheses.....	65
II. Material and methods		67
	Animals.....	68
	Dose-finding pharmacokinetic study	68
	Unilateral 6-OHDA lesion	68
	Cylinder test.....	69
	Drug treatments.....	69
	Experimental design.....	70
	Acute challenges of ondansetron study.....	70
	<i>De novo</i> ondansetron study	70
	Ratings of AIMS.....	71
	Assessment of L-DOPA anti-parkinsonian action.....	72
	Perfusions.....	72
	LC-MS/MS analysis for dopamine and its metabolites	73
	Statistical Analysis.....	73
	Pharmacokinetic study	73

Cylinder test	74
Acute challenges of ondansetron study.....	74
<i>De novo</i> ondansetron study.....	74
III. Results	75
Pharmacokinetic profile of ondansetron	76
Extent of dopaminergic denervation assessed in the cylinder test.....	77
Acute challenges of ondansetron at 0.0001 mg/kg significantly alleviated the severity of established AIMs	78
Duration of axial AIMs	78
Duration of limbs AIMs.....	78
Duration of orolingual AIMs	79
Duration of AL AIMs	79
Duration of ALO AIMs	79
Amplitude of axial AIMs	82
Amplitude of limbs AIMs.....	82
Amplitude of orolingual AIMs	82
Amplitude of AL AIMs	83
Amplitude of ALO AIMs.....	83
<i>De novo</i> study	86
<i>De novo</i> treatment with ondansetron attenuates the development of L-DOPA-induced AIMs	86
<i>De novo</i> treatment with ondansetron attenuates the development of ALO AIMs.....	92
Administration of ondansetron does not impair the therapeutic efficacy of L-DOPA in the cylinder test.....	94
IV. Discussion.....	97
Limitations and future directions	98
Pharmacokinetic study of clinically relevant doses of ondansetron	99
5-HT ₃ blockade with ondansetron alleviates previously established AIMs without impairing the anti-parkinsonian efficacy of L-DOPA.....	100

Effect of ondansetron and ALO AIMs on L-DOPA anti-parkinsonian action	105
Effect of ondansetron on the development of ALO AIMs	105
V. Conclusion	108
VI. Bibliography	110
VII. Appendix.....	172

List of tables

Table I: Genes and loci associated with Parkinson’s Disease	9
Table II: Ondansetron pharmacokinetic parameters in the 6-OHDA-lesioned rat	76
Table III: Duration rating scale of ALO AIMs in the 6-OHDA-lesioned rat	I
Table IV: Amplitude rating scale of ALO AIMs in the 6-OHDA-lesioned rat	II
Table V: Glass delta’ of right forepaw use across ondansetron treatments	III
Table VI: Glass delta’ of both forepaw use across ondansetron treatments	IV

List of figures

Figure 1: Dopaminergic synapse	19
Figure 2: Schematic diagram of the classical BG circuitry describing different states	39
Figure 3: Schematic representation of the experimental design	71
Figure 4: Plasma levels of ondansetron in a preliminary pharmacokinetic study	76
Figure 5: Performance in the cylinder test in drug-naïve lesioned animals.....	77
Figure 6: Effect of acute challenges of ondansetron on the duration of established L-DOPA induced AIMs.....	81
Figure 7: Effect of acute ondansetron treatment on the amplitude of established L-DOPA induced AIMs.....	85
Figure 8: Time course of the development of AIMs during the 22-day priming phase of the <i>de novo</i> ondansetron study.....	90
Figure 9: Effect of ondansetron on the duration of cumulative and peak AIMs severity during an acute 6 mg/kg L-DOPA challenge	93
Figure 10: Use of forepaws across treatment conditions	96
Figure 11: Equation to calculate Glass' delta	V
Figure 12: Time course of the development of AIMs during the 22-day priming phase of the <i>de novo</i> ondansetron study.....	VII

List of abbreviations

α -synuclein	alpha-synuclein
2-DG	2-deoxyglucose
5,7-DHT	5,7-dihydroxytryptamine
5-HT	serotonin
5-HT _{1A}	serotonin 1A receptor
5-HT _{2A}	serotonin 2A receptor
5-HT ₃	serotonin 3 receptor
5-HIAA	5-hydroxyindoleacetic acid
5-HTP	5-hydroxytryptophan
6-OHDA	6-hydroxydopamine
8-OHDPAT	8-hydroxy-2-(di-n-propylamino) tetralin
AADC	aromatic acid decarboxylase
AIMs	abnormal involuntary movements
AIMS	Abnormal Involuntary Movement Scale
AL	axial limbs
ALO	axial limbs orolingual
ATP	adenosine triphosphate
ATP13A2	adenosine triphosphatase 13A2
BBB	blood brain barrier
BG	basal ganglia
cAMP	cyclic adenosine monophosphate
C _{max}	peak plasma concentration
CNS	central nervous system
Complex I	NADH:ubiquinone oxidoreductase
COMT	catechol-O-methyltransferase
DA	dopamine
DAT	dopamine transporter
DNA	deoxyribonucleic acid
DOPAC	3,4-dihydroxyphenylacetic acid
EIF4G1	eukaryotic translation initiation factor 4-gamma

ERK	extracellular signal-regulated kinase
EP	entopeduncular nucleus
FBXO7	F-box only protein 7
GABA	γ -amino butyric acid
GBA	glucocerebrosidase
GI	gastrointestinal
GPCR	G protein-coupled receptor
GWAS	genome-wide association studies
GPe	globus pallidus pars externa
GPi	globus pallidus pars interna
HD	Huntington's disease
HVA	homovanillic acid
L-DOPA	L-3,4-dihydroxyphenylalanine
LB	Lewy body
LC-MS/MS	liquid chromatography-tandem mass spectrometry
LN	Lewy neurite
LID	L-DOPA-induced dyskinesia
LRRK2	leucine-rich repeat kinase 2
MAO-B	monoamine oxidase-B
MAPT	microtubule-associated protein tau
mCPBG	1-(m-chlorophenyl)-biguanide
MFB	medial forebrain bundle
MPTP	1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine
MSN	medium spiny neuron
NMDA	<i>N</i> -methyl-D-aspartate
PD	Parkinson's disease
PINK1	PTEN-induced putative kinase 1
PK	pharmacokinetic
PLA2G6	phospholipase A2, group VI
PLTS	persistent low-threshold spiking
RBD	REM sleep behaviour disorder
REM	rapid eye movement

RN	raphe nucleus
RNA	ribonucleic acid
ROS	reactive oxygen species
SEM	standard error of the mean
SERT	serotonin transporter
SNARE	soluble <i>N</i> -ethylmaleimide-sensitive factor attachment protein receptor
SNc	substantia nigra pars compacta
SNCA	alpha-synuclein gene
SNr	substantia nigra pars reticulata
STN	subthalamic nucleus
$t_{1/2}$	plasma half-life
t_{max}	time at maximal plasma levels
TH	tyrosine hydroxylase
UDysRS	Unified Dyskinesia Rating Scale
VMAT2	vesicular monoamine transporter type 2
VPS35	vacuolar protein sorting 35
VTA	ventral tegmental area

“You can’t ever reach perfection, but you can believe in an asymptote toward which you are ceaselessly striving.”

-Paul Kalanithi

Acknowledgements

First and foremost, I thank my supervisor, Dr Philippe Huot, for giving me the opportunity to pursue my graduate studies in his research group and for going above and beyond in his duties as a supervisor. His extensive knowledge on movement disorders, attention to detail, open-ended questions and critique on my project have constantly furthered my scientific critical thinking skills and my development as a researcher. His welcoming attitude to questions and encouraging support and advice on scholarship submissions were also greatly appreciated. I am also grateful for Dr Huot's willingness to meet and have discussions, often with little notice, in spite of his busy schedule. I sincerely thank him for his feedback on the Thesis and his insightful comments that forced me to further my thoughts. Thank you for imparting some of your valuable knowledge and for your patience, work ethic and understanding.

I would also like to thank all the past and current members of the Huot lab, whom I have had the pleasure of working with, for their camaraderie and support really made my MSc journey a positive experience. I thank our lab manager, Dr Adjia Hamadjida, who was really a co-supervisor in terms of responsibilities, for his guidance throughout my project. Dr Hamadjida's expertise in various techniques, particularly in stereotaxic surgeries and behavioural testing, was instrumental in my training as a graduate student, especially when I first started. I thank Dominique Bédard for her help, as without it, the behavioural studies and pharmacokinetic study wouldn't have been possible. I thank Imane Frouni for being such a wonderful colleague and a compassionate friend and for assisting with some behavioural studies. We have shared countless discussions on methods and troubleshooting experiments, and life outside research, and even travelled to our first international conference together. I look forward to our future adventures! I thank Sébastien Belliveau for his help with the necropsy and I thank Élodie Bourgeois-Cayer for the French translation of my abstract. I really appreciated their great energy and infectious enthusiasm that created such a lively atmosphere in the lab. I thank Lamia Sid-Otmane for her insightful comments and thought-provoking discussions. I thank Vaidehi Nafade for all the lengthy scientific and non-scientific discussions and for making the long commute enjoyable and worthwhile. Moreover, I would like to thank all my friends who were so supportive and always willing to lend an ear whenever I needed it.

To my mentor Dr Michel Panisset, although our interactions were few, I am grateful for your invaluable advice and recommendations that allowed me to reflect and develop my ideas. I would also like to express my thanks to all of my collaborators, Dr Francis Beaudry and Ms Fleur Gaudette and Dr Lehka Sleno.

Last but not least, I am extremely grateful to my immediate family for their endless support and encouragement throughout this challenging but rewarding adventure! I thank my grandparents for their continued support in my pursuit of higher learning and for being so understanding of my occasional early/late hours. I thank my parents for their unconditional love, support and understanding, I know it wasn't easy being my on call chauffeur to the train station every day. I thank my brother Henry for his lively disposition and humour that has always left me in good spirits. I thank my sister Karina for sharing her knowledge on global and humanitarian issues, and for our deep philosophical discussions that are often hilarious and outrageous. I look forward to your journey of higher learning.

I. Introduction

1. General Introduction

Parkinson's Disease (PD) was initially described in *An Essay on Shaking Palsy* by the British physician James Parkinson in 1817 (1). However, it was only until 1861 that the French neurologist Jean-Martin Charcot, known as “the founder of modern neurology”, coined the term “Parkinson's disease” and distinguished bradykinesia as a separate clinical feature of the illness (2). PD is one of the most common neurodegenerative disorders that affects nearly 1% of the population over 65 years of age (3, 4). PD can be defined by four cardinal motor features: tremor at rest, akinesia (or bradykinesia), rigidity and postural instability (5). In addition to these symptoms, many patients are also affected by non-motor symptoms including dementia, autonomic dysfunction, and sleep disorders (6). The onset of PD is gradual and clinical manifestations do not appear until there is a loss of approximately 40-60% of the dopamine (DA) neurons in the substantia nigra (SN) pars compacta (SNc) and about 80% of striatal nerve terminals (7-10). In more severe stages PD, neuronal loss spreads to outside the SNc to regions including the locus coeruleus, raphe nuclei (RN), olfactory bulb and cerebral cortex, and this widespread neurodegeneration may be responsible for the progression of non-motor symptoms of PD (11) (12). The pathological hallmark of the disease is the presence of intracellular proteinaceous inclusions, known as Lewy bodies (LBs), however, the role of LBs in the pathogenesis of PD is still unknown (13). Alpha-synuclein (α -synuclein) is a major component of LBs and recent studies demonstrate that specific α -synuclein conformations are directly toxic to neurons (14, 15) and can propagate via a “prion-like” mechanism of pathogenesis (16).

Currently, the most effective symptomatic drug for PD is the biochemical precursor to DA, L-3,4-dihydroxyphenylalanine (L-DOPA), which helps to relieve motor symptoms by restoring striatal DA levels. However, as L-DOPA is also converted into DA in the peripheral nervous system, chronic L-DOPA therapy results in adverse effects, notably debilitating involuntary movements, termed L-DOPA induced dyskinesia (LID) (17). Moreover, the longer the duration of treatment, the greater the number of PD patients that develop LID and approximately 80 to 90% of patients suffer from LID after 10 years of treatment (18, 19). Patients with advanced PD suffer from these erratic movements that cannot be adequately controlled with existing therapies (20), which underscores the urgent need to develop therapies that attenuate dyskinesia. In recent years, the understanding of neuronal mechanisms that

underlie the pathophysiology of LID has grown and has been associated with events including the pulsatile stimulation of dopaminergic receptors, downstream changes in proteins and genes and abnormalities in non-dopaminergic transmitter systems, which modify the activity of the basal ganglia (BG) circuitry (21).

2. Parkinson's Disease

2.1. Epidemiology of Parkinson's Disease

Epidemiological studies show that PD is an age-related disease with men at higher risk of developing the disease than women (3). In Canada, it is estimated that about 99,000 individuals are living with PD and by 2031, the number of is expected to increase by 65% to 163,700 (22). In addition, Canadians with PD tend to be older individuals with an average age of diagnosis of 66.2 years of age (22).

The global prevalence PD is estimated at between 18 to 300 per 100,000 individuals while the incidence of PD is between 10 and 50 cases per 100,000 individuals per year, respectively (23, 24). Incidence of PD is heavily age-dependent, and onset is rare before 50 years of age until a sharp increase of incidence is observed after 60 years of age (25). The disease prevalence is estimated at 1% in subjects over 65 years of age and increases to 4.3% in those over the age of 85 (26). Most of the increase is attributed to the general trend of an increasingly ageing population (27). Despite the worldwide distribution of PD, incidence rates may vary among populations. The conflicting results between individuals studies may reflect differences in research methodologies, particularly in case definitions, diagnostic criteria, and the age distribution of the study population (28). A collaborative study in four European countries using similar case-finding methods and diagnostic criteria did not reveal any differences (29). In contrast, a meta-analysis of six studies found a lower prevalence in Africa than in Europe or North America (30), but no significant difference was reported between African Americans and Caucasians living in Mississippi (31). In addition, an autopsy study found that African Americans showed the same prevalence of incidental LB disease when compared with Caucasian populations (32). Similarly, lower prevalence rates have also been reported in some Asian countries (23, 33-35) but some studies have found similar estimates to Western countries (36, 37). Accordingly, differences in environmental exposure or interethnic distribution of

susceptibility genes may also contribute to the ethnic differences in estimates of PD prevalence and incidence (38, 39). Moreover, the variation in PD prevalence reported amongst studies may be related to differences in response rates, survival and case certainty rather than ethnic differences in PD prevalence (31, 38, 40, 41). Thus, the relative contribution of genetic or environmental variations to population differences in PD incidence is still unclear (26).

Some studies report a higher prevalence of PD in men than in women (42-45) with a male-to-female ratio of about 1.5 (46-48) but other studies found no significant differences in PD prevalence between men and women (29, 49, 50). Consistent with prevalence studies, prospective studies have reported a higher incidence of PD in men than in women (29, 42, 51-53). The neuroprotective effects of oestrogens in women and X-linked genetic factors may account for the higher risk of PD in men but their role is still controversial (54).

2.2. Aetiology of Parkinson's Disease

The aetiology of PD is poorly understood but considerable advances in sequencing technology, genetics (55) and clinical studies (20) have contributed to a greater comprehension on the pathogenic processes occurring in PD. The common view today is that PD is a multifactorial disease that arises from the combined effects of exposure to environmental risk factors, genetic susceptibility, and complex genetic-environmental interactions (26, 39). Ageing is the strongest risk factor of developing PD (56) and can be explained by the increasing failure of normal physiological and biochemical processes that lead to the increased vulnerability of DA neurons to toxic insult (57). Growing evidence suggests that impairments in the regulation of protein homeostasis including processes such as protein aggregation, intracellular protein and membrane trafficking and disruptions to the ubiquitin-proteasome and lysosome-autophagy are implicated in the pathogenesis of PD (58). In addition, it has been suggested that the genetics of PD are involved in aberrations in synaptic structure and function (58), which confirmed the importance of mitochondrial dysfunction in toxin models of PD (59).

2.3. Risk factors of Parkinson's disease

2.3.1. Non-genetic risk factors

2.3.1.1. Environmental hypothesis

Evidence linking exposure to agrochemicals, including pesticides and herbicides to an increased risk of PD has been postulated for many years (60, 61). In particular, it has been demonstrated that rotenone (62) and paraquat (63-65) cause nigral dopaminergic cell death in rodents. Furthermore, individuals exposed to pesticides had a 70% higher incidence of developing PD those not exposed (66). Additional environmental factors identified include industrialization, rural environment (67), use of well water (68), intake of various metals, etc. (69-71) but studies show conflicting outcomes (72). Although several studies report a positive association between environmental risk factors and PD, no factor has been consistently implicated as the sole causative agent (73). Similarly, the degree of pesticide exposure that may lead to PD is unknown.

2.3.1.2. Discovery of MPTP-induced Parkinson's disease

The discovery of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-induced parkinsonism (74) stimulated an interest in exposure to environmental risk factors for PD. MPTP causes the degeneration of nigrostriatal dopaminergic neurons with the loss of striatal dopamine in various species, including primates, cats and mice (75-80). In 1982, several young people developed an acute and severe form parkinsonism, due to MPTP produced during their illegal synthesis process of heroin (74, 81). MPTP is highly lipophilic, and can be metabolized into 1-methyl-4-phenylpyridinium (MPP⁺), the active toxic molecule, by brain monoamine oxidase B (MAO-B) (75-80, 82). MPP⁺ is accumulated by high affinity DA transporters (DAT) into the mitochondria of dopaminergic neurons (83). Once inside the mitochondria, MPP⁺ binds to and inhibits NADH-ubiquinone oxidoreductase I (complex I) of the mitochondrial electron transport chain (84). This results in an impairment of ATP production, elevated intracellular calcium concentration and free radical generation (81). Accumulation of MPP⁺ in dopaminergic neurons causes neurodegeneration via reactive oxygen species-mediated oxidative stress and results in DA neuron death (85-87). MPTP-induced parkinsonism exhibits similar phenomena to PD, particularly the preferential loss of DA nerve terminals of the putamen and DA neuron

loss of the SNc. Primates exposed to MPTP are responsive to L-DOPA treatment, and develop motor complications after chronic administration (88). Despite the contributions of the MPTP-lesioned animal models to the knowledge of pathways implicated in PD pathogenesis, they do not fully capture all the features of the disease. For example, MPTP-induced parkinsonism is not progressive, an acute rather than chronic increase in α -synuclein occurs and LB formation is absent (57, 64, 89). In addition, the investigation of agents based on MPTP models in clinical trials has not been successful thus far. Thus, the underlying pathways in the MPTP models of PD may not all be shared with those in PD patients (64).

A recent systematic review of meta-analyses identified several environmental factors that are associated with a risk of developing PD (90). Two factors, physical activity and constipation, presented with convincing evidence for a strong association with PD. Several cohort studies support the protective effect of physical activity for PD (91, 92). Also, constipation may be an early premorbid manifestation of PD (93) and laboratory studies and laboratory studies reported an abnormal deposition of α -synuclein within the submucosal and myenteric plexuses of the enteric nervous system (94, 95). Highly significant association for increased PD risk included head injury (96), anxiety or depression (97), while decreased risk is associated with smoking (97) and high uric acid levels (98). Additional non-genetic risk factors significantly associated with development of PD include a decrease in risk with alcohol (99) and coffee consumption (100), whereas pesticides (101), well water (97), and male sex (102, 103) are linked to an increase in risk. Although substantial evidence supports the association of environmental risk factors and PD, the heterogeneity amongst the examined meta-analyses suggests that some associations may reflect reverse causation, residual confounding, information bias and sponsor conflicts. In addition, the variation in cohort studies and case-control studies, differences in exposure assessment (frequency and exposure types) may account for the biased estimates of association. Furthermore, the authors emphasize that mechanisms of several putative risk factors are poorly understood, and additional studies are required to clarify the association between these factors and the risk of developing PD.

2.3.2. Genetic risk factors

Clinical observation of increased prevalence of PD amongst relatives of patients (104, 105) and the discovery of families with genetic forms of parkinsonism (106-109) heightened

interest in the heritability of PD. However, familial aggregation does not necessarily imply genetic causation (110, 111) and a large PD twin study found no increased concordance for PD amongst monozygotic twins (112). However, for subjects with an onset before 50 years of age, a significant concordance rate in identical twins was identified, which suggests young onset PD has a greater genetic component. In contrast, a later twin study using clinical assessment and fluoro-dopa positron emission tomography (PET) to image dopaminergic function reported increased concordance amongst monozygotic twins (113), which supports a role of genetics in PD aetiology.

Significant advances in understanding the pathogenic processes of PD in the past few decades have been made due to the identification of genetic mutations and chromosomal loci associated with parkinsonism (Table I, page 9) (114-116). The majority of PD cases are sporadic (117) but Mendelian loci and the high-risk glucocerebrosidase (GBA) locus collectively account for approximately 10 – 40% of disease risk depending on the population under study (114). Genetic factors that have been identified include mutations in the genes for α -synuclein (SNCA), and leucine-rich repeat kinase 2 (LRRK2), which are responsible for autosomal-dominant PD forms, whereas mutations in the genes PARK2 (Parkin), PARK7 (PTEN-induced putative kinase 1, PINK1), PARK7 (DJ-1), and PARK9 (ATPase 13A2, ATP13A2) account for early-onset autosomal recessive PD forms (117). Recently, two other autosomal dominant PD genes, vacuolar protein sorting 35 (VPS35) and eukaryotic translation initiation factor 4-gamma (EIF4G1), have been identified (118-120). Early candidate gene studies and subsequent meta-analyses provided conclusive evidence demonstrating that polymorphisms in SNCA (121), LRRK2 (122), microtubule-associated protein tau (MAPT) (123) and glucocerebrosidase (GBA) (124) impact PD susceptibility.

High-density arrays of single nucleotide polymorphisms identified genetic susceptibility factors in genome-wide association studies (GWAS), where the frequencies of putative risk alleles are compared in patients and controls (125). Genetic variations may be susceptibility factors or disease modifiers, affecting penetrance, age at onset, severity, and progression (126). The most commonly studied candidate genes include genes involved in DA metabolism, mitochondrial metabolism, detoxification, other neurodegenerative diseases and familial PD (3, 127) and genes associated with putative risk factors for PD including oestrogen receptor gene polymorphisms (128), the tau HI haplotype (129) and the apolipoprotein E epsilon 2 allele (130).

Although the significance of many loci identified with an increase in PD risk is still unknown, they account for a population attributable risk of > 60% (131).

2.3.2.1. Risk Loci

2.3.2.1.1. GBA

The GBA gene encodes a lysosomal enzyme β -glucocerebrosidase, which is involved in glycolipid metabolism (116). Homozygous GBA mutations cause Gaucher's disease, an autosomal recessive lysosomal storage disease caused by accumulation of glucocerebrosides (132). In contrast, heterozygous mutations in GBA are associated with a higher risk of PD where approximately 5-10% of PD patients have GBA mutations as opposed to an estimated frequency of 1% in healthy controls but this may be underestimated in certain populations (133-137). Carriers of only one mutated allele have a 5-fold increased risk to develop PD compared with non-carriers, which makes GBA one of the strongest genetic risk factors reported to date (134). The high prevalence of PD amongst GBA mutation carriers, which is also age-dependent and rises up to 30% at 80 years of age (138), has led to the suggestion that GBA mutations can act as dominant factors with reduced penetrance rather than simple risk variants (139). The mechanism underlying the association of mutations in GBA with the development of PD and other LB disorders (140) is not known, but may be caused by alterations in lipid metabolism or autophagy and lysosomal function (115). In fact, the mechanism of pathogenicity may be linked to α -synuclein as intracellular glucocerebrosides facilitate the aggregation of α -synuclein into toxic oligomers and fibrils (141), which are the main constituent of LBs (142). Moreover, α -synuclein is primarily degraded through autophagy and GBA mutations interfere with autophagic clearance of α -synuclein fibrils (143, 144). Consequently, fibrils are likely to accumulate in the cell (145, 146), following which they may propagate through cell to cell transmission (147, 148).

Table I: Genes and loci associated with Parkinson's Disease

locus	gene	inheritance	clinical phenotype	pathology
Mendelian Genes				
PARK1 and PARK4	SNCA	AD	parkinsonism with common dementia	LBs
PARK2	Parkin	AR	early-onset, slowly progressing parkinsonism	LBs rarely
PARK3	unknown	AD	late-onset parkinsonism	LBs
PARK5	UCH-L1	AD	late-onset parkinsonism	unknown
PARK6	PINK1	AR	early-onset, slowly progressing parkinsonism	one case with LBs
PARK7	DJ-1	AR	early-onset parkinsonism	unknown
PARK8	LRRK2	AD	late-onset parkinsonism	usually LBs; sometimes tangles or neither
PARK9	ATP13A2	AR	early-onset parkinsonism with Kufor-Rakeb syndrome	unknown
PARK10	unknown	AD	unclear	unknown
PARK11	unknown	AD	late-onset Parkinsonism	unknown
PARK12	unknown	unknown	unclear	unknown
PARK13	HTRA2	unknown	unclear	unknown
PARK 14	PLA2G6	AR	aggressive and complex parkinsonism with pyramidal features	LBs
High-risk locus				
Gaucher's locus	GBA		late-onset parkinsonism	LBs
	SNCA		typical PD	LBs
Low-risk loci				
	LRRK2		typical PD	LBs
FTDP-17	MAPT		dementia, sometimes parkinsonism	neurofibrillary tangles

Adapted from (114-116).

2.3.2.1.2. MAPT

Most of the gene loci discovered through GWAS are present in more than 5% of the population (allele frequencies of > 10%) and carriers of the risk allele have a less than two-fold increase of disease risk over the general population average (114). The majority of these low-risk loci appear to mediate their effect by altering gene expression rather than through translational changes. The MAPT is a protein that can form aggregates similar to α -synuclein and beta-amyloid. Mutations in the MAPT gene cause a range of neurodegenerative phenotypes but some can lead to a typical PD presentation (149). The H1 haplotype at the MAPT locus has been consistently suggested as a risk factor for PD (150, 151) and gene duplications at MAPT cause frontotemporal dementia (152), which suggests that the pathogenic cascades in the tauopathies can provoke severe neurodegeneration leading to parkinsonism (153, 154). Moreover, MAPT promotes the formation of α -synuclein oligomers and fibrils (115) and in transgenic mice that exhibit the LB variant of Alzheimer's disease, cognitive decline is accelerated and associated with amyloid beta, tau and α -synuclein pathologies compared to age-matched control animals (155). Thus, synergistic interactions between α -synuclein and tau may promote their fibrillization and the formation of pathological inclusions characteristic of neurodegenerative diseases.

2.3.2.2. Autosomal dominant forms of Parkinson's disease

2.3.2.2.1. LRRK2

The most common cause of autosomal-dominant Mendelian form of PD is mutations in the LRRK2 at the PARK8 locus, which account for nearly 10% of all familial dominant inherited forms (156). The G2019S kinase domain mutation is the most frequent LRRK2 mutation (157), and responsible for 5-40% of sporadic or dominantly inherited PD, depending on the population studied (158-160). Higher G2019S prevalence rates have been reported in more isolated populations, such as the Ashkenazi Jewish (161) and North African Berber Arab (162) populations, which can be explained by a genetic founder effect (163). Patients with LRRK2 mutations tend to display late-onset PD with symptoms indistinguishable from those of sporadic PD, even though LB pathology is sometimes absent or lacking (164, 165). Thus, the disconnect between clinical manifestations of PD and the presence of LBs (166, 167) supports the theory

that inclusions may not be necessary for neurodegeneration and may instead be a consequence of PD (115). The mechanism underlying the neurodegeneration caused by LRRK2 mutations and its natural substrate are still unknown. However, cell culture studies suggest that neurotoxicity *in vitro* requires intact kinase activity (168, 169), prompting increased interest towards LRRK2 kinase inhibitors (170) as a potential neuroprotective strategy.

2.3.2.2.2. α -synuclein

SNCA mutations are the second most common cause of dominant PD (171) and various studies have reported a link between familial PD and duplications or triplications in the SNCA gene (172). The SNCA gene encodes α -synuclein, which accumulates in LBs predominantly within the brainstem. As LB pathology is also the dominant pathology observed in most cases of LRRK2-related PD, this suggests that SNCA and LRRK2 affect a common pathway that leads to α -synuclein aggregation (139). Moreover, gene triplication leads to earlier onset and faster progression of disease than duplication, which suggests a gene-dose effect between α -synuclein levels and disease severity (173). The link between α -synuclein expression levels and the appearance of PD is well-established across studies, and leads to the hypothesis that a gain-of-function by α -synuclein underlies pathogenesis of PD (115). In addition, recent *in vivo* evidence shows that it binds to and promotes assembly of soluble *N*-ethylmaleimide-sensitive factor attachment protein receptor (SNARE) complexes, which are required for the fusion of vesicles to the pre-synaptic membrane (174, 175). Triple knock-out mice lacking α -synuclein also exhibit deficits in SNARE complex assembly and develop accelerated age-associated motor impairments and early-onset mortality, but do not show neurodegeneration (168, 174).

2.3.2.2.3. VPS35

VPS35 gene encodes a major component of the retromer complex involved in endosomal trafficking to the trans Golgi (119, 176). Recent studies have identified a single missense (D620N) mutation in VPS35 as a new cause of autosomal-dominant PD in two independent exome sequencing studies on Swiss (118) and Austrian families (119). Frequency of mutation carriers is low and has been estimated to represent about 0.1% of the PD population (177). Patients with a VPS35 mutation exhibit classical late-onset, L-DOPA responsive parkinsonism similar to that of sporadic PD, with a slightly earlier age at onset (139). Specific deletion

of VPS35 in DA neurons of mice results in PD-like deficits, including loss of DA neurons and accumulation of α -synuclein and early degeneration at 2-3 months of age (178). Consistent with this data, overexpression of human D620N VPS35 variants induce the marked degeneration of SNc DA neurons and axonal pathology (179). In addition, mutations in VPS35 caused extensive mitochondrial fragmentation and cell death as well as functional deficits *in vitro*, in mouse SNc neurons *in vivo*, and in human fibroblasts from PD patient bearing the D620N mutation (180). Defects in macroautophagy, aminomethylphosphonic acid (AMPA) receptor trafficking to dendritic spines or alterations in mitochondrial dynamics and turnover have been proposed as the mechanism underlying VPS35-induced neurodegeneration (181). Although the mode of action by which it causes PD is unclear, modulation of the development of DA neurons via the wingless-related integration site pathway (182, 183) and aberrant brain iron accumulation (184, 185) have been suggested as possible mechanisms. Furthermore, recent studies demonstrate that VPS35 may interact with other PD-linked gene products including LRRK2, SNCA and Parkin (186-190) in a common pathway that leads to the neurodegeneration observed in PD.

2.3.2.3. Autosomal recessive forms of Parkinson's disease

2.3.2.3.1. Parkin, PINK1, DJ-1

Mutations in Parkin are the most common cause of autosomal recessive forms of PD, whereas mutations in PINK1 and DJ-1 are relatively less prevalent (139). Parkin gene mutations account for almost 50% of early-onset recessive familial PD and up to 15% of early onset sporadic cases (191, 192). Pathology underlying Parkin-related PD does not tend to show LBs, unlike the autosomal dominant and idiopathic forms. Clinical manifestation of Parkin mutations is often indistinguishable from that of the sporadic disease except for the earlier age at onset (generally before 45 years of age) (139). Wild-type Parkin, PINK1 and DJ-1 are involved in processes of mitochondrial quality control and regulation such as mitogenesis, mitophagy and mitochondrial homeostasis and transport (139, 163). Studies suggest that the function of these proteins in a mitochondrial quality control pathway is impaired in PD, leading to the accumulation of bioenergetically compromised mitochondria, however, it is unclear how this might give rise to substantial nigral degeneration and PD (193).

2.3.2.3.2. ATP13A2, FBXO7 and PLA2G6

More rarely, recessively inherited forms of atypical parkinsonism are caused by mutations in the ATP13A2, F-box only protein 7 (FBXO7) and phospholipase A2, group VI (PLA2G6) genes (131). Mutations in ATP13A2 were first identified from families with Kufor-Rakeb syndrome, a rare hereditary disease with typical signs of PD that also includes symptoms of more extensive neurodegeneration (194). Loss of function mutations of ATP13A2 underlie an autosomal recessive form of early-onset parkinsonism with pyramidal degeneration and dementia (194) while heterozygous mutations may be a risk factor for PD (195). ATP13A2 mutations likely play a role in lysosome degradation (131) and recent studies demonstrate that ATP13A2 can rescue against α -synuclein toxicity in a yeast, *Caenorhabditis elegans* (*C. elegans*) and neuronal culture model of PD (196). FBXO7 mutations cause early-onset autosomal recessive parkinsonism with pyramidal signs and after an initial favourable response to L-DOPA, patients often develop dyskinesia (197). Most of the reported FBXO7 mutations are loss of function (198) but no neuropathology has been described thus far. Mutations in PLA2G6 cause an early-onset recessive degenerative disorder characterized by spasticity, ataxia and dystonia but adult onset forms can manifest as dystonia-predominant parkinsonism (199) that is responsive to L-DOPA (200). PD associated with PLA2G6 is caused by the homozygous or compound heterozygous inheritance of various missense mutations (201-203).

The clinical and sometimes pathological resemblance of genetic PD to sporadic disease make it a suitable human model to identify at-risk individuals in earlier and possibly prodromal phases of the disease (116). However, monogenic causes of PD represent less than 10% of PD cases in most populations (204) whereas the majority of cases seem to arise from complex interactions among genes and between genes and environmental factors (39). Thus, environmental factors appear to be more important determinants than ethnic and genetic factors in the aetiology of PD. Further efforts are warranted to understand how genetic causes and risk factors of PD play a role in the underlying pathophysiology in hopes of developing targeted therapies that alter disease course (139).

2.4. Pathophysiology of Parkinson's disease

Based on autopsy findings in PD patients, Braak and colleagues postulated that α -synuclein aggregates form in the periphery in early stages of PD before α -synuclein aggregation in the brain (205) and also propose a six-stage system for PD based on the stereotypic pattern of α -synuclein spreading (206). The Braak model is based on the presence of LBs and Lewy neurites (LNs) where the pathogenic process begins in the lower brainstem in the dorsal motor nucleus of the vagus nerve and anterior olfactory structures (206). The disease then spreads rostrally from the dorsal motor nucleus of the vagus nerve through the medulla, pontine tegmentum, midbrain and basal forebrain before ultimately reaching the cerebral cortex. This process follows a specific pattern where susceptible regions are affected in a predictable topographic sequence (207) where severity of the lesions and the clinical manifestations of the disease increase as the pathology ascends from the brainstem (208). Accordingly, *in vitro* (209, 210), *in vivo* (211, 212) and clinical evidence (213, 214) suggest that cell types in the central nervous system (CNS) exhibit a propensity for developing Lewy pathology that shares common features. In spite of the support for Braak's hypothesis, there is criticism around whether it accurately reflects the development of PD in all patients as studies report that Braak staging fails to describe the disease progression in upwards of 50% of α -synuclein immunoreactive cases (215-217). Moreover, the absence of information on the loss of neurons and synaptic connections in the original Braak papers has been the subject of scrutiny (218, 219) as the scientific premises underlying the model remain unclear (220). Thus, the inconsistencies between the Braak model and conflicting reports of the spread of pathology require further study to determine the relationship and likely also require a deeper understanding of the mechanisms underlying the role of α -synuclein in disease progression (207).

Several lines of evidence have implicated dysfunctions in the ubiquitin-proteasome system in PD pathogenesis (221-223), which have been further supported by the identification of disease-causing mutations in genes encoding proteins involved in protein degradation in PD (224). Impairment of ubiquitination pathways and proteasomal function could result in defects in the clearance of toxic aggregates and result in their accumulation and degeneration of DA neurons (223, 225). Although systemic administration of proteasomal inhibitors modelled a behavioural and pathological phenotype reminiscent of PD (226), this model has been met with

great scrutiny due to the extensive variability in the consequences of *in vivo* proteasomal inhibition (227). Moreover, questions on the molecular connections between these systems and pathogenesis of PD remain, including the divergent fate of misfolded proteins for degradation or inclusion formation, and further studies, that likely exploit advances in genetics and technologies (228), are warranted to clarify this relationship.

2.4.1. Lewy bodies in Parkinson's Disease

Idiopathic or sporadic PD is characterized by the selective loss of neurons and appearance of abnormal cytoplasmic proteinaceous aggregates called LBs in the soma or LNs in the processes in DA neurons (173). Immunohistochemistry shows that LBs consist primarily of the protein α -synuclein (229), as well as other proteins such as ubiquitin (230) and parkin co-regulated gene (231). Studies have suggested that misfolded α -synuclein and the deposition of LBs within midbrain neurons could contribute to neuronal damage and cell death (232). In addition to SNc DA neurons, neuronal loss and Lewy pathology also occurs extensively in locus coeruleus noradrenergic neurons, RN serotonergic neurons, enteric DA neurons, post-ganglionic sympathetic noradrenergic neurons and olfactory neurons (11, 12).

2.4.2. Alpha-synuclein in Parkinson's disease

α -synuclein is of the main constituents of LBs and LNs, and accumulates widely in central and peripheral neurons of PD patients (233). Given it predominates in pre-synaptic terminals and the nuclear envelope, it plays a role in SNARE-mediated exocytosis and synaptic vesicle transport (234). Moreover, α -synuclein is present in mitochondria in PD brain and may affect mitochondrial function both *in vitro* and *in vivo*, possibly leading to the vulnerability of nigrostriatal DA neurons in PD (235-238). Accordingly, it has been demonstrated that α -synuclein aggregation may be associated with oxidative or nitrosative stress (239-241), which may be important in the pathogenesis in neurodegenerative disorders with LBs, like PD (242). Converging evidence also supports the hypothesis that α -synuclein oligomers (243, 244) and fibrils (245, 246), the pathologic form of α -synuclein, may participate in the propagation of neurodegeneration observed in PD (247). Thus, the misfolded α -synuclein fibrils present in LBs (142) and their self-propagation and spread, reminiscent of a "prion-like" process, suggest that their mode of cell-to-cell transmission is not in disagreement with the Braak staging and a

possible peripheral origin of Lewy pathology (210, 211, 248-250). Furthermore, the prion hypothesis of α -synuclein transmission is supported by evidence from transgenic mouse models of synucleinopathy and viral vector-mediated α -synuclein overexpression in rats (251, 252) and nonhuman primates (253, 254). However, no data from transgenic mice models has reported the spontaneous pathological α -synuclein (255) and conflicting findings surround the specificity of peripheral α -synuclein for PD in humans (255), for instance, some studies have found similar levels of α -synuclein accumulation in the colon of patients with PD compared with healthy controls (256-258).

2.4.3. Oxidative stress in Parkinson's disease

Oxidative stress defines a disequilibrium between the levels of reactive oxygen species (ROS) produced and the ability of a biological system to detoxify the reactive intermediates, ultimately creating a perilous state contributing to cellular damage (259). An increasing body of evidence suggests that in PD, oxidative stress and mitochondrial damage contribute to a sequence of events that lead to the degeneration of DA neurons in the SNc (57, 260-262). In addition to mitochondrial dysfunction, DA metabolism (263), neuroinflammation (259), iron (264), calcium (265) and ageing (266) also contribute to ROS production in the PD brain. Indeed, *post-mortem* brain analyses consistently show increased oxidative damage to lipids (267, 268), proteins (269), deoxyribonucleic acid (DNA) and ribonucleic acid (RNA) (270, 271). Further support for the link between oxidative stress and DA neuronal degeneration has been demonstrated by modelling motor symptoms of PD in toxin-induced animal models that cause oxidative stress such as 6-hydroxydopamine (6-OHDA), MPTP, rotenone and paraquat (272). It has also been suggested that mechanisms that contribute to neurodegeneration act in a feed forward manner where primary insults lead to oxidative stress, which damages key cellular proteins and disrupts lipid membranes that in turn cause more ROS production (259).

2.4.4. Mitochondrial dysfunction in Parkinson's Disease

In addition to the dual role of mitochondria as both a source and target of ROS (273-275), compelling evidence suggests that mitochondrial dysregulation is critical in the pathogenesis of PD (259). Mitochondria are dynamic organelles with important functions in cellular respiration, energy metabolism, calcium homeostasis, stress response and apoptosis

pathways (260). Several groups have reported decreased complex I activity in the SN of PD patients (276-278), and the finding of the downregulation of genes encoding mitochondrial proteins further supports the role of mitochondrial dysfunction in PD (279). In addition, PD-related proteins, including DJ-1, PINK1, Parkin, α -synuclein and LRRK2, are also involved in mitochondria quality control leading to exacerbations of ROS generation and susceptibility to oxidative damage (259).

2.5. Dopaminergic system in Parkinson's disease

DA neurons form four major systems within the mammalian brain: the nigrostriatal, mesolimbic, mesocortical and tuberoinfundibular systems (280) that originate from the A9, A10, and A8 groups of dopamine-containing cells, respectively (281, 282). In the nigrostriatal pathway, projections from dopaminergic neurons with cell bodies in the SN terminate in the striatum (283). The mesolimbic pathway consists of dopaminergic neurons that originate in the ventral tegmental area (VTA) and project to the nucleus accumbens and related limbic regions, whereas VTA neurons that project to the prefrontal cortex establish the mesocortical pathway (284). Last, the tuberohypophyseal pathway consists of dopaminergic projections from the hypothalamus to the pituitary gland, and its secretions regulate prolactin (285). Due to their wide connectivity within distinct pathways, DA neurons exert a variety of functions including locomotion, addiction, reward, learning and memory, cognition, stress and movement (286).

DA is a monoamine neurotransmitter synthesized in a series of enzymatic reactions (287), beginning with the conversion of the amino acid tyrosine into L-DOPA via the rate-limiting enzyme tyrosine hydroxylase (TH) (Figure 1, page 19). L-DOPA is subsequently decarboxylated into DA by the enzyme aromatic acid decarboxylase (AADC). DA is then packaged into pre-synaptic vesicles by the vesicular monoamine transporter type 2 (VMAT2) and released at nerve terminals into the synapse upon stimulation. Released DA bind to DA receptors to elicit a response in the post-synaptic cell and this interaction is important in the modulation of motor function through the BG circuitry. Extracellular DA is either metabolized by MAO-B and catechol-O-methyl transferase (COMT) in the cytosol or transported back into the pre-synaptic terminal via the DAT. Following re-entry of DA into the pre-synaptic neuron, DA can be repackaged into vesicles and recycled or degraded into the metabolite homovanillic acid (HVA).

A deficit in the number of nigrostriatal dopaminergic neurons, characteristic of PD, disrupts the dopaminergic transmission and produces abnormal motor features in affected subjects.

2.5.1. Nigrostriatal dopaminergic pathway

Dopaminergic terminals in the striatum consist of dense innervation of fibres from two specific groups of neurons in the brainstem (282, 288). The first group are neurons with cell bodies in the VTA that project to the nucleus accumbens and olfactory tubercle. The second group have cell bodies in the SNc and project primarily to the putamen and the caudate nucleus. The tegmental and nigral afferents form the nigrostriatal dopaminergic pathways.

2.5.2. Classification of dopamine receptors and their distribution in the basal ganglia

DA receptors are a family of G protein-coupled receptors (GPCRs) with five subtypes, D1-D5, that are divided into two groups (289). D1-like receptors are comprised of D1 and D5 receptors and mainly couple G proteins (290), which stimulate adenylyl cyclase and cyclic adenosine monophosphate (cAMP) production (291, 292). In contrast, D2-like receptors comprise D2, D3 and D4 receptors; they couple with $G_{\alpha i}$ / $G_{\alpha o}$ and inhibit adenylyl cyclase and negatively regulate cAMP production (293, 294). D1-like receptors have an excitatory effect by stimulating cAMP production, whereas activation of D2-like receptors is inhibitory. D2 receptors are the presynaptic receptors of the dopaminergic system and are responsible for the negative feedback on levels of synaptic DA (295).

Both D1Rs and D2Rs are highly expressed by striatal medium spiny neurons (MSNs) (296, 297) and are present at lower levels in the cortex as compared to the striatum (298). D1 receptors are expressed in striatonigral neurons containing substance P and dynorphin that project to the SN pars reticulata (SNr) and to the globus pallidus (GP) pars interna (GPi), which constitute the direct striatal output pathway (299). In contrast, D2 receptors are predominantly localized in striatofugal neurons expressing enkephalin, which project to the GP pars externa (GPe), constituting the indirect pathway (300, 301). D1 receptors are post-synaptic whereas D2 receptors are also localized on pre-synaptic nigrostriatal dopaminergic terminals, on SNc neurons, and on pre-synaptic cortico-striatal terminals where they can inhibit striatal glutamate

release (296, 302, 303). In humans and nonhuman primates, D3 receptors are mostly found in the nucleus accumbens and caudate nucleus-putamen complex but are also localized in the GPI, anterior thalamus, amygdala, hippocampus and cortex (304-306). In the human striatum, there is approximately D3:D2 receptors is approximately 1:2, and D3 receptors can co-localize with both D1 and D2 receptors (305, 307).

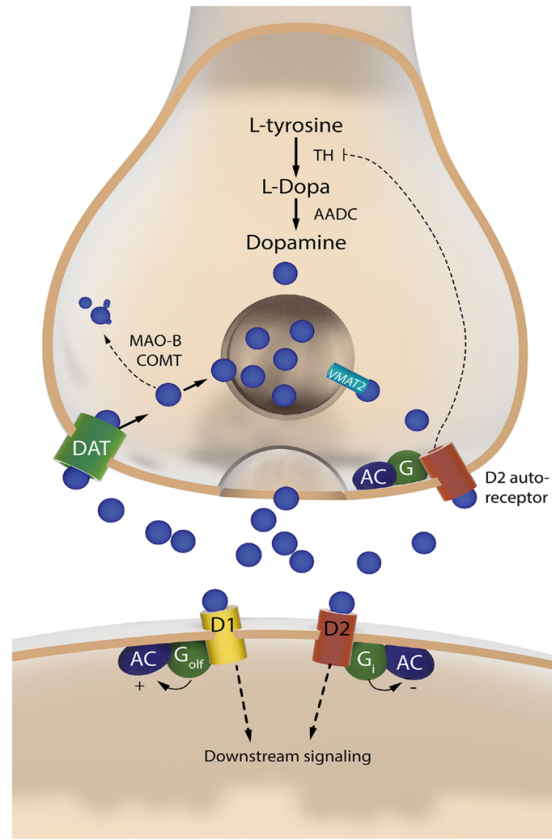


Figure 1: Dopaminergic synapse. After DA is synthesized in the pre-synaptic neuron and released into the nerve terminal, extracellular levels of DA are regulated through several mechanisms. DAT is responsible for the reuptake of DA back into the pre-synaptic neuron, VMAT2 packages DA back into synaptic vesicles, pre-synaptic D2 receptors control DA synthesis and release, and MAO-B and COMT are involved in the extracellular metabolism of DA. Following the release of DA at synaptic terminals, DA can bind to two types of DA receptors on post-synaptic neurons. The D1 receptor is coupled to Golf and activates cAMP-dependent signalling pathways while the D2 receptor is coupled to Gi and inhibits the same pathways. AADC: aromatic L-amino acid decarboxylase; AC: adenylate cyclase; COMT: catechol-o-methyl-transferase; DAT: dopamine transporter; MAO-B: monoamine oxidase B; TH: tyrosine hydroxylase; VMAT2: vesicular monoamine transporter 2. Modified from (308).

2.6. Clinical features of Parkinson's disease

Traditionally described as a motor disorder, numerous brain structures are affected at different time points along the course of the disease manifestation, and both motor and nonmotor symptoms are observed in PD.

2.6.1. Motor symptoms of Parkinson's disease

The four cardinal motor features of PD are the following: bradykinesia, muscular rigidity, resting tremor and impairment of postural balance leading to disturbances in gait and falls. Movement can be normal in early disease (309) due to the redundancy in BG activity and the capacity of the striatum to compensate functionally for lower degrees of DA deficiency (7). However, after the loss of approximately 80% striatal DA and loss 60% SNc DA neurons, motor symptoms begin to appear (7-10). Initially, the symptoms are mild and are usually confined to one side of the body but over the disease course, symptoms are increasingly impairing and involve the contralateral side as well (310). Motor features are heterogeneous and in spite of the lack of consensus on the classification of subtypes, empirical clinical observations suggest the existence of two major subtypes: tremor-dominant PD with a relative absence of other motor symptoms and non-tremor-dominant PD, a phenotype described as akinetic-rigid syndrome and postural instability gait disorder (311). Bradykinesia is defined as difficulty in planning, initiating and executing movements, as well as with performing sequential and simultaneous tasks (312). This often manifests as deficits in fine motor control (312), slower reaction times(313, 314) and slowness in performing daily activities (315-318). Moreover, bradykinesia leads to impairment of the power of voluntary movement (319, 320). Muscle rigidity is described as increased resistance to passive joint movement, and can often lead to a flexed posture (321). The combination of these motor symptoms along with disturbances in gait, leads to greater disability in PD patients. In addition to motor symptoms, PD patients also experience debilitating nonmotor symptoms.

2.6.2. Non-motor symptoms of Parkinson's disease

The spectrum of non-motor features encompasses olfactory dysfunction, sleep disturbances, autonomic dysfunction, gastrointestinal (GI) distress, memory loss and dementia, as well as neuropsychiatric conditions (6, 322-324) (325, 326). Virtually all patients with PD

exhibit at least one nonmotor symptom with an average of 7.8–11.9 nonmotor symptoms per patient (322, 327-330). Moreover, nonmotor symptoms have been reported to affect the quality of life of PD patients to a greater extent than motor features (330-332).

Olfactory dysfunction is one of the most common nonmotor symptoms in PD that affects over 80% of patients with PD (333), a prevalence with greater sensitivity for PD compared to many other clinical markers. Although robust evidence indicates that olfactory loss precedes PD (334-340), the lead time for olfactory loss is variable, and some patients may develop detectable loss before or after developing parkinsonism (309). In addition, strong evidence supports rapid eye movement (REM) sleep behaviour disorder (RBD) as a predictor of synucleinopathies, most commonly PD with or without dementia LBs (309). RBD is defined as apparent enactment of dreams during REM sleep, associated with a loss of normal REM sleep atonia (341). Five prospective studies have reported that synucleinopathies, *i.e.* PD, dementia with LBs or multiple system atrophy develop in up to 80-90% of patients with RBD (342-347). Moreover, one study showed that α -synuclein deposition was present in brains of 98% of subjects with neurodegenerative disease who also had RBD confirmed by polysomnography (348).

Additional symptoms relating to autonomic dysfunction include constipation, orthostatic hypotension, urinary and sexual dysfunction and somnolence. α -synuclein is abundant in the GI system (256), which led some to speculate that prion-like spreading might occur from the GI tract to the brain (207), and a delay in colon transit time that results in constipation could theoretically facilitate this spreading (309). Although this remains controversial, pathological evidence supports the capacity of α -synuclein to spread (349) and a preliminary report showed that vagotomy may reduce the risk of PD (350). Depression and anxiety are commonly comorbid in PD, however, their potential as a marker is limited by the low relative risks (351-355) and predictive value (309), and highly variable lead times for psychiatric manifestations (353-356). Cognitive impairment is traditionally associated to late stages of PD but mild cognitive changes are also observed in *de novo* PD (357). In general, nonmotor features precede the appearance of motor symptoms in PD, including olfactory dysfunction, RBD, constipation, urinary and sexual dysfunction, which show promise as potential markers of prodromal PD (358).

2.7. Current pharmacotherapy for Parkinson's disease

Currently available therapies are not disease-modifying or neuroprotective and they provide only symptomatic relief for motor features of the disease (359). There are non-pharmacological, pharmacological and surgical treatments that attempt to restore dopaminergic activity using L-DOPA and DA receptor agonists.

2.7.1. L-DOPA

Administration of L-DOPA with a peripheral AADC inhibitor such as carbidopa or benserazide is the most effective treatment for relief of motor symptoms of parkinsonism, particularly for controlling bradykinesia (360). The addition of carbidopa or benserazide enhances the therapeutic benefits of L-DOPA, reduces the dose of L-DOPA required, and minimize peripheral adverse effects (361).

2.7.1.1. Pharmacology of L-DOPA

DA was first synthesized in 1910 (362) and its biochemical precursor was synthesized the following year (363). In a seminal study, Ehringer and Hornykiewicz discovered that DA levels were reduced in the striatum of PD patients (364). Moreover, Hornykiewicz observed a correlation between most of the PD motor symptoms and striatal DA depletion (365). The introduction of DA replacement therapy with L-DOPA in the early 1960s revolutionized symptomatic treatment of PD (366). Unlike DA, L-DOPA crosses the blood brain barrier (BBB), and is effective at alleviating motor features of PD during early stages of treatment (367). Barbeau and colleagues reported an improvement of parkinsonism, mostly with respect to rigidity, after oral administration of L-DOPA to patients with PD (368). Furthermore, Cotzais and colleagues reported that high doses of L-DOPA had marked beneficial effect on the motor symptoms of parkinsonism, mostly bradykinesia and rigidity (369-371).

As AADC is also present outside of the brain, peripheral metabolism of L-DOPA can cause adverse side effects including hypotension, nausea and vomiting (372, 373). Thus, a peripheral acting AADC inhibitor, such as carbidopa or benserazide, that does not cross the BBB is often co-administered, to limit the peripheral decarboxylation of L-DOPA so that more L-DOPA is available to enter the brain, while minimizing the aforementioned peripheral adverse

effects. The addition of peripherally acting AADC inhibitors allowed a reduction in the required dose of L-DOPA up to 60-80% (374, 375), potentiated its efficacy, led to a faster onset of anti-parkinsonian benefit and a reduction of cardiovascular and GI side effects (371, 373, 376).

L-DOPA is absorbed in the duodenum and proximal jejunum by active transport via the large neutral amino acid system (377, 378). L-DOPA enters the body and brain by active transport (379, 380) and competes with dietary proteins and amino acids (381). Thus, high protein intake can reduce L-DOPA anti-parkinsonian action and indeed, intraduodenal delivery of L-DOPA leads to a decline of motor performance following oral protein intake (382). In the clinic, L-DOPA has a short half-life of 1.5 to 2 hours (383-388). After oral administration, L-DOPA plasma levels reach a maximum about one hour after intake, although this may vary due to the unpredictable absorption (387-389). L-DOPA plasma levels are 10–15-fold higher than L-DOPA levels in the ventricular cerebrospinal fluid (390). COMT inhibitors are often used to extend the duration of L-DOPA anti-parkinsonian action and indeed, tolcapone, entacapone and opicapone increase the area under the curve when administered with L-DOPA (391, 392).

L-DOPA is converted into DA by AADC in DA neurons from the SN and projections (393), serotonin (5-HT) neurons from the raphe complex and their striatal projections (394, 395) and noradrenergic neurons from the locus coeruleus and their projections (396). Due to the characteristic degeneration of the nigrostriatal system in PD, L-DOPA is converted in DA mostly by raphe-striatal 5-HT neurons, and to a lesser extent, by striatal intrinsic AADC-containing interneurons (397-399).

Although L-DOPA is the most effective treatment for PD, chronic administration of L-DOPA is associated with the development of motor complications including motor fluctuations and LIDs (400). LID represent a major limitation of current pharmacotherapy for PD as the majority of patients experience dyskinesia after a few years of treatment (400, 401) and underscores the need to develop effective therapeutic strategies for patients suffering from these involuntary movements.

2.7.2. Dopamine agonists

DA receptor agonists such as ropinirole, pramipexole and rotigotine are commonly employed and their main advantages over L-DOPA are: they do not require enzymatic activation

and have a longer duration of action (402). However, due to their action on DA receptors, their adverse effect profile includes: hallucinations, confusion, nausea, and increased incidence of impulse control disorders including pathological gambling and hypersexuality (403). Apomorphine, a DA agonist, is primarily used as rescue therapy for temporary relief of off-periods of akinesia in patients with fluctuating response to dopaminergic therapy (404), but is not available in Canada.

2.7.3. MAO-B inhibitors

Selective MAO-B inhibitors like selegiline and rasagiline delay the breakdown of DA in the striatum (287). As their efficacy is modest, they can be used as a monotherapy in early PD and in advanced stages of the disease, they can be administered as an adjunct to reduce off time in patients with declining response to L-DOPA (405).

2.7.4. COMT inhibitors

COMT inhibitors block the peripheral degradation of LDOPA, which leads to its increased half-life and enhanced central bioavailability (406). Two COMT inhibitors are available in Canada, tolcapone and entacapone, while opicapone is also available in Europe, although entacapone is preferred because it is not associated with hepatotoxicity (407-409). They are used as adjunctive treatment in patients who develop motor fluctuations to prolong the effect of L-DOPA (410).

2.7.5. Anti-cholinergics and amantadine

Anti-cholinergic agents like trihexyphenidyl and benztropine were historically used for the treatment of PD before the introduction of L-DOPA. Their main therapeutic effect is on tremor and they are only indicated in early PD or as an adjunct to DA replacement therapy. Amantadine modulates dopaminergic and cholinergic transmissions and is also a non-competitive *N*-methyl-D-aspartate (NMDA) receptor antagonist with a modest efficacy and improves parkinsonian symptoms in mildly affected patients with early disease and reduces LIDs in patients with advanced disease.

2.7.6. Treatments for non-motor symptoms

Non-motor symptoms in PD are being increasingly recognized as important issues diminishing the quality of life of patients, although treatment options remain inadequate (6, 326). Thus, medications used to treat related conditions are usually tried in PD, *e.g.* antidepressants for depression, atypical antipsychotics for psychosis, laxatives for constipation, etc. However, several of the currently used interventions lack robust evidence and require further research to discern their role in the management of nonmotor symptoms of PD } (326).

2.8. Surgical interventions for Parkinson's disease

Surgical brain treatments are increasingly attractive options for patients with PD, particularly in advanced stages because they diminish motor fluctuations and decrease dyskinesia severity (411). Deep brain stimulation (DBS) is the most popular surgical intervention and aims to modulate abnormal neuronal activity within a circuit to alleviate symptoms (412). Several clinical trials have demonstrated that stimulation of the subthalamic nucleus (STN) or GPi is effective in moderate to severe cases of PD (413). Inasmuch as its invasiveness, this surgical intervention is considered a symptomatic treatment limited to patients with advanced PD who no longer achieve adequate symptomatic relief with medication (411). As a result, patients who undergo DBS are on average 58.6 years of age with an average disease duration of 12 years (414-417). The precise mechanism(s) underlying the action of DBS is still unclear but the commonly accepted hypothesis is that electrical stimulation of the brain exerts inhibitory effects on structures such as neuron cell bodies close to the current and the output nuclei of the BG (418, 419).

3. L-DOPA induced dyskinesia

3.1. Clinical characteristics of dyskinesia

Dyskinesia, originates from the Greek word (δυσκίνησία) meaning “bad movement” and is medically defined as any nonvoluntary movement, and dyskinesia is clinically heterogeneous in presentation and progression (420). Typically, it firsts appear on the more severely parkinsonian side of the body (421) and affects the distal lower limb, followed by an ascending spread but with disease progression, both sides are ultimately affected (422).

LID develops with disease progression and with repeated DA replacement therapy in PD. Although administration of DA agonists can elicit dyskinesia, the prevalence is lower than L-DOPA monotherapy (423, 424). Thus, in attempt to minimize the induction and delay the onset of drug-induced dystonia and chorea, DA agonists may be used in early disease. Alternatively, once fluctuations and dyskinesia appear, DA agonists may allow to reduce the dose of L-DOPA to reduce existing dyskinesia in PD (420). In contrast, a concern with the addition of COMT inhibitors, which delay the breakdown of L-DOPA, is that L-DOPA will have a longer duration of action and may induce dyskinesia (425).

3.2. Timing of dyskinesia

LID expression is based on the timing of appearance in relation to the on-off phenomenon of the patient (420), which is defined as the switch between mobility and immobility in patients treated with L-DOPA (426). On-time refers to periods when the patient is responsive to L-DOPA and experiences improvement in mobility, whereas off-time applies to periods when the patient responds poorly to L-DOPA and impaired motor function including tremor, akinesia or rigidity, which often occurs as an end-of-dose or “wearing off” of the effect of L-DOPA, or because L-DOPA has not been taken (420). In particular, the on period is the most common time during which LID is present, *i.e.* in 70-80% of patients who experience it while being in the on-state (427). During the on period, dopaminergic stimulation in the patient’s brain is maximal or increased. Moreover, LID may be categorized into different presentation forms where the most common forms are peak-dose dyskinesia, off-period dyskinesia and diphasic dyskinesia. These forms of LID likely lie somewhere along a clinical continuum and may vary between patients, and even fluctuate between different doses in the same patients. In general, LID appears first in the foot, ipsilaterally to the side of the body more severely affected by parkinsonian symptoms, with inversion of the foot and ankle. A possible explanation may be the early loss of dopaminergic innervation in the dorsolateral striatum, which corresponds somatotopically to the foot area innervated by the SNc (428). Over the course of disease progression, dyskinesia eventually spreads to other body areas, and may follow the pattern of progression of parkinsonian symptoms (429). In addition, the forms of dyskinesia are not mutually exclusive, and a combination of choreic and dystonic movements, and the three types of dyskinesia may be observed in patients throughout the L-DOPA cycle (430).

3.2.1. Peak dose dyskinesia

Peak-dose LID is the most common subtype that occurs at high plasma levels of L-DOPA and coincides with the maximal anti-parkinsonian effect. These are often expressed as spasmodic twitching or jerking in the muscles of the superior extremities and neck. They are predominantly characterized by a choreic phenotype but can also include dystonic movements (431). They tend to be less disabling and less painful than the other forms of dyskinesia.

3.2.2. Off-period dyskinesia

In contrast, off-period dyskinesia occurs when the DA levels are falling and dopaminergic stimulation in the brain is low. Thus, the patient is subsequently in the off state or in the transition from the on to off state. This phenomenon tends to occur more commonly during the night, or prior to the first L-DOPA dose in morning or just after taking the dose (432). Phenotypically, off-period dyskinesia mainly consists of dystonic movements (433) frequently affecting the foot of the more affected side, but can also be segmental or generalized in distribution (431). A characteristic manifestation includes foot inversion and painful flexion of the toes (421). This phenomenon can be combated by taking more dopaminergic medication, particularly longer acting DA agonists and controlled release L-DOPA to avoid a decrease in DA levels over night(434).

3.2.3. Diphasic dyskinesia

Diphasic dyskinesia, also described as dystonia-improvement-dystonia (D-I-D), occurs at two different time points of a single dose cycle, at the beginning and at the end of the treatment effect, separated by an on period of minimal dyskinesia (435). In general, it is assumed at the time points of dyskinesia, L-DOPA levels are rising and falling, respectively. They affect both extremities but tend to affect the lower limbs of the most affected side more (436). It is characterized by repetitive and rapidly alternating dystonia and consists of flexion/extension of the foot or stereotyped movements (437). A notable feature is that while the lower limbs are moving involuntarily, the upper body can exhibit parkinsonian symptoms such as tremor (438).

3.3. Risk factors for the induction of dyskinesia

3.3.1. Priming

Priming is defined as the neurochemical and functional aberrant modifications in the DA-denervated BG that eventually lead to the emergence of dyskinesia in response to the repeated administration of L-DOPA or DA agonists (439). Over time, with repeated treatment, the chance of dopaminergic stimulation eliciting LID is increased and once LID has been established, the severity of dyskinesia increases (440).

Priming is produced by a two-step administration of dopaminergic drugs, including DA agonists and L-DOPA, and, encompasses an induction and expression phase. During the induction phase, the first administration of dopaminergic drugs, followed by the second administration, results in priming, *i.e.* the neurochemical and behavioural sensitization of the animal to subsequent challenge with dopaminergic drugs (441). At the behavioural level, chronic dopaminergic treatment induces dyskinesia that, once established, never stops and progressively increases in severity with further treatment (442). In the expression phase, once LID is established, the brain maintains the primed state such that even in the absence of treatments, a single challenge with L-DOPA or DA agonist will elicit dyskinesia at nearly the same severity (440). In addition, priming is associated with neurochemical maladaptive modifications in the DA-denervated striatum similar to those observed in animal models of dyskinesia induced by chronic DA replacement therapy (443). These include changes in the production of cAMP, phosphorylation of phosphoprotein of 32 kDa, and expression of mRNA encoding immediate early genes, dynorphin and glutamic acid decarboxylase isoform 67, which all regulate the activity of striatal output neurons (443).

The major factors that affect the induction of dyskinetic movements are the extent of nigral dopaminergic cell loss, the type of drug administered, and the method of drug administration (444). Nigral cell degeneration is responsible for plastic changes in BG function that lead to dyskinesia priming (308). The extent of denervation regulates the level and duration of drug exposure required to induce dyskinesia. In general, normal nonhuman and human primates do not develop dyskinesia when treated with chronic pharmacological doses of L-DOPA (445). In contrast, humans or primates exposed to MPTP, which induces nigral cell loss,

and PD patients with severe nigral denervation develop dyskinesia rapidly after starting L-DOPA therapy (446).

The method of drug delivery is regarded as one factor that is important in the development of dyskinesia, in addition to other determinants including nigrostriatal dopamine denervation and dopamine receptor sensitivity. The greater propensity of L-DOPA to induce dyskinesia than DA agonists is attributed to more than differing duration of action and/or plasma half-life and pharmacology of receptors (447). Orally administered L-DOPA has a short half-life of 60-90 minutes (448) and due to its central and peripheral pharmacokinetic effects amongst many other factors, it is associated with a gradually shorter action duration with continued disease progression (449). In contrast, orally administered DA agonists have half-lives of several hours or more, notably cabergoline has a half-life of 72-96 hours (450, 451). Therefore, intermittent oral doses of L-DOPA with a shorter duration of action may be associated with pulsatile stimulation of dopaminergic receptors, whereas administration of longer-acting DA agonists results in the tonic and phasic release of DA and more continuous physiological dopaminergic stimulation, which results in a lower incidence of dyskinesia (452, 453).

The different pharmacological profile of L-DOPA compared to DA agonists may also underlie variations in dyskinesia induction. Orally-administered DA agonists display a higher affinity for D2 DA receptors compared to D1 receptors (454, 455), whereas L-DOPA interacts with all five subtypes of DA receptors present in the basal ganglia (456, 457). Evidence suggests that dyskinesia is associated with specific alterations in D1 receptor function and D2-selective DA agonists may induce less dyskinesia because of this pharmacological difference (458). In addition, several DA agonists also demonstrate affinity for non-dopaminergic receptors, for example, cabergoline also antagonizes 5-HT_{2A} receptors (451), which has been demonstrated to exert an anti-dyskinetic effect (459, 460). Thus, the relatively lower association of dyskinesia with DA agonists than L-DOPA may be due to their lack of selectivity for the D1 receptor and the broader pharmacology of L-DOPA (450).

Moreover, results from clinical trials have been conflicting on the association between continuous delivery of L-DOPA, such as intraduodenal L-DOPA infusion, also known as duodopa, and the risk of dyskinesia development (461). In two small trials, duodopa administration over a 12-h period daily over 6 months but not over a 14-h period daily over 18 months, reduced the severity of dyskinesia (462, 463). In contrast, a recent study reported no

effect of duodopa on dyskinesia severity (464). In addition, this method of continuous drug delivery still encompasses some pulsatility of post-synaptic dopaminergic receptors, and in the event of uninterrupted drug administration, there is the issue of tolerance (461). Therefore, while pulsatile L-DOPA may be implicated in the development of dyskinesia, compared to other factors, the extent of its contribution remains relatively unclear.

3.4.Risk factors for developing dyskinesia

Processes that underlie the induction of dyskinesia are different from those responsible for the execution of involuntary movements in response to subsequent drug treatment (439). The risk factors for LID include the age of PD onset, duration of L-DOPA treatment and dose, which suggests that the progressive loss of DA neurons and L-DOPA exposure are implicated in the development of LID (465-467). Cotzias and colleagues, who are credited with the first successful use of L-DOPA to treat PD, were also the first to report the incidence of LID (371). Studies subsequently noted the high incidence and variation of LID, and treatment-limiting effect of L-DOPA. Moreover, several studies reported that continuous rather than intermittent exposure to L-DOPA is associated with a decrease in the incidence of LID. Accordingly, continuous infusion of L-DOPA via an intestinal gel increased on time without dyskinesia compared to immediate-release oral formulation of L-DOPA (464).

3.4.1. Duration of L-DOPA therapy

The duration of L-DOPA therapy is considered an important risk factor for the development of LID. After five years of L-DOPA treatment, about 50% of patients are reported to have developed LID (423, 468, 469) and after 15 years, the incidence rises to more than 90% of patients suffering from dyskinesia (19). LID occurs more frequently with longer duration of L-DOPA treatment (470). However, it remains to be determined to what extent each of treatment duration, the effect of neurodegeneration, disease duration and dose contribute to the development and expression of LID (420). Thus, disease duration and severity, are also shown to correlate highly with the duration of therapy as well as the prevalence of dyskinesia.

3.4.2. Impact of L-DOPA dose

In general, L-DOPA does not typically induce dyskinesia in normal individuals (471, 472). In hemiparkinsonian rats, the dosage is critically involved in dyskinesia via loss of synaptic depotentiation (473). That L-DOPA induces dyskinesia and alleviates extrapyramidal symptoms is generally considered as a continuous dose-dependent pharmacological spectrum (474). A landmark paper by Ahlskog and Muentner looked at the epidemiology of LID in studies from the pre-L-DOPA era and from the modern era (468). The frequency of LID between prospective clinical trials and observational studies yielded similar figures and the overall frequency of LID applies to both. However, dyskinesia occurred earlier in the pre-L-DOPA era during L-DOPA treatment than the modern era, which is partially attributed to the longer durations of pre-existing PD. Moreover, the reported difference in LID frequency suggests that greater depletion of dopaminergic striatal terminals may increase the likelihood of developing LID. Similarly, LID occurs later now than in earlier series of studies, which may be explained by the higher dose used in the past, which led to the earlier appearance of dyskinesia (475). A large retrospective study found that the risk of developing LID was a higher initial L-DOPA dose (476), which is in agreement with a later cross-sectional study, adjusted for other risk factors (476). In line with this data, a placebo-controlled clinical trial also reported that patients with LID were taking higher L-DOPA doses at the appearance of dyskinesia compared to patients without dyskinesia (477). However, the cumulative dose of L-DOPA does not differ significantly between the groups, as observed in a retrospective study (478). Furthermore, it is difficult to determine the exact effect of L-DOPA dosage as PD patients may receive different doses at different time points throughout the day as the drug regimen changes throughout the course of disease (420).

Studies suggest that females are more likely to develop dyskinesia than males (479, 480) but it appears that sex cannot fully explain this phenomenon. In fact, given the same dose of L-DOPA with an AADC inhibitor, females are exposed to a higher plasma concentration time curve compared to males, when adjusted for kilogram of body weight (481). It seems that body weight affects the pharmacokinetics of L-DOPA, which in turn, may influence the onset of dyskinesia (480, 482). This is further supported by a subanalysis of a prospective clinical trial, which reported that dose per kilogram body weight is the most significant factor in the

development of LID (483). In contrast, multiple logistic regression analysis found that female gender, absolute dose, body weight and disease duration were insignificant (483). Thus, treatment with high L-DOPA dose relative to patient's body weight seems to be a significant risk factor for LID (484).

3.4.3. Impact of L-DOPA type

L-DOPA is currently available in several formulations including oral standard-release, controlled-release and dispersible tablets. Controlled-release formulations theoretically reduce fluctuations in plasma L-DOPA levels and clinical trials have reported decreased off-time and reduced L-DOPA dosing frequency when compared to immediate-release formulations (485-487). Other studies, however, suggest that controlled-release is not superior to standard release (488) and is associated with increased incidence of dyskinesia (488-492). To date, there is no consensus in selecting one formulation over another in the treatment of dyskinesia. An increasingly popular approach to treat motor complications in advanced PD patients is continuous intrajejunal infusion. Despite its cost and technical demands, this method of delivery improves motor fluctuations in PD and may reduce both duration and severity of dyskinesia (493, 494).

3.4.4. Young age at onset

Several epidemiological studies indicate that a young age at the onset of PD is a significant risk factor for LID (495). In addition, the "DA turnover" to DA synthesis and storage rate are inversely correlated with the onset age of PD (496). Young-onset patients might have more compensatory mechanisms to dopaminergic cell loss in the BG, rendering them more vulnerable to the development of LID (497). Moreover, young-onset patients have more nigral abnormalities while late onset patients have more cortical abnormalities (498). The relationship between the age of onset and the development of LID may be partially explained by genetic influences (499). Some genetically determined forms of PD at young onset have been reported to have a higher risk of developing LID, and a higher prevalence of dyskinesia has been reported in patients with a family history of PD (495, 500-503) than those without (495, 502). Mutations in the genes PARK 2, PARK 6 and PARK 7 are associated with young onset PD, and have been reported to have relatively higher rates of dyskinesia (504-506). Recent studies, however,

disagree with these reports and observe that carriers of PARK2 or PARK8 mutations do not develop LID more frequently than age- and disease duration-matched non-carriers (507, 508).

3.4.5. Genetic risk factors

Genetic factors may also contribute to the variability in incidence, severity and latency from treatment onset, and result in different susceptibility to develop dyskinesia. In addition, the interindividual difference and the high prevalence of LID in young-onset PD patients further supports the possibility of genetic susceptibility for LID (484). Genetic polymorphisms of pre-synaptic and post-synaptic structures could be potential substrates for genetic susceptibility to LID (484) and the occurrence of dyskinesia is associated with specific polymorphisms for the DA receptors or DAT genes (509-511), the COMT gene and the mu-opioid receptor gene.

3.5. Dyskinesia rating scales in Parkinson's disease

Despite the developments in pharmacological and surgical treatments for advanced PD, progress has been limited by the lack of a widely accepted clinical rating scale for dyskinesia (512). The challenge can be attributed to the variability in the anatomical distribution of dyskinesia, intensity of movements, disability or impact on daily living (513). In addition, quantification of dyskinesia using rating scales is subject to inter- and intra-rater reliability and also needs to clearly discriminate from other motor parkinsonian features. Amongst the clinically-available scales used to assess dyskinesia, only a few meet the minimal criteria and it is often at the discretion of the investigators and clinicians to select one that best fits the need of the assessment. Lang-Fahn (514) and PD-DYS-26 (515) are more patient-oriented scales while more objective assessments can be obtained for impairment and disability with the Abnormal Involuntary Movement Scale (AIMS) (516) and the Rush Dyskinesia Rating Scale (517). Although the AIMS displays high inter-rater reliability for tardive dyskinesia, data suggests it does not have a specific reference to LID, which limits its use in PD patients (518). In contrast, the Unified Dyskinesia Rating Scale (UDysRS), the newest rating scale developed specifically for the assessment of dyskinesia in PD, encompasses both patient-based and rater-based ratings of disability and impairment, and provides a more comprehensive measurement tool for the burden of dyskinesia (519). Furthermore, the intra-rater and inter-rater reliabilities and reproducibility of the UDysRS have been well established with a strong clinimetric profile (519-521).

3.6. Pharmacological management of dyskinesia

Pharmacological treatment for dyskinesia is based on adjustments to the intervals and doses of dopaminergic treatments, adjunct oral drugs with direct anti-dyskinetic effects and continuous administration of anti-parkinsonian drugs via pumps. To date, only two orally administered agents have been shown to relieve dyskinesia without worsening motor disability, amantadine and clozapine.

Amantadine is a non-selective NMDA receptor antagonist and has been reported to reduce LID in animal models and in PD patients (522). The effectiveness of this agent in LID has provided support for the pathogenic role of changes in striatal NMDA receptors in dyskinesia (522). In addition, a randomized, placebo-controlled trial reported that treatment with amantadine significantly improved on time with dyskinesia and reduced AIMs after a L-DOPA challenge (523). Furthermore, in a placebo-controlled crossover trial, dyskinesia severity was reduced by nearly 50% on amantadine compared with placebo (523). Amantadine is efficacious as an oral anti-dyskinetic drug with a sustained effect that lasts for at least one year (524). However, its potential as a treatment is limited by its ability to worsen neuropsychiatric problems, particularly in elderly patients (525).

Clozapine is an atypical anti-psychotic and although its exact mechanism of action is unclear, it may be attributed to its affinity to 5-HT_{2A} (5-HT_{2A}) receptors (526). Uncontrolled studies of clozapine for dyskinesia have estimated a reduction by around 50 % with high doses (527). Moreover, a randomized, placebo-controlled 10-week study reported that clozapine is associated with a significant reduction in on-time with dyskinesia compared with placebo, without changes in off-time duration (528). The use of clozapine is limited by potential adverse events, agranulocytosis and myocarditis, although evidence shows that this risk decreases over time (529, 530).

3.7. Surgical options for dyskinesia

Treatment of LID is one of the most common indications for neurosurgery in PD and procedures such as STN-DBS, or pallidotomy or GPi-DBS, which are thought to have a direct effect on dyskinesia, can provide relief of motor symptoms of PD and help control LID (531). Since the introduction of “chronic” high-frequency DBS by the French neurosurgeon Alim

Benabid (532), DBS procedures have been pivotal to improve motor symptoms (533), whereas ablative surgery is considered an alternative and only used when DBS is not feasible (415, 534, 535). However, controversial issues including the timing of therapeutic intervention, the selection of stimulation target and adverse effects (536) as well as the restricted selection criteria and invasiveness of these procedures present considerable risks to patients with PD compared to pharmacological interventions. Thus, the present Thesis will focus more on the relevance of pharmacological management of LID but advances in DBS and pallidotomy are reviewed in (412, 444).

3.8. Basal ganglia circuitry in dyskinesia

The interaction between DA and post-synaptic receptors is crucial in the modulation of motor function and classically described by the BG circuitry. BG are a group of interconnected nuclei located bilaterally in the diencephalon and midbrain and contain the striatum, GPe, GPI, SNc, SNr and STN (537). The BG form a neural network that relays information from the motor cortex to the thalamus, forming closed “cortico-striato-thalamo-cortical” loops (538). In addition to an involvement in the planning, initiation, and execution of voluntary movement (539, 540), these structures have broader roles in motor learning, executive functions, behaviours and emotions (541, 542). Thus, these loops functionally convey information for both motor and non-motor processes (543).

Clinical-pathological observations during the 20th century found that lesions to the putamen, GP and STN were associated with movements disorders (544, 545), which heavily influenced our understanding of BG function (546). There are two opposing views on the anatomical substrate of information processing at the BG, the “parallel processing” and “information funneling” hypotheses (547). The “parallel processing” hypothesis infers that the processing of different types of cortical information is largely independent via parallel and segregated circuits (547). Five parallel circuits have been identified thus far: motor, oculomotor, orbitofrontal, dorsolateral prefrontal, lateral orbitofrontal and anterior cingulate loops (543). Each circuit appears to receive inputs from separate cortical areas, travel portions of the BG and thalamus and project back upon cortical input areas, forming a partially “closed” loop (543, 548-550). In contrast, the “information funneling” hypothesis proposes the convergence of cortical information along the cortico-striato-pallido/nigro-thalamo-cortical system, and emphasizes the

contrast between the three-dimensional geometry of the axonal and dendritic arborizations in the striato-pallido-nigral circuit (547).

3.8.1. Physiological state

The BG and related nuclei can be categorized into input, output and intrinsic nuclei (551). Input nuclei receive incoming information from different sources, primarily from cortical, thalamic and nigral areas. The primary input structure of the BG is the striatum, which receives excitatory input from the cortex. In contrast, output nuclei send basal ganglia information to the thalamus and consist of the GPi and the SNr. Finally, intrinsic nuclei such as the GPe, the STN and the SNc are the intermediary between the input and output nuclei in the relay of information.

3.8.1.1. Structures

3.8.1.1.1. Striatum

The striatum serves as the major input structure of the BG and the origin of its name refers to the striated appearance of white corticofugal fibres (552). The caudate nucleus and putamen are referred to as the striatum (553). In primates, the fibres of the internal capsule separate the caudate nucleus medially and the putamen laterally (554), whereas in rodents, the structures are fused together (555). There is also a ventral component of the striatum defined as the nucleus accumbens, which is functionally and anatomically connected to limbic brain structures (556). Functionally, the caudate-putamen complex is associated with movement regulation, whereas the ventral striatum is involved in mediating neurological functions relating to motivation, reward and emotion (557, 558). The striatum receives different afferent projections, including dopaminergic fibres from the midbrain (559), serotonergic fibres from the dorsal and medial raphe nucleus (560) and noradrenergic fibres from the locus coeruleus (561). Glutamatergic fibres originating from the cerebral cortex project to the striatum in a somatotopically-organized manner (562-564). The targets of cortical input are gamma-aminobutyric acid (GABA)-containing inhibitory MSNs (565, 566) that represent the majority of striatal neurons. These GABAergic output cells are homogeneously distributed such that the striatum lacks a distinct cytoarchitectural organization, in contrast with the laminar organization of the cortex (555). MSNs are divided into two subtypes, which form the direct and indirect

pathways (567). The remaining striatal neurons are larger in size and serve as local interneurons composed of three subtypes: 2% contain either parvalbumin or calretinin and 1% contain the peptides somatostatin and acetylcholine (568, 569). Striatal interneurons are distributed in axons and the majority synapse onto spiny projection neurons (569).

3.8.1.1.2. Substantia nigra

The SN is a midbrain structure that comprises two distinct components, the SNr and the SNc. The name “substantia nigra”, also referred to as “locus niger”, or black substance, refers to the high concentration of neuromelanin, a dark pigment derived from L-tyrosine, found in dopaminergic neurons (570). In contrast, the SNr, consists of dendritic arborizations, and is located ventrally to the SNc (571). Functionally, the SNr is a major output structure of the BG circuitry involved in sensorimotor integration and organization of behaviour (572). Dopaminergic neurons of the SNc project to the caudate nucleus and putamen, where they synapse with MSNs and release dopamine (557, 573).

3.8.1.1.3. Subthalamic nucleus

The STN is the uniquely placed in the BG circuitry as the sole structure emitting glutamatergic fibres (299). It receives excitatory glutamatergic afferents from the frontal lobes with especially large inputs from the motor cortex (574, 575). The STN also receives inhibitory GABAergic input from the GPe (576). In contrast, the STN sends excitatory glutamatergic output to the GPi, GPe and SNr (577-580).

3.8.1.1.4. Output nuclei

The output nuclei, the GPi/SNr complex, act as a single functional unit through inhibitory projections to the thalamus, which in turn, influences frontal lobe cortical regions (299). The GPi and SNr have similar cyto- and chemoarchitectural characteristics and to some extent, similar types of afferent and efferent systems (546). They receive afferent projections including glutamatergic afferents from the STN and inhibitory GABAergic afferents from the striatum and GPe (299). In both structures, there is a relative segregation of limbic and sensorimotor inputs (581). The GPi and SNr consist of inhibitory GABAergic neurons with a

firing rate of discharge that tonically inhibits their targets (546). Their output predominantly projects to the ventral anterior, ventral lateral and mediodorsal thalamus (543, 582, 583).

3.8.1.2. Direct versus indirect pathways

To mediate voluntary movement, equilibrium is maintained between two opposing pathways originating in the striatum and regulated by the SNc DA projections that synapse with striatal GABAergic neurons (299).

The first population of MSNs express D1 receptors and preproenkephalin-B, an opioid peptide cleaved to produce co-transmitters including substance P and dynorphin (584). This population forms the direct pathway, and provide direct inputs to the output neurons of the GPi and to the SNr (Figure 20A, page 39). These structures then project to the ventral lateral and centro-median parts of the motor thalamus, which in turn, project towards the motor cortex. Striatofugal neurons subsequently send GABAergic projections inhibiting the tonic activity of GPi/SNr, which leads to a dis-inhibition of thalamic glutamatergic neurons. The thalamus then sends an excitatory glutamatergic projection that activates the motor cortex. The behavioural result is a facilitation of voluntary movement (585).

In contrast, the second population of MSNs express enkephalin and D2 receptors (584). These neurons project towards relay structures prior to arriving on the GPi/SNr and form the indirect pathway (Figure 2A, page 39). D2 neurons of the striatum project to the GPe, which in turn, sends GABAergic efferent fibres to the STN. The STN then provides excitatory projections to the output neurons of the BG. Striatopallidal MSNs subsequently emit an inhibitory GABAergic projection to the GPe, and further to the glutamatergic neurons of the STN. In turn, the STN activates the GPi/SNr. Both nuclei send an inhibitory GABAergic projection to the motor thalamus (ventral lateral and centro-medial nuclei) that leads to a decrease in excitatory thalamic output to the motor cortex. The net result is suppression of voluntary movement (585).

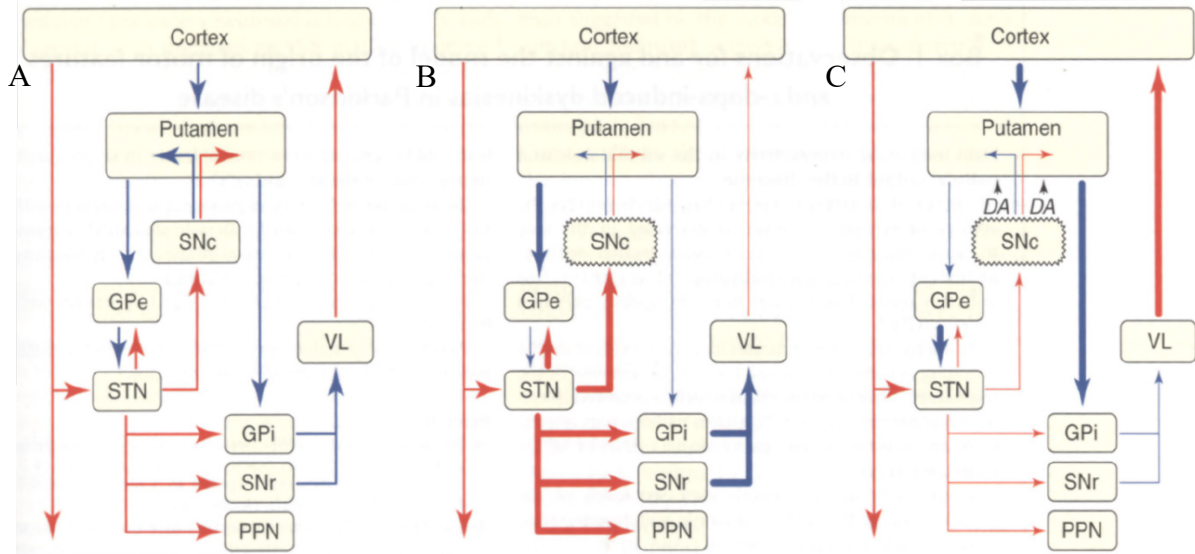


Figure 2: Schematic diagram of the classical BG circuitry describing different states. A. The model under physiological conditions shows an equilibrium between the direct and indirect pathways. B. The loss of nigrostriatal DA neurons in PD leads to the hyperactivity of the indirect pathway and the underactivity of the direct pathway. C. Chronic administration of L-DOPA and the eventual development of LID may be due to the hyperactivity of the direct pathway and underactivity of the indirect pathway. Reproduced from Obeso (586).

3.8.2. Pathophysiological state: parkinsonism and dyskinesia

3.8.2.1. Basal ganglia circuitry in Parkinson's Disease

In PD, the extensive degeneration of SNc dopaminergic neurons disrupts the equilibrium between the direct and indirect pathways and favours the hypokinetic state (7). Activity of the striatofugal neurons of the direct pathway diminishes, whereas the MSNs of the indirect pathway become overactive, motor symptoms arise as a result of this imbalance (Figure 2B, page 38). With striatal dopamine loss, both pathways lead to the inhibition of GPe and subsequent dis-inhibition of the glutamatergic fibres of the STN (572). The consequent hyperactivity of the GPi/SNr causes the inhibition of motor thalamic nuclei and decreased thalamic output. The result is the underactivity of motor cortical areas, which is reported to occur in the primary sensory motor cortex and supplementary motor area (587) in the parkinsonian state. In a groundbreaking series of experiments by Mitchell and colleagues, the neuronal metabolic marker 2-deoxyglucose (2-DG) was used to reveal the activity states of BG subnuclei in the MPTP-lesioned nonhuman primates (588-590). It was reported that the STN

was hyperactivated, while the GPe and thalamic nuclei were hyperinhibited (588-590). These findings suggested that output structures of the BG are hyperactivated in PD (590), and were later confirmed through electrophysiology and mRNA expression studies (591-593). However, assessment of levels of neural activity in BG based on metabolic markers instead of electrophysiological methods is difficult to interpret because the increase in metabolism can reflect excitatory or inhibitory processes and the balance between pre- and post-synaptic metabolic activity is unclear (594).

3.8.2.2. Basal ganglia circuitry in dyskinesia

Early attempts to describe the pathophysiology of LID proposed a disequilibrium between the direct and indirect pathways that is opposite with respect to PD (595-597). A greater emphasis was placed on the indirect pathway in the pathogenesis of the dyskinetic state and it was proposed that MSNs of the indirect pathway become underactive, leading to disinhibition of the GPe (Figure 2C, page 38) (538). Subsequently, this causes the over-inhibition of the STN and leads to the underactivity of the GPi/SNr. This imbalance disinhibits the motor thalamus and motor cortex, giving rise to the overactivation of motor cortical areas and consequent excessive abnormal movements, which characterize the dyskinetic PD patients (598, 599). In MPTP-lesioned nonhuman primates, Mitchell and co-workers demonstrated that at the peak dose of DA agonist-induced dyskinesia, there was an increased uptake of 2-DG in the STN and GPi, indicating that these structures were hyperinhibited (600). In addition, there was a decrease in the uptake of 2-DG in the motor thalamus, which reflected its hyperactivated state in dyskinesia (600). In contrast, the proposed underactivity of the indirect pathway in LID is generally inconsistent with experimental findings, which presents limitations of the classic model of BG circuitry (538). For example, the underactivation of the indirect pathway due to an overactive GPe is not consistently seen in dyskinetic MPTP-lesioned nonhuman primates (601). Furthermore, MSNs of the indirect pathway are not necessarily underactive, as levels of striatal PPE-A mRNA are actually upregulated in dyskinetic compared to non-dyskinetic PD patients (602, 603).

Bezard and colleagues later emphasized the role of the direct pathway in the pathogenesis of LID (604). The authors suggest that underactive/abnormal firing of the BG output nuclei in dyskinesia (591, 605-610) is primarily caused by overactivation of MSNs of the

direct pathway. Indeed, functional overactivity of the direct pathway in LID has been demonstrated at the cellular level by upregulated striatal mRNA expression of PPE-B and prodynorphin (602, 611-614), and the supersensitization of striatal D1 receptors (614). Moreover, treatment with the selective dopamine D1 receptor agonist ABT-431 in PD patients elicits dyskinesia to a similar extent to that of L-DOPA (615), which further supports the hypothesis of the hyperactivation of the direct pathway in dyskinesia. Overall, these findings are consistent with the mechanism suggested in the classic functional model, where the hyperactive direct pathway mediates over-inhibition of the BG output, resulting in the underactivation of these nuclei.

Recently, Nadjar and colleagues showed that both the phenotype and the targets of striatofugal neurons, are preserved after dopamine denervation in the parkinsonian state and even after chronic L-DOPA treatment in non-dyskinetic and dyskinetic subjects (616). Although these results suggest that the phenotypical plasticity of the striatofugal system is not affected by the experimental condition, it does not exclude the possibility of plastic changes in the striatum (617). In fact, the size of the dendritic tree and density of dendritic spines of MSNs is reduced in the striatum of PD patients compared with controls (618), consistent with the pruning reported to occur in rodents (619) and in MPTP-primates (620, 621). Taken together, these plastic changes contribute to the development of L-DOPA related adverse events by altering the flow of information through the striatum and the rest of the BG (617).

3.8.3. Present: changes to the classic model

In the 1980s, based on the anatomy, neurochemistry, and electrophysiology studies available at the time, the classic model of the functional organization of the BG circuitry was first proposed (617). As discussed above, the classic model is based on the segregation of the direct and indirect pathways where neural processing occurs in a feed-forward manner to achieve behavioural outcome (538). Although the classic functional model has advanced our understanding of functional mechanisms in normal and disease states, the model is too simplistic and limited to describe the pathophysiology underlying PD and LID (538). For example, underactivity of the BG output nuclei in dyskinesia (605, 608, 609) cannot fully account for disease pathogenesis (607). Similarly, lesioning the GPi does not result in dyskinesia (539, 622) and in fact, pallidotomy of the GPi effectively alleviates LID in MPTP-lesioned primates (623)

and PD patients (624, 625), which is contrary to the outcome proposed by the classic functional model.

Experimental reports demonstrate a greater complexity in the neural organization and information processing within the BG (617). This has prompted deviation from the classical model originally described by Alexander and Crutcher (543) to a new functional model, which considers the dynamic neural network in the BG circuitry (597, 626). As demonstrated by major experimental findings in PD and LID, the BG are not simply a “go through” structure, where the connectivity and functional interactions occur along a unilateral fashion along the cortico-basal ganglia-thalamo-cortical circuits (627, 628). The organization of the BG circuitry appears to be interconnected rather than segregated as striatofugal axons consistently collateralize to both the GPe and GPi (629). In addition, major changes to the classic indirect pathway include the GPe as a key structure for inhibitory modulation of the striatum and output nuclei (628, 630, 631) and the STN as another major input station that receives and sends glutamatergic projections (551, 574, 632). Thus, the model now incorporates internal feedback loops (597, 626), and reciprocal connections are found between many nuclei of the circuitry (633-635). Furthermore, the reorganization includes functional dual di-synaptic control of the GPe and GPi (597, 628) via parallel cortical projections to the striatum and STN (538). The corticostriatal projection uses the striatum to exert inhibition of the pallidal segments (538) while the cortico-STN projection uses the STN to mediate fast excitatory input to these structures (636). Indeed, parallel cortico-basal ganglia loops have been confirmed in humans by fMRI and PET studies (637).

3.8.4. Serotonergic system and basal ganglia: implication in Parkinson’s Disease and dyskinesia

The basal ganglia are enriched with a variety of neurotransmitters such as DA, glutamate, acetylcholine and 5-HT (638). In the last decade, there has been growing interest in the role of the serotonergic system in PD and LID, which will be further discussed in detail (section: 3.10. Serotonergic system in dyskinesia, page 47). Briefly, serotonergic neurons in the raphe nucleus project axonal fibres to multiple brain areas including the basal ganglia (639). In the DA-denervated brain in PD, striatal serotonergic terminals contribute increasingly to the conversion of L-DOPA into DA (640, 641). However, the lack of auto-regulatory feedback mechanisms to

control DA release results in the aberrant release of DA into the extracellular space (639). Consequently, fluctuations in DA levels lead to the supersensitivity of postsynaptic dopaminergic receptors and the expression of dyskinesia (642).

The 5-HT receptors are divided in 7 classes (5-HT₁₋₇) with at least 14 subtypes and are all members the GPCR family, except for the 5-HT₃ receptor (see section: 5-HT₃ receptors, page 52). These GPCRs activate an intracellular second messenger to mediate excitatory or inhibitory neurotransmission (643). 5-HT neurons express three subtypes of autoreceptors, amongst which the 5-HT_{1A} and 5-HT_{1B} are the most abundant (644). 5-HT_{1A} receptors are found in the soma and on dendrites (645, 646), whereas 5-HT_{1B} receptors are more abundant in terminals (647) and together, these autoreceptors fine-tune the synaptic release of 5-HT to maintain synaptic levels within a physiological range (645). Agonists of these receptors reduce neurotransmitter release from 5-HT neurons (645, 648-650), and since DA and 5-HT are localized in the same synaptic vesicles after exogenous L-DOPA administration, it is conceivable that they also decrease the release of L-DOPA derived DA from 5-HT terminals (394, 640, 651).

The BG nuclei receive serotonergic afferents that mainly originate from the dorsal raphe nuclei (see review in (652)). Furthermore, the BG contain 5-HT, its metabolite 5-hydroxyindoleacetic acid (5-HIAA) (560, 653, 654), the 5-HT transporter (SERT) and 5-HT receptors 5-HT₁ to 5-HT₇. The distribution of 5-HT receptors in these structures is heterogeneous and varies between species (655). 5-HT modulates the activity of BG nuclei by acting on 5-HT receptors and helps maintain the balance between the direct and indirect pathways (656).

Parent and colleagues conducted an immunohistochemistry study to visualize the 5-HT system innervation of BG in human and nonhuman primates (657). The SN is the most densely innervated BG subnucleus and nigral 5-HT innervation originates from axons and arborizes immediately upon entering the SN (657). In addition, 5-HT mainly exerts an inhibitory effect on the activity of SNc DA neurons projecting to the striatum and SNr GABAergic neurons projecting to the thalamus and brainstem in humans and primates (657). Conversely, 5-HT depletion decreases firing rate and increases burst activity of SNr neurons (658). In spite of the apparent inhibitory effect of 5-HT input on the SNc (659, 660), lesioning the dorsal RN does

not alter SNc activity (661), so it is still unclear how 5-HT transmission modulates the activity of dopaminergic SNc neurons (655).

In addition, the striatum receives dense serotonergic afferents from the dorsal RN, where local administration of 5-HT inhibits the majority of the striatal cells (662-664). Furthermore, many 5-HT varicosities in the striatum could be visualized in close apposition with the pigmented cell bodies of the SNc (657). Stimulation of pre-synaptic 5-HT_{1A} and 5-HT_{1B} receptors inhibits striatal 5-HT release (648, 665) and activation of the 5-HT_{1A} receptor also decreases glutamate release from corticostriatal projections (666-668). In contrast, the 5-HT₂ receptor exerts an inhibitory effect on striatal neuron activity, mainly by targeting MSNs (669, 670).

In the STN, 5-HT can act at the pre- and post-synaptic levels and, given the functional diversity of 5-HT receptors, exert multiple effects (671-673). The activity of STN neurons is modulated by 5-HT afferents and in primates, 5-HT axonal varicosities are apposed almost only upon dendritic spines or branches and many terminals do not form synaptic contacts (657). Pharmacologic lesion of the DRN and 5-HT depletion increases STN firing frequency and bursting activity *in vivo* (674, 675).

Electrophysiology studies in rats indicate that 5-HT controls its effect by both pre- and post-synaptic mechanisms at the pallidal level (676). The decrease in 5-HT concentrations can lead to changes in pallidal activity and contribute to abnormal synchronous oscillations in BG components (657). Moreover, in the GPe, 5-HT depletion decreases the firing frequency and increases the proportion of bursty and irregular neurons (677), these results have been confirmed by a patch-clamp recording study where 5-HT perfusion increased the firing rate of GPe neurons (678). In addition, 5-HT can decrease the pre-synaptic release of glutamate and GABA from subthalamic and striatopallidal terminals, respectively, by acting on 5-HT_{1B} receptors (679).

In PD and subsequent L-DOPA replacement therapy, the 5-HT system adapts to DA depletion by adopting anatomical and functional transformations (655). However, the changes occurring after dopaminergic lesion in animal models of PD differ across research groups, which may reflect methodological differences including the parkinsonian state, the age of the animals, injection site, toxin concentration, and the time between surgery and performing the studies (655). Similarly, striatal 5-HT levels have been reported to be increased (680, 681), unchanged

(641, 682) or decreased (683) in parkinsonian animals. On the other hand, the dorsal RN also undergoes adaptive changes after the dopaminergic degeneration such as the increased 5-HT_{1A} expression in MPTP-lesioned primates (684). Overall, the effects of 5-HT in the BG depend on the specific nucleus and its receptor population (655). For example, 5-HT exerts an inhibitory action on striatal MSNs via direct or indirect activation of 5-HT receptors, as well as in the STN and SNr *in vivo*, whereas the overall effect of 5-HT is excitatory in the GPe (655).

3.9. Dopaminergic system in dyskinesia

Mechanisms involved in the pathophysiology of LID are complex and have been investigated in studies using animal models and parkinsonian patients.

In general, DA cell loss in the nigrostriatal pathway and chronic administration of L-DOPA or DA agonists, are viewed as necessary conditions for the appearance of LID (685).

3.9.1. Dopamine receptor supersensitivity and dopamine sensitization

Denervation-induced supersensitivity of DA receptors has been recognized as a plausible mechanism of LID. However, it is likely more complex than simply an increase in the density of striatal DA receptors (686-689); thus, according to this theory, then LID might appear with first dose of L-DOPA. However, LID does not usually emerge at the first exposure to L-DOPA but gradually develops over years of L-DOPA therapy, as discussed above. The development of LID appears to be related to an increase in the activity of D1, D2, D3 receptor subtypes while additional studies are required to discern the contribution of D4 and D5 receptors (685).

3.9.2. D1-like family of dopamine receptors

In early studies in primates, it was shown that D1 agonists were as effective as D2 agonists to improve parkinsonian symptoms, while inducing less dyskinesia (690-692). Later studies, however, implicated a more important role of D1 receptors in dyskinesia. A study in drug-naïve MPTP-lesioned primates found that chronic administration of a D1 receptor agonist led to the development of dyskinesia (693). Consistent with this data, recent studies in 6-OHDA-lesioned rats reported that D1 agonists induce dyskinesia, and that pharmacological blockade of D1 receptors was more effective than D2 receptor antagonism at alleviating dyskinesia (457, 694, 695). Furthermore, genetic knockout of D1 receptors completely

suppressed LID in parkinsonian mice, whereas D2 receptor knockout mice developed LID similar to wild-type mice (696).

Autoradiographic studies on D1 receptors *in vivo* and on *post-mortem* tissues of animal models and in PD patients have been conducted with no general consensus (697), which may be due to the differences in experimental assays and the subregion of the striatum measured (685). Although the association between the expression of D1 receptors and dyskinesia is unclear, the sensitivity of D1 receptors, measured by GTP γ S binding, was reported to be linearly related to the severity of LID (686). L-DOPA induced a decrease in the sensitivity of D1 receptors in non-dyskinetic MPTP-lesioned primates, whereas its sensitivity was increased in dyskinetic animals (686). Moreover, D1 receptors are internalized in the cytoplasm in 6-OHDA-lesioned rats compared to normal rats (698), which is also observed in PD patients, where D1 receptors are preferentially localized to the cytoplasm compared to healthy controls (699). However, it is unclear if the change in subcellular distribution of D1 receptors is a consequence of distribution in development, priming process or expression of LID (685).

D1 receptors interact with a variety of receptors and trigger signalling pathways that have an influence on dyskinesia development. D1 receptors interact with ionotropic glutamate NMDA receptors at the post-synaptic striatal level, and may also form hetero-oligomeric complexes (700). This interaction affects the trafficking, signalling and desensitization of both receptors (701, 702) and importantly, these complexes are lost in dyskinetic 6-OHDA-lesioned rats (703). Furthermore, extracellular signal-regulated kinase (ERK) is part of the intracellular pathways of both NMDA and D1 receptors (704). ERK intracellular signalling is associated with LID priming process (705) and the expression of LID is reduced with pharmacological inhibition of ERK intracellular signalling (706, 707). On the other hand, dopaminomimetic agents induce the expression of transcription factors such as c-jun, c-fos, Δ FosB, FosB in striatal neurons in normal (708, 709) and hemiparkinsonian animals (350) and require the activation of D1 but not D2 receptors (709). No study has reported the involvement of the D5 receptor in PD and LID (685).

3.9.3. D2-like family of dopamine receptors

Once primed to express LID, D2 agonists will trigger abnormal involuntary movements (AIMs) in 6-OHDA-lesioned rats (710, 711) and LID in MPTP-lesioned primates (712, 713) and PD patients (714), producing greater dyskinesia than D1 agonists (690). Results from autoradiographic studies were more consistent than with D1 receptors, but again, the inconsistency observed could be due to variation in experimental assays and the striatal region studied (685). Expression of D2 receptors in the striatum remained unchanged (715, 716) or increased (717, 718) with MPTP lesion in primates and in untreated PD patients (719, 720). D2 receptor agonists reduced MPTP-induced upregulation of D2 receptors (721, 722) not as efficiently as L-DOPA whereas D1 receptor agonists had no effect or produced an increase of D2 receptors (686). A PET study in *de novo* PD patients reported similar observations (723, 724). *In situ* hybridization studies in primates demonstrated that MPTP induced an upregulation of striatal D2 receptor mRNA that was completely reversed by L-DOPA treatment (725) or unaffected (686, 726), whereas D2 receptor agonists decreased or reversed this expression (727). The influence of D2 receptor trafficking in PD and LID is not yet established (685).

3.10. Serotonergic system in dyskinesia

The chemical structure of 5-HT was identified in 1953 (728) and a few years later, its function as a neurotransmitter in the CNS was proposed (729). 5-HT plays major roles in the regulation of mood and emotion, cognition, feeding and satiety, circadian and sleep-wake cycle regulation, pain, and motor control through a variety of receptor subtypes (730, 731). 5-HT is synthesized from L-tryptophan by the following reactions: the tryptophan hydroxylase enzyme generates 5-hydroxytryptophan (5-HTP), which is then converted to 5-HT by the AADC, the same enzyme that catalyzes the conversion of L-DOPA into DA. In the brain, 5-HT neurons are clustered within the midbrain raphe nuclei (areas B1-B9) (732) and include the dorsal, median, magnus, obscuris, and pontis RN (733). Anatomical studies further subdivided the midbrain RN into two main groups based on position within the brainstem and axonal projections. The rostral group, which contains the dorsal RN comprises the B7 and B8 cell clusters whereas the median RN consists of the B5, B8 and B9 cell clusters. 5-HT plays a key role in the CNS as 5-HT neuron soma from the midbrain send projections throughout the entire CNS (734). In fact, one 5-HT cell body can be responsible for up to 500,000 cortical varicosities (735), which underscores the

widely distributed innervations derived from these nuclei, including the majority of 5-HT innervation to the forebrain regions (736, 737). Moreover, within the dorsal RN, 5-HT neurons are topographically organized and studies also demonstrate that 5-HT neurons within each sub-region differ morphologically, electrophysiologically, and molecularly with respect to receptor expression (738).

There is a growing appreciation for the multifaceted effect of 5-HT in PD and recent studies suggest that the 5-HT system is heavily implicated in the pathophysiology of LID (641). In animal models of PD, it has been suggested that the 5-HT system is an important source of striatal L-DOPA derived DA release (641, 739-741). In PD patients undergoing L-DOPA therapy with severe degeneration of the nigrostriatal DA system and compromised function in the remaining DA neurons (742-744), the 5-HT neuron derived “false transmitter” can help improve motor disability (641). With chronic administration of L-DOPA, and particularly in advanced stages of the disease, the lack of a regulatory mechanism to control synaptic neurotransmitter levels can provoke dyskinesia (745).

3.10.1. Pre-clinical evidence for the involvement of the serotonergic system in dyskinesia

Pharmacological studies also demonstrate the involvement of 5-HT neurons in the appearance of LID in animal models, as discussed above. It is generally assumed that in early stages of disease, L-DOPA is taken up into spared nigrostriatal DA neurons of PD animal models and patients, converted into DA, stored into vesicles and released in a physiologically regulated manner (746). The resultant DA release from these dopaminergic terminals within the striatum accounts for the therapeutic action of L-DOPA and is finely regulated by D2 autoreceptors and the DAT (746). With disease progression, however, fewer DA terminals can convert exogenous L-DOPA into DA. Other cellular compartments can compensate for the loss of DA neurons in mediating the conversion of L-DOPA to DA and neurotransmitter release. 5-HT neurons possess the same enzymatic machinery as dopaminergic terminals as they express AADC and VMAT2 (394, 395, 747) and thus, are able to convert L-DOPA to DA and to mediate its storage into synaptic vesicles (746). Various studies have demonstrated that 5-HT neurons can store and release DA *in vivo* and *in vitro* (644). The first report implicating 5-HT neurons as a source of DA release was provided by Tanaka and colleagues (743). In the study, removal

of 5-HT innervation by 5,7-dihydroxytryptamine (5,7-DHT) administration reduced L-DOPA derived extracellular DA levels by about 80% in the striatum of 6-OHDA-lesioned rats (743). Importantly, this also led to a near-complete suppression of LID in L-DOPA primed parkinsonian rats. Furthermore, the same group also showed a similar reduction in extracellular DA levels after co-administration of the 5-HT_{1A} agonist 8-hydroxy-2-(di-n-propylamino)tetralin (8-OHDPAT) with L-DOPA (744). Another group used a similar approach to demonstrate that lesion to the 5-HT system suppresses L-DOPA induced rotational behaviour and striatal c-Fos expression in 6-OHDA-lesioned rats (748). Taken together, these studies suggest that the action of L-DOPA in PD depends, at least in part, on its conversion to DA in 5-HT neurons (308).

Risk factors that underlie the development of dyskinesia suggests that it is the progression of dopaminergic degeneration rather than the duration of L-DOPA treatment that is responsible for the emergence of LID over time (749). Indeed, parkinsonian animals only develop severe dyskinesia with extensive DA denervation whereas partially lesioned animals show no or only mild dyskinesia (745). Using a viral vector delivery of short hairpin RNA for TH, Ulusoy and colleagues induced significant DA deficiency, and reported that DA-depleted rats were resistant to the induction of dyskinesia following administration of high dose of L-DOPA as opposed to control animals with similar striatal DA depletion (750). These findings may be explained by the relatively preserved striatal DA terminals after inhibition of DA synthesis, which act as a buffering system for exogenous L-DOPA (745). Rat transplantation studies also confirmed the ability of pre-synaptic DA compartment to prevent excessive DA receptor stimulation, as L-DOPA primed dyskinetic rats tended to normalize response to L-DOPA after receiving ventral mesencephalic dopaminergic neuronal grafts into the lesioned striatum (739, 751). Similarly, in the clinic, as neurodegeneration progresses in PD patients, susceptibility to dyskinesia also increases over time (752). DA surges reflect uptake and conversion of exogenous L-DOPA by cells other than nigrostriatal DA neurons (443). Accordingly, several studies have demonstrated that 5-HT neurons become the main source of DA release in severely DA-denervated animals and that 5-HT neuron derived DA release is important in LID (308). In 6-OHDA lesioned rats, Carta and colleagues demonstrated that DA release from the 5-HT system is responsible for the appearance of LID (641). In fact, animals

subject to toxic lesion of the 5-HT system by the neurotoxin 5,7-DHT or pharmacological blockade of these neurons by 5-HT_{1A} and 5-HT_{1B} agonists leads to the silencing of dyskinesia upon treatment with L-DOPA (641). Moreover, studies consistently found that 5-HT_{1A} and 5-HT_{1B} agonists exert anti-dyskinetic effects in animal models of LID (641, 753-755) .

In early stages of disease, the therapeutic efficacy of L-DOPA and the physiological release of DA by 5-HT neurons is beneficial because the remaining DA terminals can buffer the 5-HT neuron-derived DA and avoid excessive post-synaptic DA receptor stimulation (642). DA D2 autoreceptors are located on the pre-synaptic membrane and activate a feedback control mechanism to fine-tune neurotransmitter release and allow the maintenance of physiological-like synaptic DA levels (745). However, with disease progression, the therapeutic efficacy of L-DOPA is partly compromised and, 5-HT neurons contribute increasingly to the conversion of exogenously administered L-DOPA to DA, eventually producing excessive DA receptor activation (745). Moreover, unlike DA neurons, 5-HT neurons lack autoregulatory feedback to control DA release (465, 756). As a result, the non-physiological release of DA leads to large fluctuations in synaptic DA levels, causing pulsatile stimulation of striatal DA receptors and aberrant downstream signalling cascade (757).

3.10.1.1. 5-HT₁ agonists in the treatment of dyskinesia

Munoz and colleagues observed that a combination of 5-HT_{1A} and 5-HT_{1B} receptor agonists, using low doses of 8-OH-DPAT and CP-94253, synergistically suppressed L-DOPA induced AIMs in 6-OHDA-lesioned rats (758). These results were also obtained in dyskinetic MPTP-lesioned macaques (758). Accordingly, a rat microdialysis study reported a reduction of extracellular DA levels that account for the potent anti-dyskinetic effect of 5-HT_{1A} and 5-HT_{1B} receptor agonists (759). In a rat PET study, Nahimi and co-workers showed that administration of 8-OH-DPAT reverses L-DOPA induced decrease of [¹¹C]-raclopride binding and increases extracellular DA in 6-OHDA lesioned rats (760).

Agonists for the 5-HT_{1A} receptor have shown acute and chronic efficacy in animal models and clinical studies for LID but at the expense of the therapeutic efficacy of L-DOPA (641, 740, 741, 761-763). The 5-HT_{1A} receptor agonist sarizotan demonstrated efficacy in reducing dyskinesia in rodent and primate models of PD, as well as in idiopathic PD patients in early open-label studies (762). However, the anti-dyskinetic effect was not significantly

different compared to placebo in two Phase III clinical trials (764). Similarly, the partial non-selective 5-HT_{1A} receptor agonist buspirone reduced LID in patients (765) but two other studies found that this effect compromised the therapeutic efficacy of L-DOPA (766, 767). 5-HT_{1B} receptor agonists can produce anti-dyskinetic effects in animal models of PD but no clinical trials have been performed with these yet (768).

3.10.1.2. 5-HT_{2A} antagonists in the treatment of dyskinesia

5-HT_{2A} receptors are localized post-synaptically and in general, they exert an excitatory effect (768). Rahi and colleagues demonstrated an increase in 5-HT_{2A} receptors in the striatum of dyskinetic primates when compared with non-dyskinetic animals (769). Preclinical and clinical studies have shown the efficacy of drugs acting on 5-HT_{2A} receptors in controlling L-DOPA-induced motor complications (770, 771) but the results are contradictory. In MPTP-lesioned primates, the selective 5-HT_{2A} inverse agonist pimavanserin reduced LID without worsening motor scores (459), whereas another antagonist, ritanserin, alleviated LID but worsened L-DOPA anti-parkinsonian action (772). In addition, the 5-HT_{2A} antagonist volinanserin did not reduce LID in hemiparkinsonian rats (773). Thus, further work is required to establish whether 5-HT_{2A} antagonists can be beneficial in dyskinetic patients (768).

3.10.1.3. Clinical evidence for the involvement of the serotonergic system in dyskinesia

Clinical evidence on the effectiveness of 5-HT modulation in LID is still scarce (745). An open-label double-blind study on the efficacy of sarizotan, a partial 5-HT_{1A} receptor agonist in dyskinetic patients was terminated for lack of efficacy (764, 774). This may be attributed to its antagonistic activity at the D₂ receptors (775), as well as its action on only the 5-HT_{1A} receptor, whereas experimental evidence has demonstrated that simultaneous targeting of 5-HT_{1A/1B} auto-receptors exerts a synergistic effect to attenuate LID (741, 758). In fact, a Phase II study with the mixed 5-HT_{1A/1B} agonist eltoprazine has shown promising results (776).

Consistent with microdialysis experiments in rats, a PET study provided support for the association between dyskinesia and dysregulated DA release (752). Dyskinetic patients showed higher synaptic DA levels one hour after L-DOPA administration compared to non-dyskinetic subjects, which led the authors to propose that dyskinetic patients have difficulty maintaining

DA levels within certain limits, likely caused by progressive degeneration of DA neurons and consequent reduced ability to mediate controlled DA release. In line with this view, a PET study using the radioligand [^{11}C]-3-amino-4-(2-dimethylaminomethylphenylsulfanyl)-benzotrile to evaluate 5-HT terminal function and the radioligand [^{11}C]-raclopride to evaluate striatal DA release found that dyskinetic PD patients showed relative preservation of serotonergic terminals and no difference compared to non-dyskinetic PD patients (777). Furthermore, in dyskinetic PD patients, the same L-DOPA dose induced higher striatal synaptic DA concentration in PD patients with LIDs compared to non-dyskinetic PD patients, which is in agreement with previous studies (752, 778). Administration of the partial 5-HT_{1A} agonist buspirone, prior to L-DOPA treatment, reduced L-DOPA evoked rise in striatal synaptic DA and attenuated LID (777). Further dividing the LID group by severity into milder and severe forms, it was found that buspirone modulated DA levels to a greater extent in PD patients with mild LIDs compared to those with severe LIDs (777). The authors concluded that striatal serotonergic terminals contribute to LID in human PD via aberrant processing of exogenous L-DOPA and release of DA as false neurotransmitter. Finally, the SERT-to-DAT binding ratio increases in PD patients that experience LIDs, when compared with non-dyskinetic PD patients, which further supports the notion that when dopaminergic innervation in the striatum is low, the 5-HT system is critical to the development of LIDs (779).

Collectively, these experimental findings provide strong evidence supporting the pivotal role of 5-HT neurons in the induction and expression of LID. An important challenge for future clinical studies will be to preserve the therapeutic effect of L-DOPA following pharmacological dampening of 5-HT neuron activity (745).

4. 5-HT₃ receptor

Although many serotonergic drugs tested have demonstrated efficacy in reducing dyskinesia, most have been at the expense of impairing L-DOPA anti-parkinsonian action (639). In MPTP-lesioned non-human primate models of PD, for example, selective 5-HT_{1A} and 5-HT_{1B} receptor agonists reduced dyskinesia but induced suppression of locomotor activity and increased motor disability (780, 781). The 5-HT₃ receptor is an interesting target to study in the context of dyskinesia because several drugs are clinically available to modulate its function and

the 5-HT₃ receptor has been shown to modulate striatal DA release, as discussed in detail (section: 5-HT₃ receptors in Parkinson's Disease and L-DOPA-induced dyskinesia, page 57).

The 5-HT₃ receptor is the sole ligand-gated ion channel amongst the 5-HT receptor family, while the other 5-HT receptors are all metabotropic GPCRs that modulate an intracellular second messenger system (782). The 5-HT₃ receptor was first identified in the guinea pig ileum, and then more widely distributed in the peripheral nervous system (PNS) (783). The presence of 5-HT₃ receptors in the brain was initially a subject of controversy (784) until adequate ligands were developed to conduct membrane binding and autoradiography studies (785-788), which suggested the presence of 5-HT₃ receptors in the CNS. In the PNS, the activation of 5-HT₃ receptors regulates autonomic, parasympathetic and sensory functions (789). 5-HT₃ receptors located on vagal sensory afferents exert pronounced effects on the cardiovascular system (790, 791) and also control motility and peristalsis throughout the gastrointestinal tract (792). In addition, 5-HT₃ receptors regulate nociceptive processing (793-795), which is consistent with their expression in the dorsal root ganglion and neurons in dorsal horn of the spinal cord (795-798). In the CNS, 5-HT₃ receptors in the hippocampus and nucleus accumbens are implicated in anxiety (799). Moreover, 5-HT₃ receptors are implicated in drug addiction and alcohol consumptions in rats (800) and in humans (801) and they are also important for cognitive function in elderly patients (802).

4.1. Localization of 5-HT₃ receptors

The distribution of 5-HT₃ receptors has been studied through autoradiographic, immunohistochemistry and *in situ* hybridization techniques with variation across different species (803), likely reflecting differences in the methodology and choice of ligands. In the PNS, 5-HT₃ receptors have been detected on pre- and post-ganglionic autonomic neurons and on neurons of the sensory and enteric nervous system (804-806).

In the CNS, when compared to other 5-HT receptors, the 5-HT₃ receptor displays a relatively lower density (805, 807). The highest density of 5-HT₃ receptors are found within the dorsal vagal complex in the brainstem (807, 808), which comprises the nucleus tractus solitarius, area postrema and dorsal motor nucleus of the vagus nerve (784). Outside the brainstem, the highest levels of 5-HT₃ receptors are expressed in regions such as the hippocampus (788), amygdala and superficial layers of the cerebral cortex (807, 808). However, the distribution

within the forebrain displays species variations, and in humans, for example, there are relatively high levels of 5-HT₃ receptors within the caudate nucleus and putamen (809) whereas low levels are detected within cortical regions (809-811). In contrast, autoradiographic and homogenate binding studies in the rat brain have demonstrated high levels of 5-HT₃ receptors in the neocortex (786, 812-816), hippocampus (786, 812-815), amygdala (786, 812-814) and dorsal vagal complex (813, 814), whereas a low density of 5-HT₃ receptors is detected in the striatum (786, 815), RN (813), SN (813) and nearly absent in the cerebellum (786, 815). An autoradiographic binding study conducted in the mouse, ferret and rabbit brains also showed a similar distribution of 5-HT₃ receptors within the brain (808). Moreover, an immunohistochemistry study in the Syrian hamster brain also reported similar results with high levels of the 5-HT_{3A} (5-HT_{3A}) subunit within the neocortex and amygdala, and intermediate levels in the striatum, SN, GP and DRN (817). In general, 5-HT₃ receptors are concentrated in regions involved in the initiation and coordination of the vomiting reflex, which may explain the relevance of 5-HT₃ receptor antagonists in chemotherapy-induced emesis (782) as well as pain processing and control of anxiety (818).

Consistent with the mapping of 5-HT₃ receptors in autoradiographic studies, *in situ* hybridization studies indicate that in the rodent brain, 5-HT_{3A} receptor mRNA transcripts are similarly distributed to radiolabelled 5-HT₃ receptor binding sites (819). 5-HT₃ mRNA is present in interneurons in the hippocampus and prefrontal cortex (820, 821) and this distribution indicates that 5-HT₃ receptors may mediate the indirect inhibition of excitatory pyramidal neurons via activation of GABAergic interneurons. Furthermore, 5-HT₃ receptor-like immunoreactivity is primarily associated with GABA-containing neurons in the cerebral cortex and hippocampus that often co-localize with the peptide hormone cholecystinin (804, 822, 823) or the calcium-binding protein calbindin (804).

4.2. 5-HT₃ receptor subtypes and properties

5-HT₃ receptors share electrophysiological and structural patterns with the nicotinic acetylcholine and GABA type A receptors, other members of the Cys-loop superfamily (824). A functional channel consists of five symmetrically-arranged subunits that surround a central ion-conducting pore (825). In rodents, two subunits have been cloned thus far: 5-HT_{3A} (826) and 5-HT_{3B} (827) receptor subunits, whereas three additional subunits have been identified in

humans: 5-HT_{3C}, 5-HT_{3D} and 5-HT_{3E} subunits (828-830). The subunits can be arranged to form a homomeric (5-HT_{3A} only) receptor or heteromeric (5-HT_{3A} and 5-HT_{3B} subunits) receptor (827). Although the 5-HT_{3C}, 5-HT_{3D} and 5-HT_{3E} subunits are likely to form only heteromeric receptors with 5-HT₃ receptor subunits, their function is still being debated (828, 831).

In contrast to the 5-HT_{3A} subunits, the 5-HT_{3B} subunit does not form functional homopentameric channels because of its retention in the endoplasmic reticulum (832). 5-HT₃ receptors in the CNS and PNS may be constructed of different subunits and, although it is known that all receptors contain the 5-HT_{3A} subunit, the distribution of the 5-HT_{3B} subunit is still unclear (833). Immunohistochemical studies suggested that the expression of the 5-HT_{3B} subunit is restricted to the PNS (798, 834) but *in situ* hybridization studies showed that the 5-HT_{3B} subunit mRNA is present in the human brain (827). Furthermore, immunocytochemical studies report that 5-HT_{3B} subunits are found in rat hippocampal neurons (835). Thus, it has been proposed that the 5-HT_{3B} subunit is either present in low levels in the CNS or in very discrete localized cell populations (833). The function of 5-HT₃ receptors depends on receptor composition (836, 837), and expression of the 5-HT_{3B} subunits leads to an increase in single channel conductance and lower permeability to Ca²⁺ (827, 838). Furthermore, heteromeric receptors show faster activation and deactivation kinetics than homomeric receptors (839). The differences observed between *in vitro* and *in vivo* studies (803) may be explained by the fact that *in vitro* studies tend to be performed in cultured cells expressing only homomeric 5-HT₃ receptors (840). Whether the 5-HT_{3B} subunit is a major determinant of 5-HT₃ receptor function in the CNS is still being debated (836, 841, 842) and may depend on species-specific expression patterns (843).

4.3. Physiology and pharmacology of 5-HT₃ receptors

The 5-HT₃ receptor is permeable to Na⁺, K⁺ and Ca²⁺ (844, 845) and its function depends on whether it localizes to nerve terminals or post-synaptic cells (803). Differences in the cellular localization of pre- and/or post-synaptic 5-HT₃ receptors within different cerebral regions appear to depend on the nature of the neuron that bears these receptors (846, 847). The preferential localization on nerve endings is consistent with a physiological role of the 5-HT₃ receptor in the control of neurotransmitter release (848). Activation of pre-synaptic 5-HT₃ receptors is followed by rapid membrane depolarization, which causes a rapid rise in cytosolic

Ca^{2+} concentration by inducing Ca^{2+} influx and mobilizing intracellular Ca^{2+} stores, and modulates the release of neurotransmitters and neuropeptides including DA, cholecystokinin, acetylcholine, GABA, substance P or 5-HT itself (849, 850). In contrast to 5-HT₃ receptors found predominantly in pre-synaptic regions associated with axons and terminals, in the hippocampus, they are mostly located on post-synaptic receptors in somatodendritic regions (846). Here, activation of post-synaptic 5-HT₃ receptors leads to depolarization by Na^+ influx and K^+ efflux (851) where it mediates fast synaptic transmission (852, 853). Furthermore, pre- and post-synaptic 5-HT₃ receptors exhibit distinct electrophysiological profiles with differences in single channel conductance, kinetics and re-sensitization time-course (827, 854, 855). For example, the permeation properties differ such that pre-synaptic 5-HT₃ receptors are highly permeable to Ca^{2+} (851, 856-858), whereas post-synaptic receptors are less permeable to Ca^{2+} compared to Na^+ and K^+ (844, 859).

Activation of the 5-HT₃ receptor by its physiological ligand 5-HT, leads to the influx of cations through the open ion channel, which causes depolarization of the cell (860). In addition to 5-HT, DA may be another endogenous ligand for 5-HT₃ receptors, as it displays low-affinity agonism of the 5-HT₃ receptor (861). Frequently used 5-HT₃ receptor agonists are 1-(*m*-chlorophenyl)-biguanide (mCPBG), 2-methyl-5-HT and phenylbiguanide. However, they do not readily penetrate the BBB (862, 863), which limits their usefulness in *in vivo* studies. In recent years, SR57227A has been proposed as a high affinity agonist of the 5-HT₃ receptor. SR57227A may be a useful tool to study the function of 5-HT₃ receptor in both *in vitro* and *in vivo* studies (864) given its ability to cross the BBB and its affinity to central 5-HT₃ receptors (863, 865, 866). However, due to the emetogenic and anxiogenic effects of 5-HT₃ agonists, they have no therapeutic potential (839).

In contrast to 5-HT₃ receptor agonists, a variety of highly specific and potent antagonists have been developed in the last three decades and they are currently the gold standard to treat chemotherapy-induced emesis (867). 5-HT₃ receptor antagonists can be identified by the suffix setron, and competitively bind to the orthosteric ligand binding site of 5-HT₃ receptors (837). However, the nature of receptor antagonism varies, which may account for differences in their pharmacokinetic profile (868). The affinities of common 5-HT₃ receptor antagonists are in the low nanomolar concentration range (7.73 to 10.45 nM) and include dolasetron, ondansetron,

granisetron, tropisetron and palonosetron (869). 5-HT₃ antagonists are only non-selective at concentrations 100-fold or greater in excess of those required to antagonize the 5-HT₃ receptor (870, 871) and their non-selective effects include agonism of 5-HT receptors (870, 872), antagonism of non-5-HT₃ receptors (873, 874), and local blockade of ionotropic receptors (875).

In spite of the actions cited above, pharmacological blockade of 5-HT₃ receptors does not modify normal animal behaviour or physiological function in healthy volunteers except for intestinal transit time (818). However, 5-HT₃ receptor antagonists demonstrated clinical efficacy in various forms of emesis like chemotherapy-induced, radiotherapy-induced, and post-operative emesis (867, 876) and extends to other indications such as irritable bowel syndrome (877, 878), anxiety (879), chronic fatigue syndrome (880), alcohol abuse (881), fibromyalgia (882) and migraine (883). Although the use of 5-HT_{3A} knockout mice has not contributed much to the role of 5-HT₃ receptors (805), studies that investigate the effects of specific genetic alterations of 5-HT₃ receptors (884) may further illuminate the function of these receptors.

4.4. 5-HT₃ receptors in Parkinson's Disease and L-DOPA-induced dyskinesia

Pharmacological modulation of the 5-HT system, particularly the 5-HT_{1A} and 5-HT_{1B} receptors, as discussed above, has demonstrated efficacy in preclinical and clinical studies of dyskinesia. Although the 5-HT₃ receptor has been understudied in the context of PD and its role is unknown in LID, its distribution in BG draws attention to the potential of the 5-HT₃ receptor as a novel therapeutic target for dyskinesia. Administration of pharmacological compounds including clozapine (528, 885, 886), mirtazapine (887), quetiapine (770), AQW051(888) and AZD0328 (889) have reduced the severity of dyskinesia in animal models of PD and/or in clinical settings. However, given the non-selective effects of these therapeutic agents, namely as antagonists of the 5-HT₃ receptor (837, 890-896), it is conceivable that pharmacological blockade of 5-HT₃ receptors may have contributed to the anti-dyskinetic effect of these compounds.

The 5-HT₃ receptor is poorly characterized in the BG compared to other members of the 5-HT receptor family (652) and only a few radioligand binding experiments have used highly-specific drugs to study its distribution in these nuclei. There is some controversy amongst the

literature concerning the variation in the binding sensitivity of the selected ligands (897). For instance, radioligand binding of the 5-HT₃ receptor within the rodent striatum differed depending on the molecule used, with low levels with the antagonist radioligands [3H]-GR 65630 (786) while strong levels of binding were observed with the agonist radioligand [3H]-mCPBG (814) and the antagonist radioligand [125I]-iodozacopride (898). Some authors have reported the presence of 5-HT₃ receptors in the striatum of different mammals and intriguingly, they observed relatively higher receptor densities of 5-HT₃ receptors in human striatum compared to rat striatum (788, 809, 812, 813, 899-903), although the functional significance of this species difference is poorly understood. Of note, homogenate binding studies from patients with Huntington's disease (HD) and PD suggest that the 5-HT₃ receptor is localized to GABAergic output neurons of the caudate putamen and not predominantly located on DA neurons (904). In fact, the density of 5-HT₃ receptors was not affected by the neurodegeneration associated with PD, whereas a significant proportion of HD cases showed decreased 5-HT₃ receptor binding in the striatum. HD is neuropathologically characterized by the degeneration of neurons with cell bodies within the caudate putamen, which include MSNs. Thus, these studies suggest that at least a proportion of 5-HT₃ receptors is localized on neurons which degenerate in HD but not on DA terminals which degenerate in PD (904). Furthermore, the use of rat striatal synaptosomes showed the presence of functional pre-synaptic 5-HT₃ receptors as well as its known post-synaptic localization (851, 856, 857). In contrast, membrane binding assays and immunolabelling of the rat and human brains only detected low levels of the 5-HT₃ receptor in the SN (813, 898, 900, 905).

The dysregulated release of DA in the striatum is a potential pre-synaptic mechanism responsible for the progression of dyskinesia in parkinsonian animals and patients with PD. In addition to the distribution of the 5-HT₃ receptor in the striatum and SN, studies also demonstrate that the 5-HT₃ receptor modulates central dopaminergic activity. Indeed, 5-HT agonists stimulate the striatal release of DA *in vitro* (865, 906-910) and *in vivo* (911, 912), which is reversed by 5-HT₃ antagonists (913). In further support of these findings, behavioural studies also report that the 5-HT₃ receptor modifies nigrostriatal DA transmission-mediated motor responses such as stereotypy (914), orofacial dyskinesia (915) or rotations (863). Ondansetron is a potent and highly-selective prototypical 5-HT₃ receptor antagonist used as an anti-emetic in

patients receiving cancer chemotherapy (916). It has been demonstrated that ondansetron reduces basal concentration of DA in the nucleus accumbens (917) and modifies mesolimbic DA activity in the rat and marmoset brains (918, 919), such as the inhibition of amphetamine induced hyperactivity. Moreover, in clinical studies, administration of ondansetron led to improvements in tardive dyskinesia and psychotic symptoms (920) and attenuated psychosis in advanced PD patients (921, 922).

5. Animal models of Parkinson's disease

Animal models of PD, categorized into toxin or genetic, have led to the discovery of novel symptomatic treatments and uncovered mechanisms underlying some features of the disease (59, 923-925). Pharmacological interventions can be used to mimic the motor deficits of parkinsonism including neuroleptic-induced catalepsy (926) or reserpine-induced akinesia (927, 928). The most characterized neurotoxin-induced animals models are the complex I inhibitor of the respiratory chain MPTP in primates (925, 929) and mice (283, 930) and the 6-OHDA rodent model of PD (931). On the other hand, genetic models in mice have more recently been developed (924) including the transgenic overexpression of mutant genes (α -synuclein and LRRK2) or the knockdown or knockout of autosomal recessive genes (PINK1, PARKN, DJ-1) (932), and in transgenic mice, preformed fibrils seed Lewy pathology and decrease survival time (933). Moreover, viral vector-mediated overexpression of α -synuclein in rats (251, 252, 934) and primates (253, 254, 935) have reproduced hallmarks of PD including Lewy-like synucleinopathy, progressive dopaminergic cell loss, and even a parkinsonian behavioural phenotype in rodents (936). Although genetic models have provided greater insight into the molecular mechanisms underlying PD, the recapitulation of genetic alterations discussed above (see section: genetic risk factors of PD, page 5) tends to elicit modest loss of DA neurons (937-940) and does not fully capture the neuropathology of PD (924). Moreover, in spite of the reports of alterations in the motor function and behaviour of animals (939, 941, 942), the behavioural phenotypes are often distinct from the human condition (924). As the experimental work presented in this Thesis was conducted in the 6-OHDA-lesioned rat, the next sections will discuss in depth this animal model of PD.

5.1. The 6-OHDA-lesioned rat

The 6-OHDA-lesioned rat model is the classical and most widely used toxin-based animal model used for both *in vitro* and *in vivo* investigations, and its popularity may be attributed to its cost-effectiveness and minimal labour requirements (943). The catecholamine neurotoxin 6-OHDA is transported into cell bodies and fibres of both dopaminergic and noradrenergic neurons and destroys these neurons on the ipsilateral side relative to its administration (944). Its neurotoxic effect is based on the inhibition of mitochondrial respiratory enzymes (945), which causes oxidative stress and mitochondrial damage (946). Subsequently, these neurons can no longer exert their normal physiological functions and ultimately, die (947). As 6-OHDA has poor penetration across the BBB, it is injected intra-cerebrally into the site of interest (948). Furthermore, the neurotoxin is only relatively selective since it has the ability to destroy dopaminergic and noradrenergic fibres, a concern when injecting into the medial forebrain bundle (MFB), which includes ascending fibre systems from the raphe pontine nucleus and locus coeruleus, respectively (946). Therefore, to achieve the selective destruction of dopaminergic neurons and spare noradrenergic fibres, subjects are pre-treated with despiramine, a noradrenaline transporter blocker, prior to the 6-OHDA lesion (946).

The neurotoxin 6-OHDA can be injected virtually anywhere along the nigrostriatal tract and a seminal study found that 6-OHDA could be injected at the origin of the ascending nigrostriatal DA pathway to produce a nearly complete depletion of DA in the ipsilateral striatum (949). Injection of 6-OHDA into the MFB leads to a close to complete nigrostriatal lesion, with up to 100% loss of dopaminergic terminals in the striatum (950). Generally, 6-OHDA is injected into one of three target sites: SNc, MFB or the striatum (951), where it induces varying degrees of DA denervation (946), depending on what the experimental end-point is. To model PD, the animal model should recapitulate both dopaminergic cell loss and behavioural deficits associated with idiopathic PD (944). For instance, injections of the toxin at the origin of nigrostriatal DA bundle produce large (>97%) DA depleting lesions, which model an advanced stage of PD, while injections in the terminal field of the nigrostriatal pathway produce a partial and slower progressing lesion (952, 953) that would be of greater interest for neuroprotective interventions.

In the present study, it is of interest to study the effects of a therapeutic intervention that alleviate AIMs in the 6-OHDA-lesioned rat model, which resembles more advanced stages of PD. Thus, the injection of 6-OHDA into the MFB, which results in severe DA denervation, was more appropriate for the following experiments.

5.1.1. Injection of 6-OHDA into the MFB

The nigrostriatal dopaminergic pathway consists of the A9 cell group, located in the SNc (944). Axons of these neurons run along the MFB and terminate in the dorsal striatum (954). In PD, the dopaminergic neurons of the A9 cell group undergo extensive loss and cause a dramatic decline in striatal DA, leading to motor impairments (955). Unilateral injection of the MFB causes total destruction of A9 and A10 cell groups (956), resulting in near total depletion of DA in the ipsilateral striatum, denervation supersensitivity of post-synaptic DA receptors in ipsilateral striatum and the characteristic rotational behaviour in response to both D-amphetamine and apomorphine (944).

Although bilateral 6-OHDA lesions of the MFB more closely resemble the bilateral pathology observed in idiopathic PD (944, 957), the survival and problems with daily living such as swallowing and adipsia of the animals limits the use of the bilateral model (958).

Unilateral lesioning the MFB causes can asymmetry in the motor behaviour of rats. Following lesion, rats initially tend to turn preferentially towards the side of the lesion (949, 959), a postural motor asymmetry of behaviour that may recover only slightly if depletion is near total (944). When challenged with drugs acting on the DA system, rat displays active rotational behaviour due to the imbalance in DA activity between two striata, which causes rotational asymmetry such that the animal rotates away from the side of greater activity (931).

5.1.2. Compensation

An important consideration is that by inducing a DA-depleting lesion in rats, animals transition from a normal state to a state of severe parkinsonian symptoms (944). It is possible that compensatory mechanisms come into action to antagonize these neurobiological deficits, for instance, in the rat, PD symptoms may recover to some extent over time (960-963), whereas human idiopathic PD is a progressive disease with PD symptoms worsening with time (944). In

addition, unilateral 6-OHDA-lesioned rats shows compensatory serotonergic hyperinnervation of striatum, an effect that is not observed in human PD (964). This animal model of PD should be interpreted with caution as it may not be predictive of changes in 5-HT receptors in PD.

6. Behavioural testing

6.1. Cylinder test

It is crucial to assess the extent of DA-denervating lesions in animal models and tests of physiological motor behaviour can be performed to estimate lesion severity. Although many interventions have demonstrated anti-dyskinetic efficacy in animal models, their action may at times be due to a motor depressant effect, which is of no benefit in the context of PD as it would exacerbate parkinsonian symptoms (965). Therefore, it is important to assess that the anti-parkinsonian efficacy of a treatment does not interfere with physiological motor behaviour. With the cylinder test, a measure of forelimb use during spontaneous exploration (966), it is possible to determine whether the efficacy of the anti-dyskinetic treatment compromises the therapeutic efficacy of L-DOPA.

The cylinder test, originally described by Schallert and Tillerson, assesses the independent use of each forelimb during explorative activity (966). This test takes advantage of the rats' innate drive to explore a novel environment by standing on their hind limbs, known as rearing, and using their forelimbs to contact the wall (965). Forelimb asymmetry is scored as independent weight bearing contacts on the cylinder wall of the ipsilateral and contralateral forepaws, as well as movements by both forepaws (944). Subsequently, investigators compute a limb use asymmetry score that expresses the performance of each limb as a percentage of total wall contacts. Normal rats use the right and left forepaw indifferently in this test, whereas unilaterally 6-OHDA-lesioned rats use the forepaw contralateral to lesion in about 10-30% of total supporting wall contacts (885). Use of the paw ipsilateral to lesion in $\geq 70\%$ of all wall contacts is indicative of 88% nigrostriatal dopaminergic denervation and is used as a cut-off threshold for animal inclusion in studies (967).

The cylinder test offers several advantages as a measure of physiological motor performance (965). On a conceptual level, the test is a true measure of spontaneous forelimb use as the movements exhibited by the rat in the testing cylinder are identical to those performed

in its home cage. In addition, it is simple, objective and rapidly executed and does not require pre-training of animals or extensive manipulation. Moreover, the inter-rater reliability is very high ($r > 0.95$), even with relatively inexperienced raters (966). Some investigators have argued against drug-induced rotational behaviour as a reliable indicator of nigrostriatal DA depletion (948, 968). Accordingly, the cylinder test is a drug-free test sensitive to the disrupting effects of DA denervating lesions and allows the animals to remain in a drug-naïve state, which is critical to the success of certain experimental paradigms. In addition, 6-OHDA-lesioned rats show robust forepaw asymmetry and the cylinder test is sensitive to the motor improvement produced by anti-parkinsonian compounds (see section: administration of ondansetron does not impair the therapeutic efficacy of L-DOPA in the cylinder test, page 96). Treatment with L-DOPA improves performance in the cylinder test, *i.e.* greater use of the contralateral parkinsonian forepaw, and is disrupted by the appearance of L-DOPA induced AIMs. The main drawback of the cylinder test is its relatively narrow dynamic range (from 20-50% of the contralateral forepaw) and with many repetitions of the test, the animal loses interest in exploring the novel environment and the total number of wall contacts gradually declines (965). Therefore, it is important to limit the frequency of testing sessions to a twice-weekly or weekly basis to avoid compromising test sensitivity.

6.2. ALO AIMs

Traditionally, experimental studies of LID were exclusively performed in nonhuman primates. Existing literature on LID in the parkinsonian rodent assumed that the responsiveness to L-DOPA could only be measured with tests of contralateral rotation and many investigators expressed scepticism about modelling PD symptoms and treatment related dyskinesia in rodents (885). In fact, it was believed that only primates could show the repertoire of movement disorders displayed by patients (604), which could not be evaluated in rats. For decades, the Ungerstedt model was the gold standard model of rodent research (931) and used as a screen for potential anti-parkinsonian agents where, following administration of drugs that stimulate dopamine receptors, the animal turns away the site of lesion, *i.e.* displays contralateral turning (969).

In the late 1990s, Cenci and collaborators were the first to develop the AIMs rating scale in the L-DOPA-treated 6-OHDA-lesioned rat (602). In addition to the sensitized rotational

behaviour displayed by dyskinetic rats, they also exhibit abnormal movements and postures affecting the trunk, limb and orofacial muscles contralateral to lesion. The quantification of AIMs in rats gradually replaced the test of contralateral rotation (970, 971), which does not always correlate with the development of dyskinesia (885). L-DOPA-induced AIMs in rats present functional and phenomenological analogies to LID in PD patients (965). Phenotypically, the movements are complex and involve different muscle groups that include tonic torsion of the upper trunk and neck and are associated with repetitive head movements and rapid flexion of the forelimb, similar to the choreiform on-dyskinesia exhibited by patients (965). Functionally, these movements are involuntary and disabling as are LID in PD patients (965).

The standard L-DOPA treatment to induce dyskinesia consists of a priming phase of single daily intraperitoneal injections of L-DOPA for two to three weeks, depending on the dose of L-DOPA administered, followed by a maintenance of priming by two to four injections per week to maintain stable dyskinesia over long-term (965). Once established, the brain maintains its primed state, and even after stopping the initial therapy, a single acute L-DOPA or DA agonist administration can elicit LID at nearly the same severity (440). With a daily dose of 6-10 mg/kg L-DOPA (combined with 15 mg/kg benserazide), approximately 50 to 80% of the treated rats develop AIMs by the end of the treatment period (965). The latency for the initial appearance of dyskinesia varies amongst individual rats (602) and the incidence of dyskinesia can be boosted and its latency shortened with higher L-DOPA doses (972).

To quantify drug-induced AIMs, rats are individually placed in a transparent cylinder and observed for two minutes every twenty minutes for three hours following the injection of L-DOPA (885, 973). Rat AIMs are classified into four subtypes based on their topographic distribution: axial AIMs, which are dystonic postures or choreiform twisting of the neck and body towards the side contralateral to lesion; limb AIMs, which are abnormal purposeless movements of the forelimb and digits contralateral to lesion; orolingual AIMs are empty jaw movements and contralateral tongue protrusions; and locomotive AIMs which are increased locomotion with contralateral side bias (946). Although locomotive AIMs, contralateral rotation, are part of the dyskinetic expression, it is not a specific predictor of dyskinesia as it may result from increased locomotor activity in rats that display sensorimotor asymmetry, as discussed above. The relative presentation of different AIM subtypes may differ amongst the

animals but is very consistent in the same animal upon repeated testing (965). To increase the sensitivity of the test, the authors included an additional scale based on the amplitude of dyskinetic movements (Table IV, page II). The amplitude scale is scored simultaneously with the duration scale as described above (based on the duration and frequency). Lastly, only movements that are phenomenologically distinguishable from stereotypic behaviour for the rat are classified as dyskinetic, whereas enhanced manifestations of normal motor activities (grooming, gnawing, rearing and sniffing) are not included in the rating (974).

In seminal studies, Lundblad and colleagues compared the effects of drugs with varying dyskinesigenic potential on motor performance in the 6-OHDA-lesioned rat model (885). They reported an attenuation of AIMs by non-dopaminergic compounds with proven anti-dyskinetic efficacy in patients and/or primates. In contrast, AIMs were not induced by anti-parkinsonian treatments with low dyskinetic potential in primates (975). These studies were the first demonstrations that clinically-relevant measures of parkinsonian akinesia and dyskinesia could be obtained in rats (885). Furthermore, it demonstrated that rat AIMs share similar pharmacological properties to primate models of LID, particularly with respect to modulation of neurotransmitter systems to control the expression of dyskinesia (885, 973, 976-978).

Another study compared the effects of various non-dopaminergic compounds on both L-DOPA-induced AIMs and L-DOPA-induced motor improvement in 6-OHDA lesioned rats (973). Again, the AIMs model demonstrated a high degree of predictive validity as interventions with proven anti-dyskinetic action in primate models and in PD patients also modulated rodent axial, limb and orolingual (ALO) AIMs. However, treatments that specifically alleviated the severity of trunk, limb and orofacial dyskinesia did not necessarily reduce locomotive AIM scores. Treatments that specifically produced a decrease in ALO AIMs scores neither interfered with normal rat behaviour (*e.g.* locomotion, exploration, grooming) nor affected the locomotive AIM scores or rotarod activity.

7. Objectives and hypotheses

As presented above, growing evidence supports a pivotal role of the 5-HT system in the pathogenesis of LID. However, no studies have evaluated the effect of 5-HT₃ receptors on dyskinesia expression. Thus, the present study seeks to determine and validate the efficacy of

5-HT₃ receptor antagonism as a new therapeutic strategy to alleviate L-DOPA-induced AIMs in the 6-OHDA-lesioned rat model of PD. More specifically, we hypothesize that:

1. 5-HT₃ receptor blockade reduces the severity of established L-DOPA-induced AIMs;
2. 5-HT₃ receptor blockade attenuates the priming process that leads to the development of L-DOPA induced AIMs;
3. 5-HT₃ receptor blockade does not impair the therapeutic efficacy of L-DOPA on parkinsonian features.

To validate these hypotheses, we will meet the following aims:

1. To determine the effect of acute challenges of the highly-selective 5-HT₃ antagonist ondansetron at alleviating established L-DOPA-induced AIMs;
2. To determine the effect of selective 5-HT₃ receptor blockade with ondansetron on the development of L-DOPA-induced AIMs, in the context of a *de novo* study;
3. To assess whether the anti-dyskinetic benefit of 5-HT₃ antagonism with ondansetron is achieved without compromising the anti-parkinsonian action of L-DOPA.

Positive outcomes of the proposed experiments would provide support for 5-HT₃ receptor antagonism as a new and effective therapeutic approach to alleviate L-DOPA induced dyskinesia in the 6-OHDA lesioned rat model of PD. Moreover, positive results would demonstrate the potential of a new target to achieve an anti-dyskinetic effect without impairing L-DOPA anti-parkinsonian action. Given that ondansetron and other 5-HT₃ receptor antagonists are clinically available and well tolerated, positive outcomes could rapidly lead to Phase IIa clinical trials, and enhance the quality of life of PD patients.

II. Material and methods

Animals

Adult female Sprague-Dawley rats (225 – 250 g, Charles River, Saint-Constant, Canada) were group-housed in a temperature, humidity- and light-controlled environment (under 12-h light/dark cycle, on 07:00) with free access to food and water. Experimental protocols were approved by Centre de Recherche du Centre Hospitalier de l'Université de Montréal Animal Care Committee in agreement to guidelines established by the Canadian Council on Animal Care. Upon arrival, rats were left undisturbed to acclimatize to the housing conditions for at least 5 days before experiments.

Dose-finding pharmacokinetic study

Based on doses of ondansetron used in the literature (979), a preliminary dose-finding pharmacokinetic (PK) study was conducted to determine clinically-relevant plasma levels of ondansetron in the rat. Blood was collected from animals ($N = 2$) by jugular vein puncture using a sparse sampling technique, as previously described (980), serial blood samples of 150 μL were collected prior to, and at the following time points: 2 min, 5 min, 15 min, 30 min, 1 h, 2 h, 3 h, and 4 h after subcutaneous administration of ondansetron 0.01 mg/kg. Samples were gently inverted and placed on ice pending centrifugation (1500g for 10 minutes at 4°C). Following centrifugation, aliquots of approximately 80 μL of plasma were stored at -80°C until analysis. The analytical method to quantify ondansetron in the plasma consisted of protein precipitation followed by liquid chromatography-tandem mass spectrometry (LC-MS/MS), which was done in collaboration with Dr Francis Beaudry and Ms Fleur Gaudette from Faculté de Médecine Vétérinaire de l'Université de Montréal and Centre de Recherche du Centre Hospitalier de l'Université de Montréal.

Unilateral 6-OHDA lesion

Animals were rendered hemi-parkinsonian by unilateral injection of 6-OHDA into the right MFB as previously described (967). Animals were pre-treated with pargyline (5 mg/kg) and desipramine (10 mg/kg) 30 min prior to surgery. Under general anaesthesia (3% isoflurane in 95% O₂, 5% CO₂), rats were positioned onto a stereotaxic frame (Kopf Instruments, Tujunga, USA). 6-OHDA (7 mg/mL) was dissolved in 0.02% ascorbic saline. The 6-OHDA solution (2.5

μL) was injected into the right MFB using a 10 μL Hamilton microsyringe with a 30-gauge needle (at a flow rate of 0.5 μL/min) at the following coordinates: anteroposterior -2.8 mm, lateromedial -2.0 mm and dorsoventral -9.0 mm from Bregma and skull surface (981), with the incisor bar set at 3.3 mm below ear bars. The injection was done over 5 min and the needle was left in place for an additional 5 min before slowly retracting the needle to avoid reflux. At the end of surgery and for two additional days post-op, animals received carprofen (5 mg/kg) as analgesic treatment.

Cylinder test

Following a 14-day post-lesion recovery period, animals underwent the cylinder test to assess the extent of dopaminergic degeneration (966). Rats were placed in a transparent cylinder (14 cm diameter × 18 cm height) and recorded for 10 min. A mirror was placed behind the cylinder to enable the evaluator to count forelimb movements when the animal was turned away from the camera. Several behaviours were scored to determine the extent of forelimb-use asymmetry displayed by the animal during the 10-min period and analysed *post hoc*. The first limb to contact the wall during a rear or weight-shifting movement was scored as an independent wall placement for that limb. A subsequent placement of the other limb on the wall while maintaining the initial movement was scored as a “both” movement. A simultaneous placement of both forepaws on the walls was also considered a “both” movement. Another wall movement score was attributed only if both paws were removed from the vertical surface. Only animals exhibiting preferential use of the un-lesioned forelimb in $\geq 70\%$ of the rears were selected to undergo further behavioural pharmacological testing. As mentioned above, this rearing asymmetry score indicative of $\geq 88\%$ striatal dopamine depletion (967).

Drug treatments

Despiramine hydrochloride, pargyline hydrochloride, 6-OHDA hydrobromide, L-DOPA methyl ester, benserazide hydrochloride and ondansetron hydrochloride were purchased from Sigma-Aldrich (St Louis, USA). All drugs were dissolved in saline (sodium chloride 0.9% w/v) except for 6-OHDA and ondansetron hydrochloride, which were dissolved in ascorbate-saline and dimethyl sulfoxide at 100 mg/mL, respectively, and the latter was diluted to the appropriate concentrations in saline. All solutions were subcutaneously injected in a volume of 1.0 mL/kg

body weight. In the acute challenge study, treatments were randomized according to a Latin square design and behavioural testing sessions were separated by at least 48 h of drug washout.

Experimental design

The experimental design for the acute challenges and *de novo* studies is described in Figure 3, page 70.

Acute challenges of ondansetron study

6-OHDA lesion surgery was performed on 35 Sprague-Dawley rats. Following assessment of parkinsonism, 18 lesioned animals (~ 51%) exhibiting severe rearing asymmetry were selected and were primed daily with L-DOPA/benserazide (10/15 mg/kg, from now on referred to as L-DOPA) for 14 days to elicit stable and reproducible AIMs. Once AIMs were expressed, on days on behavioural testing, rats were administered L-DOPA (6/15 mg/kg) in combination with ondansetron (0.0001, 0.001, 0.01 0.1 and 1mg/kg) or vehicle, and AIMs were assessed, as described below (Section: ratings of AIMs, page 71), by a blinded rater.

***De novo* ondansetron study**

In another set of experiments, rats were rendered hemi-parkinsonian by 6-OHDA injection in the MFB as described above. Following recovery and after assessment of the extent of lesion, rats were administered a once daily treatment of L-DOPA (6/15 mg/kg) in combination with ondansetron (0.0001 mg/kg, group 1, or 0.001 mg/kg, group 2, both $n = 9$) or vehicle (group 3, $n = 7$) for 22 days. ALO AIMs were assessed on days 1, 8, 15 and 22 by an experimenter blinded to treatment conditions. After a 3-day washout period, animals were administered an acute 6/15 mg/kg L-DOPA challenge and AIMs severity was assessed.

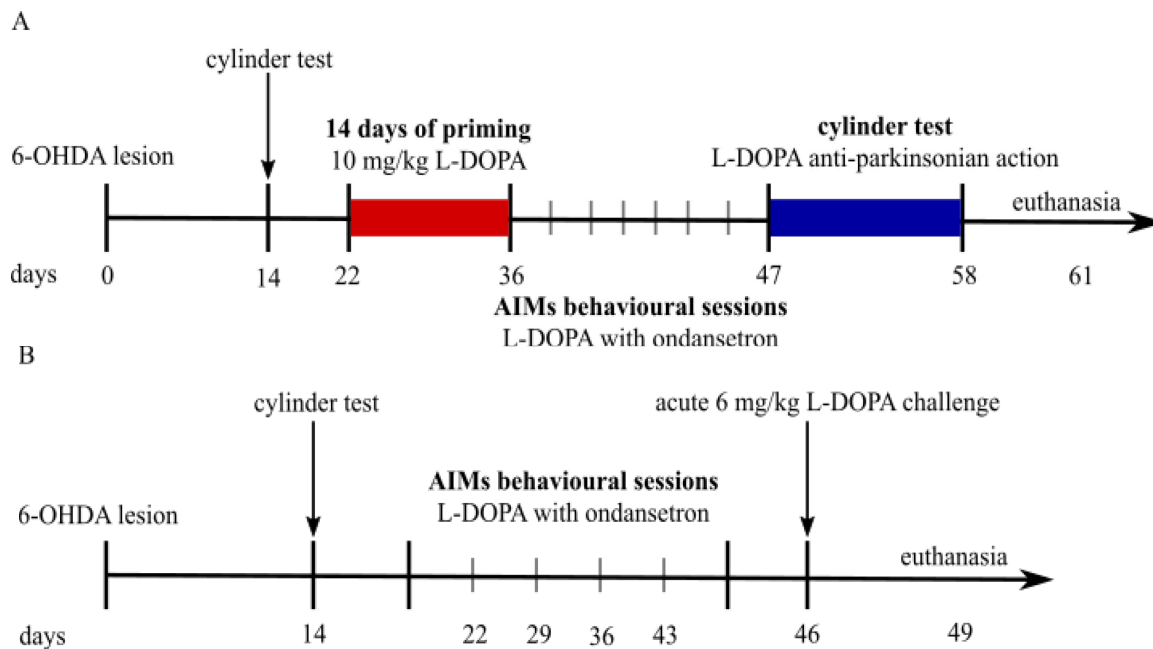


Figure 3: Schematic representation of the experimental design. A. Timeline of the acute challenge experiments. 6-OHDA lesioned animals underwent a L-DOPA priming phase to induce dyskinesia and the effect of acute ondansetron on the severity of ALO AIMs was evaluated. **B.** Timeline of the de novo experiments. Animals received treatment concurrently as their first L-DOPA dose, and this treatment regimen was maintained daily with weekly assessments of the progression of ALO AIMs, followed by an acute L-DOPA challenge after a washout period.

Ratings of AIMs

On days of behavioural scoring, after injection of L-DOPA, rats were put in individual glass cylinders and observed for 2 minutes every 20 minutes over a 180 min testing session, as previously described (967). The severity of dyskinesia was evaluated using a validated rat AIMs scale, where ALO AIMs were each scored (885). Each AIMs subtype was rated on a duration severity scale from 0 to 4 (Table III, page I) in each monitoring interval where: 0 = no dyskinesia; 1 = occasional signs of dyskinesia; 2 = frequent signs of dyskinesia; 3 = continuous dyskinesia but interrupted by external stimuli and 4 = continuous dyskinesia not interrupted by external stimuli. The amplitude of AIMs was rated from 0 to 4 (Table IV, page II). Axial AIMs are the twisting of the neck and upper body toward the contralateral side to the lesion and amplitude are rated according to the following scale: 1= sustained deviation of the head and neck at $\sim 30^\circ$ angle; 2 = sustained deviation of the head and neck at an angle of 60° or more; 3

= sustained twisting of the head, neck and upper trunk at an angle greater than 60° but up to 90° and 4 = sustained twisting of the head, neck and trunk at an angle greater than 90°, causing the rat to lose balance from a bipedal position. Limbs AIMS consist of jerky or dystonic movements of the contralateral limb and are rated as follows: 1 = tiny movements of the paw around a fixed position; 2 = movements leading to visible displacement of the limb; 3 = large displacement of the limb with contraction of shoulder muscles and 4 = vigorous limb displacement of maximal amplitude, with concomitant contraction of shoulder and extensor muscles. Orolingual AIMS consist of movement of jaw muscles and tongue protrusions and amplitude are rated as: 1 = twitching of facial muscles accompanied by small masticatory movements without jaw opening; 2 = twitching of facial muscles accompanied by masticatory movements that occasionally result in jaw opening; 3 = movements with broad involvement of facial muscles and masticatory muscles, with frequent jaw opening and occasional tongue protrusion and 4 = involvement of all of the above muscles to the maximal possible degree. The ALO AIMS score represents the sum of axial, limbs and orolingual AIMS scores during the behavioural session, and this expression of AIMS scores is sensitive to the anti-dyskinetic effects of drugs used in the clinic (885, 973). The axial limbs (AL) AIMS score represents the sum of axial and limbs AIMS scores on all monitoring periods.

Assessment of L-DOPA anti-parkinsonian action

To assess whether the anti-dyskinetic effect of ondansetron affects the therapeutic efficacy of L-DOPA, preference for the un-lesioned forelimb in the cylinder test was evaluated as described above. Rats used in the acute challenge study underwent a 3-day washout period, after which they were administered a low dose of L-DOPA (3/15 mg/kg), to avoid triggering AIMS, in combination with vehicle or ondansetron (0.0001 0.001, 0.01, 0.1, 1 mg/kg). 45 min later, at peak anti-parkinsonian action, animals underwent the cylinder test, in which the number of rears of each paw was counted, *post hoc*, by a treatment-blinded experimenter.

Perfusions

Perfusions were performed at least 48 h post-administration of L-DOPA. Under general anaesthesia (4% isoflurane in 95% O₂, 5% CO₂), rats were euthanised by exsanguination by

transcardial perfusion with saline (982). After the toe pinch-response was used to determine depth of anaesthesia, animals were secured in a supine position on the work surface. Using sharp scissors, incisions were made along the thoracic midline and lateral to the ventral ribcage to expose the thoracic field. The rib cage was then cut through to open up the thoracic cavity and, using blunt scissors, the diaphragm was separated from the chest wall and the tissue connecting the sternum to the heart was cleared. The heart was subsequently secured with blunt forceps before an 18-gauge perfusion needle was inserted into the left ventricle and the needle position was secured by clamping a hemostat near the point of entry. An incision to right atrium was then made with scissors and animals were infused with a steady flow of their corresponding body weights of 0.9% saline (until the fluid exiting the right atrium was clear). Brains were rapidly removed, flash-frozen at -55°C in isopentane and stored at -80°C until collection of striatal tissue.

LC-MS/MS analysis for dopamine and its metabolites

The extent of nigrostriatal lesion was determined LC-MS/MS (983). A 30- μm diameter tissue punch of the left and right striata was obtained and deposited into separate 1.5 mL sterile microcentrifuge tubes and stored at -80°C until LC-MS/MS analysis (984). A piece of comparable size of cerebellum was also dissected from each brain as a control (985). Quantification of the biogenic amine DA and its metabolites 3,4-dihydroxyphenylacetic acid (DOPAC) and HVA in the striatal punches is currently being conducted in collaboration with Dr Lehka Sleno from Université de Québec à Montréal and the results will be presented in a peer-review article.

Statistical Analysis

Pharmacokinetic study

Calculations of the peak plasma concentration (C_{max}), time of C_{max} (t_{max}) and half-life ($t_{1/2}$) were done with Microsoft Office Excel (Microsoft Corporation, Redmond, USA) (986).

Cylinder test

Data from the cylinder test were graphed as the mean \pm standard error of the mean (SEM) and were analysed using one-way analysis of variance (ANOVA) with the Greenhouse-Geisser correction; *post hoc* comparisons with the lesion control and L-DOPA/vehicle control groups were performed using Tukey's *post hoc* test.

Taking into consideration the variation in standard deviations across treatment conditions in Figures 10B and 10C, a measure of effect size was provided using Glass' delta (see Tables V and VI in Appendices III and IV). This estimate is recommended for unequal variances and under the assumption that any measure on control is untainted by the effect, the standard deviation of the control group is used to standardize the differences between means to minimize bias (987) as calculated by the formula for Glass' delta (Figure 11 in Appendix V).

Acute challenges of ondansetron study

AIMs scores were expressed as the median with interquartile interval. In the acute study, comparisons of AIM scores used the cumulative score over the entire testing session or the peak of L-DOPA action, the interval from 40-120 min post-drug administration. Results were analysed using nonparametric Friedman test, followed by Dunn's *post hoc* test.

***De novo* ondansetron study**

The *de novo* AIMs timecourse analysis was performed using two-way ANOVA, followed by Tukey's *post hoc* test. Data from the *de novo* challenge AIMs scores underwent a squareroot transformation (988), and were subsequently analysed by one-way ANOVA, followed by Tukey's *post hoc* test.

The threshold for statistical significance was assigned at $P < 0.05$. Statistical analyses were performed with GraphPad Prism 7.03 (GraphPad Software Inc., La Jolla, California, USA).

III. Results

Pharmacokinetic profile of ondansetron

As illustrated in Figure 4 (page 76), the plasma levels of subcutaneous administration ondansetron are mapped over a time course. The ondansetron PK parameters assessed in the preliminary dose-finding ondansetron PK are summarised in Table II (page 76). C_{\max} was 2.31 ng/mL while t_{\max} and $t_{1/2}$ were 15 and 39 min, respectively.

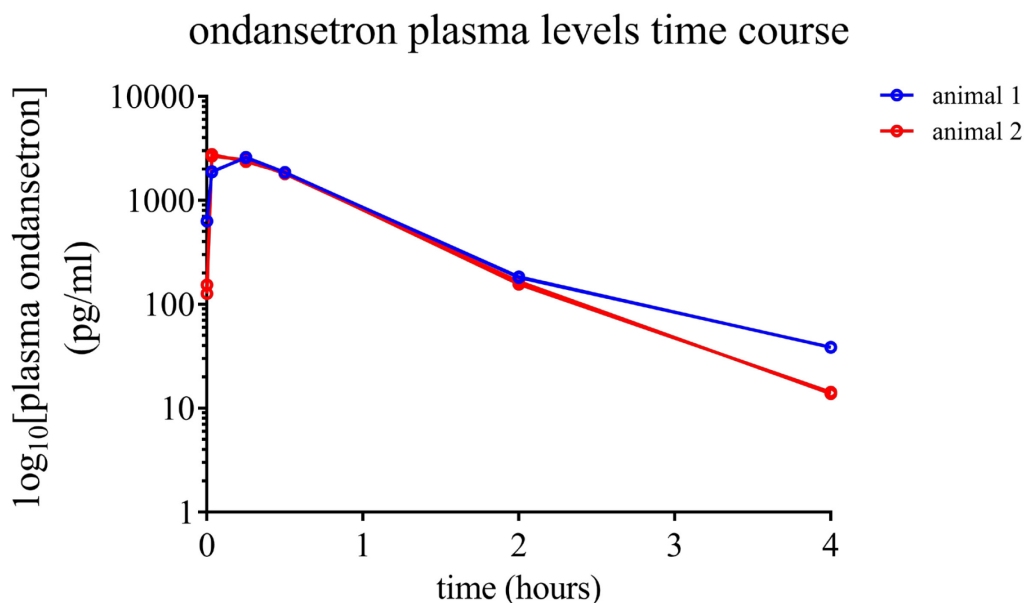


Figure 4: Plasma levels of ondansetron in a preliminary pharmacokinetic study. Logarithmic (\log_{10}) time course of ondansetron plasma levels following subcutaneous administration of 0.01 mg/kg ondansetron ($n = 2$). Data are presented as the mean.

Table II: Ondansetron pharmacokinetic parameters in the 6-OHDA-lesioned rat

	ondansetron
C_{\max} (ng/mL)	2.31
t_{\max} (min)	15
$t_{1/2}$ (min)	39

C_{\max} : maximal plasma concentration; t_{\max} : time at maximal plasma levels $t_{1/2}$: plasma half-life. Ondansetron 0.01 mg/kg was administered subcutaneously to the animals ($n = 2$). Data are presented as the mean.

Extent of dopaminergic denervation assessed in the cylinder test

Following 6-OHDA lesion, only animals that displayed marked forelimb asymmetry, i.e. used the right (un-lesioned) forepaw to initiate \geq than 70% of wall contacts during the cylinder test, a score that is indicative of \geq 88% striatal DA depletion (967), were selected for the behavioural studies. As shown in Figure 5A (page 77), animals that underwent the acute challenges of ondansetron study displayed forelimb asymmetry ($F(2, 39) = 296.6, P < 0.0001$, one-way ANOVA) with marked preferential use of the right forepaw in 83% of wall contacts when compared to 0.4% with the left forepaw and 15% with both forepaws, respectively (both $P < 0.0001$, Tukey's *post hoc* test). As illustrated in Figure 5B (page 77), in the *de novo* ondansetron study, animals also displayed marked forelimb asymmetry ($F(2, 78) = 1017, P < 0.0001$, one-way ANOVA), and preferred the use of the right forepaw in 85% rears when compared with 0.8% and 15% of rears using the left forepaw and both forepaws, respectively (both $P < 0.0001$, Tukey's *post hoc* test).

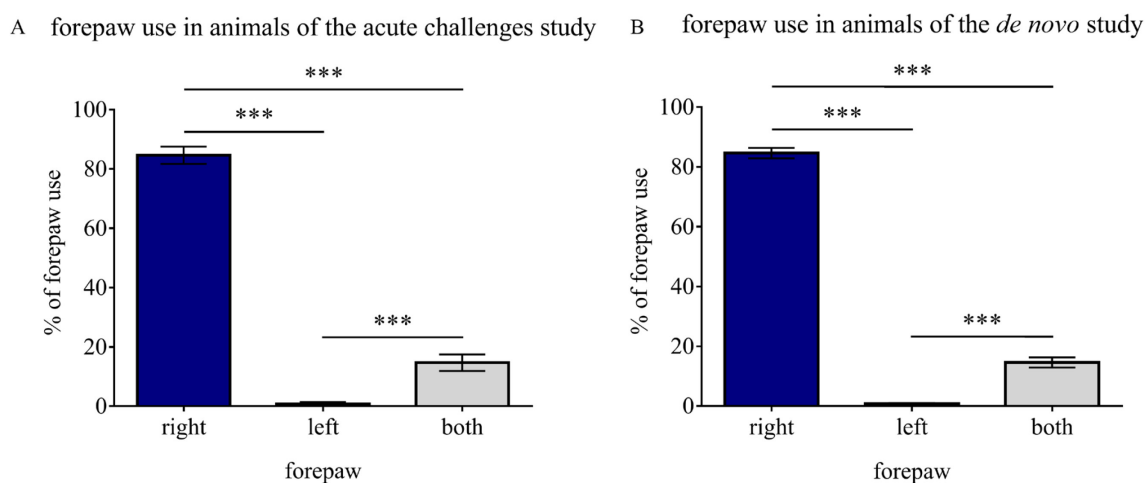


Figure 5: Performance in the cylinder test in drug-naïve lesioned animals. **A.** Animals ($n = 18$) selected to undergo acute challenges of ondansetron show a marked preference for the un-lesioned (right) forepaw in 83% of rears compared with 0.4% and 17% of rears using the lesioned (left) forepaw and both forepaws, respectively. **B.** In the *de novo* ondansetron study, animals ($n = 24$) prefer the right forepaw in 85% of rears while the left forepaw and both forepaws, only accounted for 0.8% and 15% of rears, respectively. Data are presented as the mean \pm SEM. **: $P < 0.01$, ****: $P < 0.0001$.

Acute challenges of ondansetron at 0.0001 mg/kg significantly alleviated the severity of established AIMs

In the acute challenges of ondansetron experiments, daily administration of L-DOPA 10 mg/kg for two weeks to 6-OHDA-lesioned rats induced stable and reproducible dyskinetic behaviour. Animals subsequently received acute challenges of ondansetron or vehicle in combination with L-DOPA and AIMs severity was assessed.

Duration of axial AIMs

As shown in Figure 6A and 6B (page 80), administration of ondansetron in combination with L-DOPA reduced the duration of cumulative and peak axial AIMs (Friedman Statistic [FS] = 32.16, $P < 0.001$ and FS = 21.14, $P < 0.01$, respectively). Thus, when ondansetron 0.0001 mg/kg and 0.001 mg/kg was added to L-DOPA, the duration of cumulative axial AIMs was reduced by 55% and 52%, respectively, when compared to L-DOPA/vehicle ($P < 0.001$ and $P < 0.01$, Dunn's *post hoc* test, Figure 6A, page 80). The addition of ondansetron 0.0001 mg/kg also reduced peak axial AIMs duration when compared with vehicle by 49%, compared to L-DOPA alone ($P < 0.01$, Dunn's *post hoc* test, Figure 6B, page 80). Peak axial AIMs duration was also reduced with treatment of ondansetron 0.001, 0.01 and 1 mg/kg compared to vehicle but did not reach statistical significance.

Duration of limbs AIMs

As illustrated in Figure 6C (page 80), adding ondansetron to L-DOPA resulted in a significant reduction in the duration of cumulative limbs AIMs (FS = 20.76, $P < 0.001$). Thus, administration of ondansetron 0.0001 and 0.001 mg/kg in combination with L-DOPA reduced the duration of cumulative limbs AIMs was reduced by 49% and 41%, respectively, when compared with L-DOPA/vehicle ($P < 0.01$ and $P < 0.05$, Dunn's *post hoc* test). Administration of ondansetron did not significantly diminish the duration of peak limbs AIMs (FS = 10.71, $P > 0.05$, Figure 6D, page 80). The duration of limbs AIMs scores was also reduced with treatment of ondansetron 0.001, 0.01 0.1 and 1 mg/kg compared to vehicle but did not reach statistical significance.

Duration of orolingual AIMs

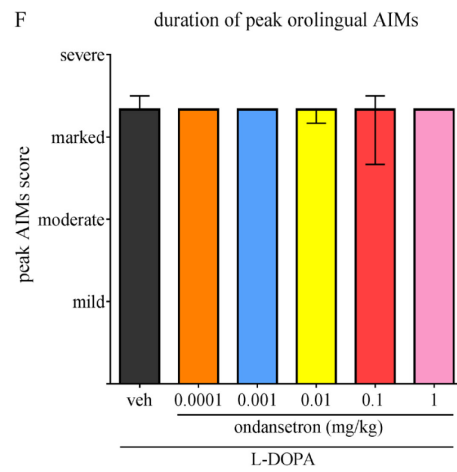
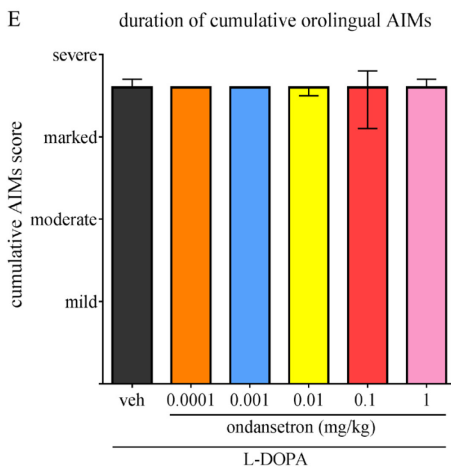
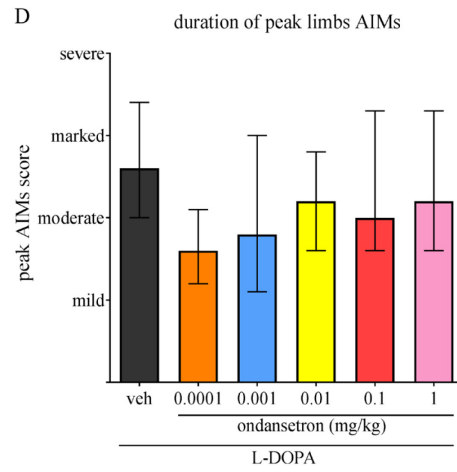
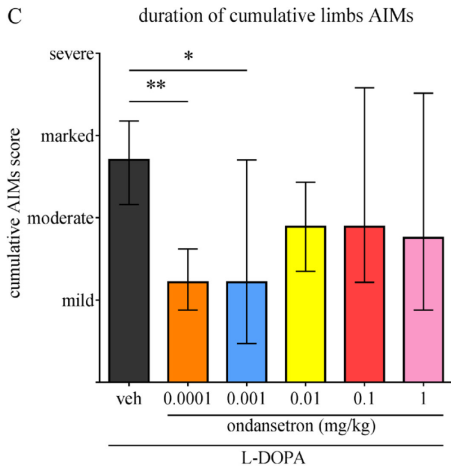
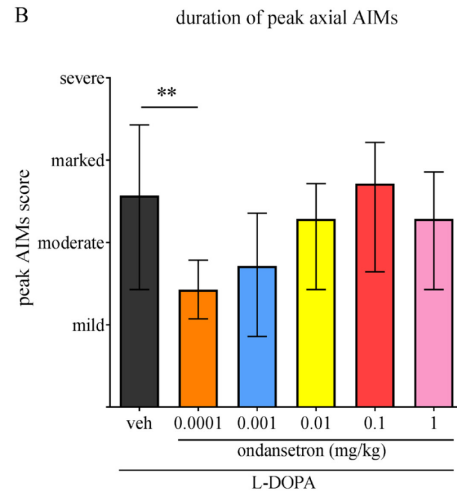
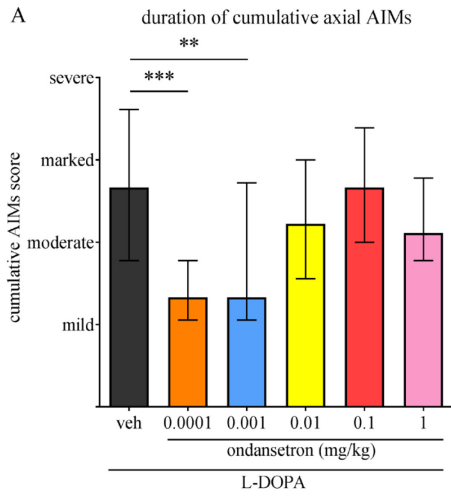
As illustrated in Figures 6E and 6F (page 80), the severity and duration of cumulative and peak orolingual AIMs was comparable between the doses of ondansetron (0.0001, 0.001, 0.01, 0.1 and 1 mg/kg) and vehicle (FS = 5.826, $P > 0.05$ and FS = 4.833, $P > 0.05$, respectively).

Duration of AL AIMs

As shown in Figures 6G and 6H (page 81), the addition of ondansetron to L-DOPA resulted in a significant decrease in the duration of cumulative and peak AL AIMs (FS = 29.01, $P < 0.0001$ and FS = 19.43, $P < 0.01$, respectively). Thus, treatment with ondansetron 0.0001 and 0.001 mg/kg led to a marked decrease in the duration of cumulative AL AIMs by 57% and 47%, respectively, when compared with L-DOPA/vehicle ($P < 0.001$ and $P < 0.01$, Dunn's *post hoc* test, Figure 6G, page 81). Animals that received ondansetron 0.0001 mg/kg also exhibited a 54% reduction in the duration of peak AL AIMs, when compared with vehicle ($P < 0.001$, Dunn's *post hoc* test, Figure 6H, page 81). The duration of cumulative and peak AL AIMs scores was also diminished with administration of ondansetron 0.001, 0.01, 0.1 and 1 mg/kg compared to vehicle but this was not statistically significant.

Duration of ALO AIMs

As shown in Figures 6I and 6J (page 81), administration of ondansetron in combination with L-DOPA led to a significant reduction in the duration of cumulative and peak ALO AIMs, respectively (FS = 23.93, $P < 0.0001$ and FS = 17, $P < 0.001$). Thus, administration of ondansetron 0.0001 and 0.001 mg/kg decreased the duration of cumulative ALO AIMs by 53% and 43%, respectively, when compared with vehicle ($P < 0.01$ and $P < 0.05$, Dunn's *post hoc* test, Figure 6I, page 81). The duration of peak ALO AIMs was reduced with ondansetron 0.0001 mg/kg compared to vehicle by 51% ($P < 0.01$, Dunn's *post hoc* test, Figure 6J, page 81). In addition, cumulative and peak ALO AIMs scores were also diminished with treatment of ondansetron 0.001, 0.01, 0.1 and 1 mg/kg compared to vehicle but did not reach statistical significance.



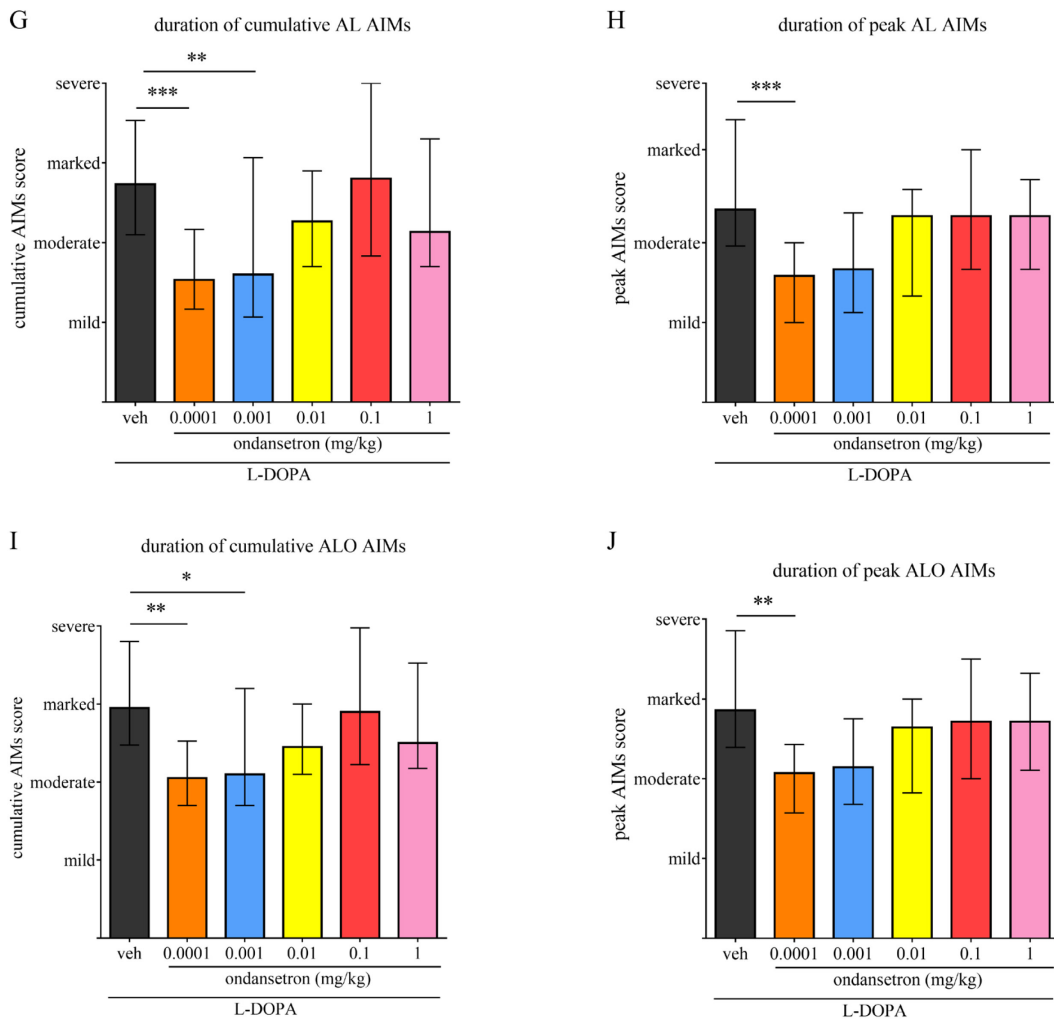


Figure 6: Effect of acute challenges of ondansetron on the duration of established L-DOPA induced AIMs. **A.** Administration of 0.0001 mg/kg ondansetron in combination with L-DOPA diminished the duration of cumulative and **B.** peak axial AIMs when compared with vehicle. **C.** Similarly, L-DOPA/ondansetron 0.0001 mg/kg resulted in a reduction in the duration of cumulative limbs AIMs, when compared with vehicle but had **D.** no effect on peak limbs AIMs when compared with L-DOPA/vehicle. **E. F.** Adding ondansetron to L-DOPA had no effect on orolingual AIMs when compared to L-DOPA/vehicle. **G.** With respect to the duration of AL AIMs, ondansetron 0.0001 mg/kg resulted in less severe cumulative AL AIMs, **H.** as well as a decrease in the duration of peak AL AIMs. **I.** The combination of ondansetron 0.0001 mg/kg and L-DOPA led to a significant reduction in the severity of cumulative ALO AIMs **J.** and a marked decrease in the duration of peak AIMs scores. Cumulative and peak duration AIMs scores are expressed as median with interquartile interval. $n = 18$ for all treatment conditions. *: $P < 0.05$; **: $P < 0.01$, ***: $P < 0.001$ and ****: $P < 0.0001$.

Amplitude of axial AIMS

As illustrated in Figures 7A and 7B (page 84), during the assessment of AIMS, ondansetron significantly decreased the amplitude of cumulative and peak axial AIMS was reduced with ondansetron (FS = 54.34, $P < 0.0001$ and FS = 46.61, $P < 0.01$, respectively). Thus, after administration of ondansetron 0.0001 and 0.001 mg/kg, the amplitude of axial AIMS diminished by 67% and 66%, respectively, when compared with L-DOPA/vehicle (both $P < 0.0001$, Dunn's *post hoc* test, Figure 7A, page 84). In addition, peak axial AIMS also diminished with 0.001 and 0.001 mg/kg ondansetron by 60% and 61%, respectively, when compared with vehicle (both $P < 0.0001$, Dunn's *post hoc* test, Figure 7B, page 84). Higher doses of ondansetron (0.01, 0.1 and 1 mg/kg) also reduced cumulative and AIMS amplitude scores but did not reach statistical significance.

Amplitude of limbs AIMS

As shown in Figures 7C and 7D (page 84), the addition of ondansetron to L-DOPA resulted in a decrease in the amplitude of cumulative and peak limbs AIMS, respectively, when compared with L-DOPA/vehicle (FS = 28.79, $P < 0.0001$ and FS = 20.48, $P < 0.01$, respectively). Thus, ondansetron 0.0001 and 0.001 mg/kg diminished the amplitude of cumulative limbs AIMS by 48% and 55%, respectively, compared to vehicle ($P < 0.01$ and $P < 0.001$, Dunn's *post hoc* test, Figure 7C, page 84). In addition, animals treated with ondansetron 0.0001 and 0.001 mg/kg exhibited a decrease in peak axial AIMS amplitude scores, compared to vehicle, by 44% and 45%, respectively (both $P < 0.05$, Dunn's *post hoc* test, Figure 7D, page 84). The amplitude of AIMS scores was also reduced with higher doses of ondansetron compared to vehicle but did not reach statistical significance.

Amplitude of orolingual AIMS

As shown in Figures 7E and 7F (page 84), treatment with ondansetron (0.0001, 0.001, 0.01 0.1 and 1 mg/kg) resulted in similar amplitude severity levels of cumulative and peak orolingual AIMS when compared with vehicle (FS = 1.744 and FS = 3.46, both $P > 0.05$).

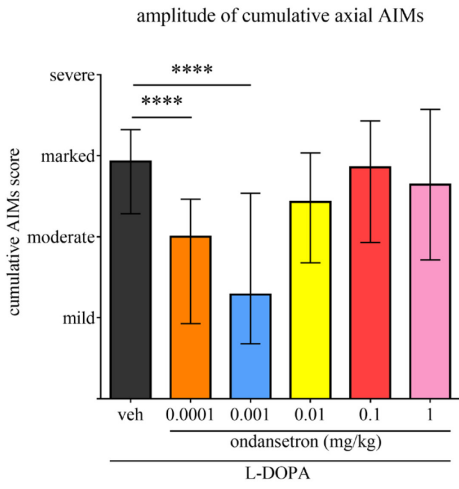
Amplitude of AL AIMs

As illustrated in Figures 7G and 7H (page 85), when administered with L-DOPA, ondansetron reduced the amplitude of cumulative and peak AL AIMs (FS = 47.25 and FS = 40.49, both $P < 0.0001$). Thus, treatment with ondansetron 0.0001 mg/kg and 0.001 mg/kg resulted in a 61% and 64% decrease in the amplitude of cumulative amplitude AL AIMs scores, respectively, when compared with vehicle ($P < 0.0001$ and $P < 0.001$, Dunn's *post hoc* test, Figure 7G, page 85). Furthermore, peak AL AIMs amplitude was diminished with ondansetron 0.0001 and 0.001 mg/kg by 60% and 58%, respectively, compared to vehicle (both $P < 0.0001$, Dunn's *post hoc* test, Figure 7H, page 85).

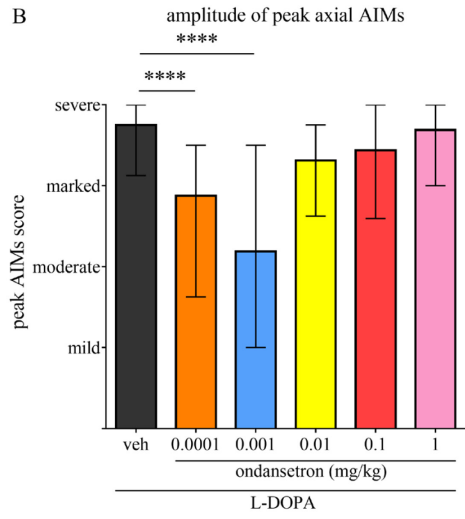
Amplitude of ALO AIMs

As shown in Figures 7I and 7J (page 85), adding ondansetron in combination with L-DOPA led to a significant reduction in the amplitude of cumulative and peak ALO AIMs, respectively (FS = 30.07, $P < 0.0001$ and FS = 22.3, $P < 0.001$). Thus, adding ondansetron 0.0001 and 0.001 mg/kg decreased the amplitude of cumulative ALO AIMs by 51% and 54%, respectively, when compared with vehicle ($P < 0.01$ and $P < 0.001$, Dunn's *post hoc* test, Figure 7I, page 85). In addition, the amplitude of peak ALO AIMs was also reduced with ondansetron 0.001 and 0.001 mg/kg, when compared to vehicle, by 51% and 44%, respectively ($P < 0.01$ and $P < 0.05$, Dunn's *post hoc* test, Figure 7J, page 85).

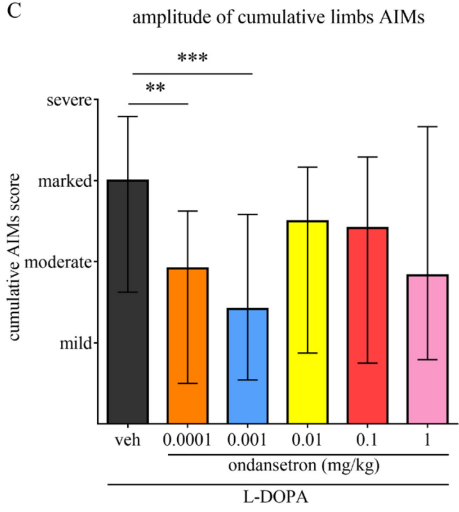
A



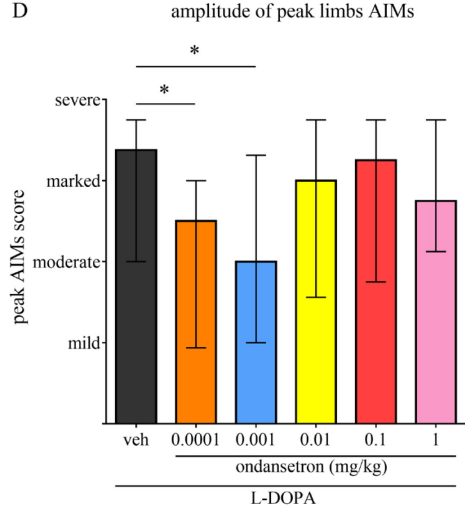
B



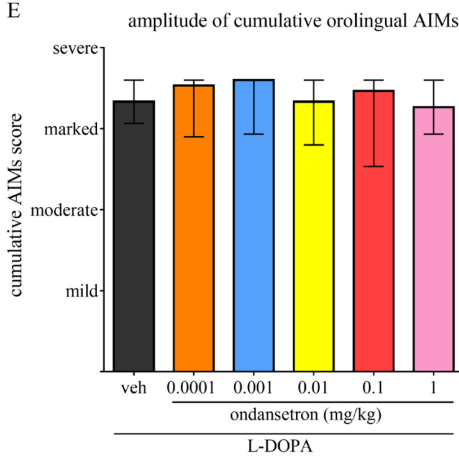
C



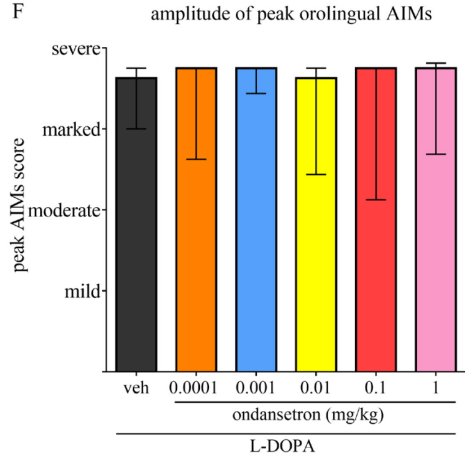
D



E



F



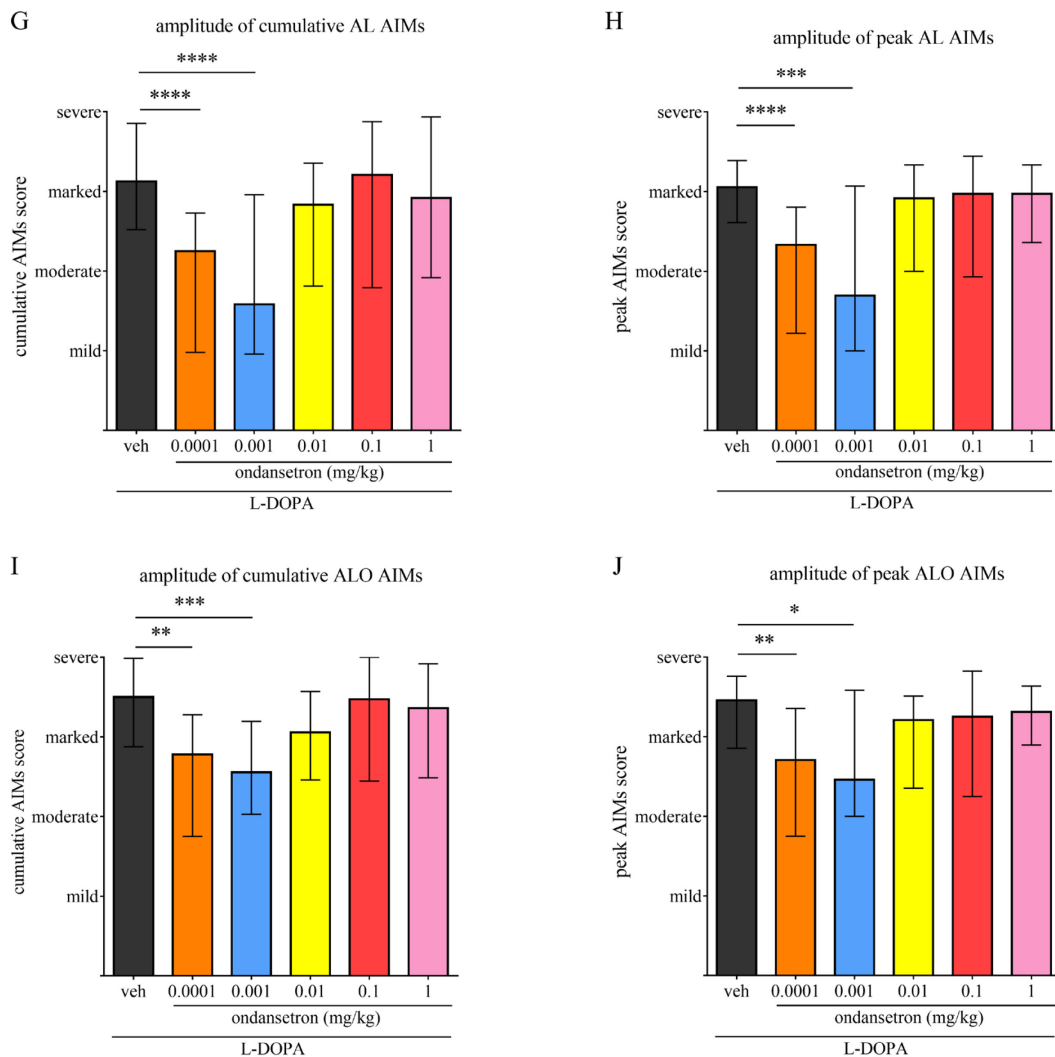


Figure 7: Effect of acute ondansetron treatment on the amplitude of established L-DOPA induced AIMs. **A.** When administered with L-DOPA, 0.0001 mg/kg ondansetron alleviated the severity of cumulative and **B.** peak axial AIMs when compared with vehicle. **C.** Similarly, ondansetron 0.0001 mg/kg/L-DOPA resulted in a reduction in the amplitude of cumulative limbs AIMs, when compared with vehicle; **D.** this effect was also observed in the lower amplitude of peak limbs AIMs when compared with vehicle/L-DOPA. **E.** The addition of ondansetron led to comparable amplitude in cumulative orolingual AIMs severity, **F.** as well as peak amplitude orolingual AIMs, when compared with vehicle. **G.** With respect to the amplitude of AL AIMs, ondansetron 0.0001 mg/kg resulted in less severe cumulative AL AIMs, and **H.** a decrease in the amplitude of peak AL AIMs. The combination of ondansetron 0.0001 mg/kg and L-DOPA resulted in a significant reduction in **I.** cumulative ALO AIMs severity and **J.** peak ALO AIMs scores. Cumulative and peak duration AIMs scores are expressed as median with interquartile interval. $n = 18$ for all treatment conditions. *: $P < 0.05$; **: $P < 0.01$, ***: $P < 0.001$ and ****: $P < 0.0001$.

***De novo* study**

***De novo* treatment with ondansetron attenuates the development of L-DOPA-induced AIMs**

Axial AIMs

As illustrated in Figure 8A (page 89), the addition of ondansetron to L-DOPA resulted in a decrease in the duration of axial AIMs during the 22-day priming phase ($F_{\text{time}}(3,88) = 2.285$, $P > 0.05$; $F_{\text{treatment}}(2,88) = 8.111$; $P < 0.001$; and $F_{\text{interaction}}(6,88) = 0.354$, $P > 0.05$, two-way ANOVA). On day 8, animals treated with 0.0001 mg/kg ondansetron/L-DOPA exhibited a 53% reduction in the duration of axial AIMs when compared with the vehicle/L-DOPA group ($P < 0.05$, Tukey's *post hoc* test). The amplitude of axial AIMs was significantly different across the three treatment groups ($F_{\text{time}}(3,88) = 1.29$, $P > 0.05$; $F_{\text{treatment}}(2,88) = 3.62$; $P < 0.05$; and $F_{\text{interaction}}(6,88) = 0.23$, $P > 0.05$, two-way ANOVA, Figure 8B, page 89). Ondansetron 0.001 mg/kg shows a slight reduction in the duration of axial AIMs whereas, the amplitude axial AIMs was increased with respect to the vehicle-treated animals.

Limbs AIMs

As shown in Figure 8C (page 89), administration of ondansetron in combination with L-DOPA did not lead to significant reduction in the duration of limbs AIMs ($F_{\text{time}}(3,88) = 2.496$; $F_{\text{treatment}}(2,88) = 2.87$; and $F_{\text{interaction}}(6,88) = 0.6427$, each $P > 0.05$, two-way ANOVA). Similarly, ondansetron did not significantly diminish the amplitude of limbs AIMs ($F_{\text{time}}(3,88) = 1.455$; $F_{\text{treatment}}(2,88) = 1.181$; and $F_{\text{interaction}}(6,88) = 0.2405$, each $P > 0.05$, two-way ANOVA, Figure 8D, page 89).

Orolingual AIMs

As illustrated in Figure 8E (page 89), adding ondansetron to L-DOPA did not significantly affect the duration of orolingual AIMs ($F_{\text{time}}(3,88) = 1.052$; $F_{\text{treatment}}(2,88) = 0.9368$; and $F_{\text{interaction}}(6,88) = 0.5472$, each $P > 0.05$; two-way ANOVA). In contrast, administration of ondansetron resulted in a significant decrease in the amplitude of orolingual AIMs ($F_{\text{time}}(3,88) = 0.8776$, $P > 0.05$; $F_{\text{treatment}}(2,88) = 38.12$; $P < 0.0001$; and $F_{\text{interaction}}(6,88) = 1.105$, $P > 0.05$, two-way ANOVA, Figure 8F, page 89). Thus, ondansetron 0.0001 mg/kg

significantly reduced the amplitude of orolingual AIMs on days 1, 8, 5 and 22 by 21%, 32%, 37% and 24%, respectively, when compared with vehicle ($P < 0.05$, $P < 0.001$, $P < 0.0001$ and $P < 0.05$, Tukey's *post hoc* test). The dose of 0.001 mg/kg ondansetron showed a similar trend in the development of orolingual AIMs as the vehicle group.

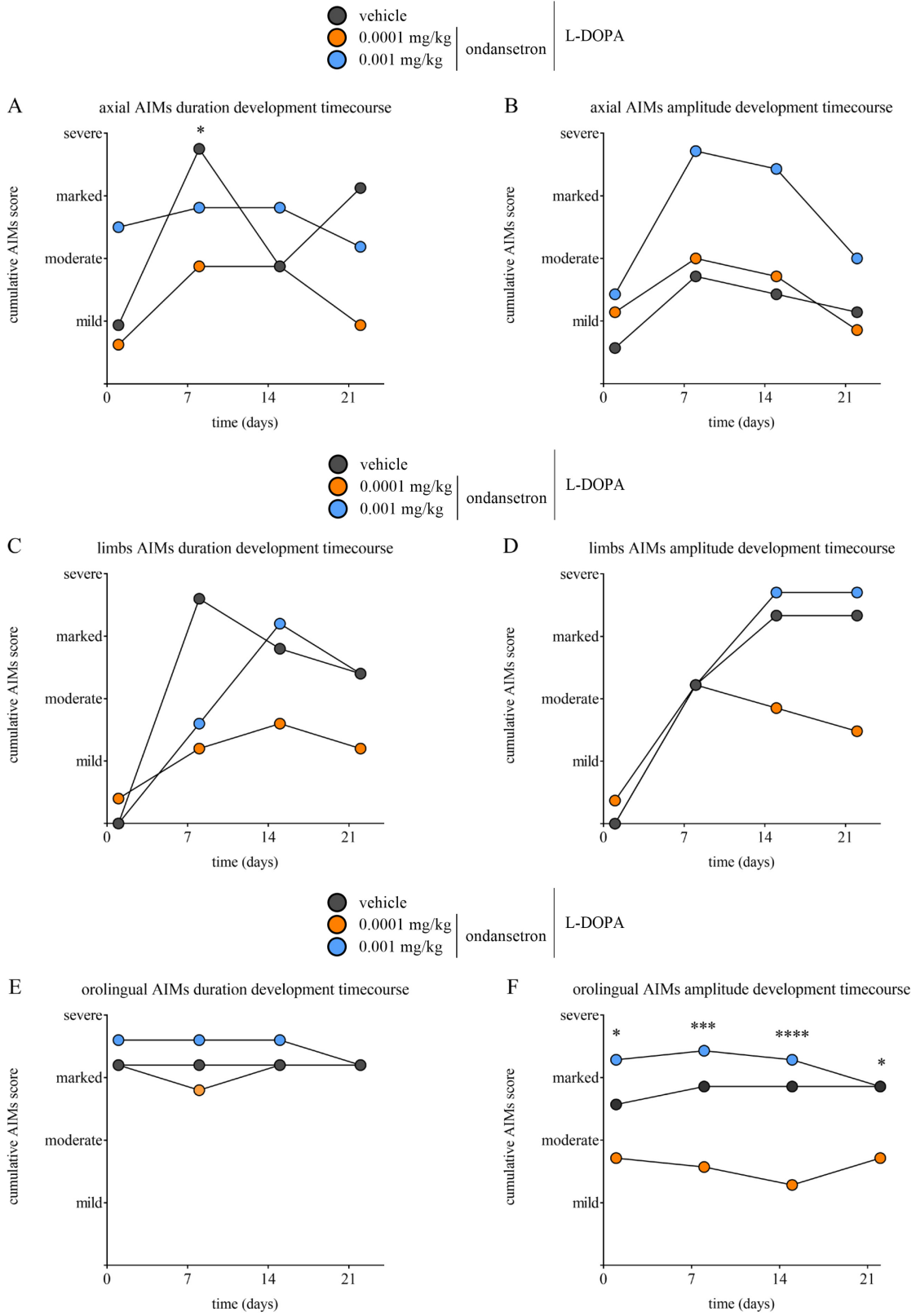
AL AIMs

As shown in Figure 8G (page 90), the addition of ondansetron significantly reduced the duration of AL AIMs ($F_{\text{time}}(3,88) = 3.064$; $P < 0.05$; $F_{\text{treatment}}(2,88) = 7.953$; $P < 0.001$; and $F_{\text{interaction}}(6,88) = 0.4133$, $P > 0.05$, two-way ANOVA). Thus, on day 8, animals that received ondansetron 0.0001 mg/kg with L-DOPA exhibited a 53% shorter duration of AL AIMs, when compared with those that received vehicle/L-DOPA ($P < 0.05$, Tukey's *post hoc* test). In contrast, ondansetron did not lead to a significant reduction in the amplitude of AL AIMs ($F_{\text{time}}(3,88) = 1.623$; $F_{\text{treatment}}(2,88) = 2.959$; and $F_{\text{interaction}}(6,88) = 0.1974$, each $P > 0.05$, two-way ANOVA) but it appears that animals administered 0.0001 mg/kg ondansetron exhibited a slight decrease in the amplitude AL AIMs, when compared with the control group (Figure 8H, page 90). Treatment with 0.001 mg/kg ondansetron produced a modest decrease in the duration of AL AIMs with respect to vehicle-treated animals while the severity of amplitude was unaffected.

ALO AIMs

As illustrated in Figure 8I (page 90), adding ondansetron resulted in a significant reduction in the duration of ALO AIMs ($F_{\text{time}}(3,88) = 2.969$; $P < 0.05$; $F_{\text{treatment}}(2,88) = 8.797$; $P < 0.001$; and $F_{\text{interaction}}(6,88) = 0.6006$; $P > 0.05$, two-way ANOVA). Thus, on day 15, administration of ondansetron 0.0001 mg/kg in combination with L-DOPA led to a 33% decrease in the duration of ALO AIMs, when compared with vehicle ($P < 0.01$, Tukey's *post hoc* test). Similarly, ondansetron treatment also diminished the amplitude of ALO AIMs ($F_{\text{time}}(3,88) = 0.3526$; $P > 0.05$; $F_{\text{treatment}}(2,88) = 13.87$; $P < 0.0001$; and $F_{\text{interaction}}(6,88) = 0.9623$; $P > 0.05$, two-way ANOVA). On day 8, 0.0001 mg/kg ondansetron led to a 31% reduction in the amplitude of ALO AIMs, when compared with vehicle ($P < 0.05$, Tukey's *post hoc* test, Figure 8J, page 90). On day 15, ondansetron 0.001 mg/kg showed an 49% increase in the amplitude of ALO AIMs, when compared with the 0.0001 mg/kg dose ($P < 0.01$, Tukey's *post hoc* test).

Timecourse graphs that present the data as median with interquartile range are shown in Figure 12 (Appendices VI and VII).



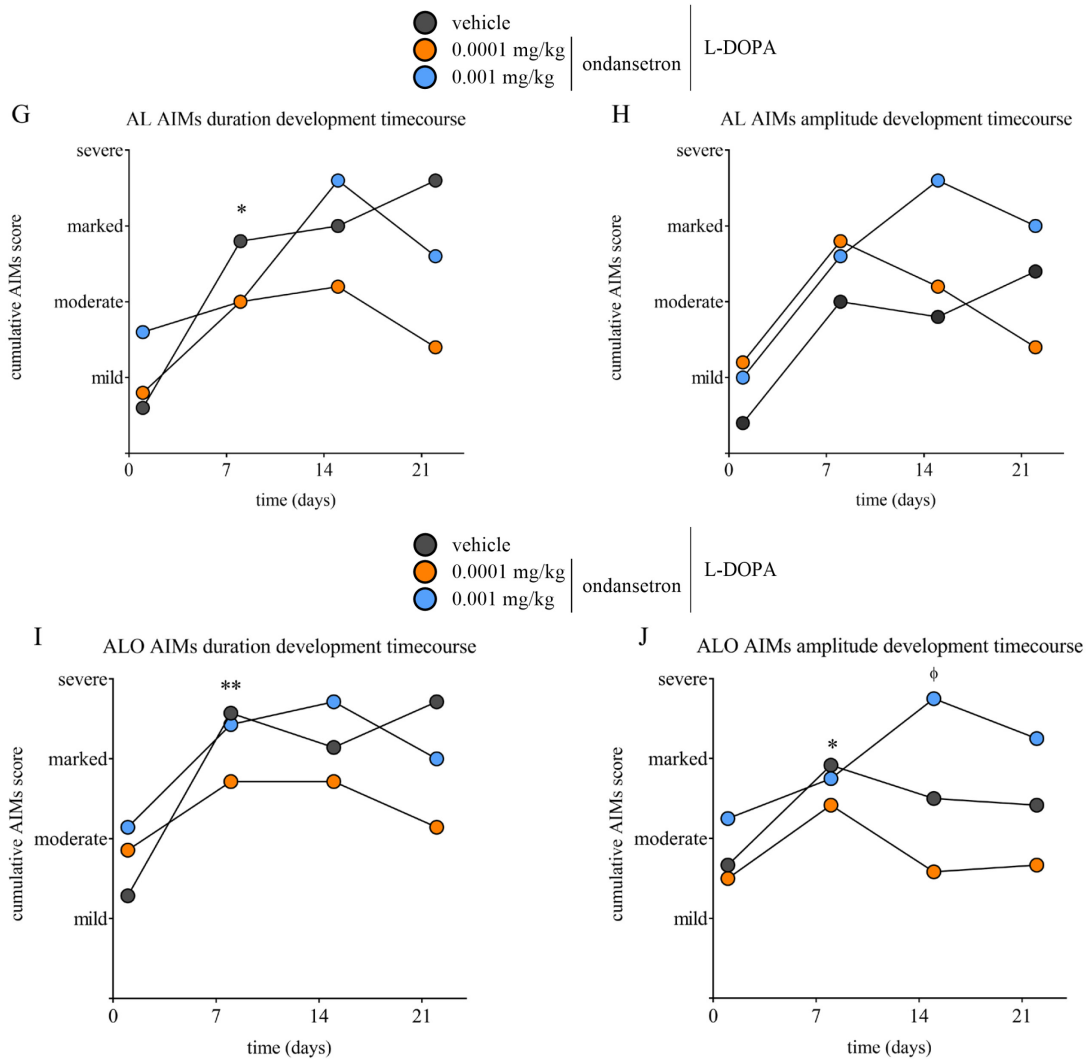


Figure 8: Time course of the development of AIMs during the 22-day priming phase of the *de novo* ondansetron study. On day 8 of priming, **A.** the duration of axial AIMs was significantly reduced in animals previously treated with L-DOPA/0.0001 mg/kg ondansetron ($n = 9$), by 53%, when compared with L-DOPA/vehicle ($n = 7$) ($P < 0.05$, Tukey's *post hoc* test). **B.** In contrast, the amplitude of axial AIMs was not significantly reduced in animals that were previously treated with L-DOPA/ondansetron 0.0001 mg/kg ($n = 9$) compared to animals that received L-DOPA/vehicle. **C.** The duration and **D.** amplitude of limbs AIMs was comparable between animals administered ondansetron 0.0001 mg/kg and vehicle. **E.** The duration of orolingual AIMs was not affected by ondansetron treatment. **F.** Administration of L-DOPA/ondansetron 0.0001 mg/kg resulted in a significant decrease in the amplitude of orolingual AIMs on days 1, 8, 5 and 22 by 21%, 32%, 37% and 24%, respectively, when compared with vehicle ($P < 0.05$, $P < 0.001$, $P < 0.0001$ and $P < 0.05$, Tukey's *post hoc* test). **G.** On day 8, when administered with L-DOPA, 0.0001 mg/kg ondansetron significantly reduced the duration of AL AIMs, by 53%, when compared with L-DOPA/vehicle ($P < 0.05$,

Tukey's *post hoc* test). **H.** The amplitude of AL AIMS was comparable between L-DOPA/0.0001 mg/kg ondansetron and L-DOPA/vehicle. **I.** Administration of ondansetron 0.0001 mg/kg resulted in a significant decrease in the duration of ALO AIMS on day 15, by 33%, when compared with L-DOPA/vehicle ($P < 0.01$, Tukey's *post hoc* test). **J.** The amplitude of ALO AIMS was reduced in animals that were treated with L-DOPA/ondansetron 0.0001 mg/kg, by 31%, compared to animals that received L-DOPA/vehicle ($P < 0.05$, Tukey's *post hoc* test). Data are graphed as the median. *: $P < 0.05$; **: $P < 0.01$, ***: $P < 0.001$, ****: $P < 0.0001$ vehicle versus ondansetron 0.0001 mg/kg; ϕ : $P < 0.01$ ondansetron 0.0001 mg/kg versus ondansetron 0.001 mg/kg.

***De novo* treatment with ondansetron attenuates the development of ALO AIMs**

To determine if any reduction of AIMs observed during the priming phase was due to a symptomatic effect by ondansetron or resulted from an interference with AIMs development, following the 22-day priming phase, rats entered a 3-day washout period, after which they were administered an acute L-DOPA challenge (Figure 9, page 93).

As shown in Figures 9A and 9B (page 93), rats previously administered L-DOPA/ondansetron demonstrated a reduction in the duration of cumulative and peak ALO AIMs ($F(2,22) = 4.814$ and $F(2,22) = 4.389$, both $P < 0.05$, one-way ANOVA). Treatment with L-DOPA/ondansetron 0.0001 mg/kg and 0.001 mg/kg did not statistically diminish the duration of cumulative and peak ALO AIMs when compared with L-DOPA/vehicle (both $P > 0.05$, Tukey's *post hoc* test). In contrast, the dose of 0.0001 mg/kg ondansetron led to a 32% and 47% decrease in the duration of cumulative and peak ALO AIMs, respectively, when compared with the dose of 0.001 mg/kg (both $P < 0.05$, Tukey's *post hoc* test).

As illustrated in Figure 9C (page 93), the addition of ondansetron led to a significant reduction in the amplitude of cumulative ALO AIMs ($F(2,22) = 5.996$, $P < 0.01$, one-way ANOVA). Thus, administration of L-DOPA/ondansetron 0.0001 mg/kg resulted in a decrease in the amplitude of cumulative ALO AIMs by 33% and 34%, respectively, when compared with L-DOPA/vehicle and L-DOPA/ondansetron 0.001 mg/kg, respectively (both $P < 0.05$, Tukey's *post hoc* test). As shown in Figure 9D (page 93), ondansetron diminished the amplitude of peak ALO AIMs ($F(2,22) = 3.513$, $P < 0.05$, one-way ANOVA). Thus, ondansetron 0.0001 mg/kg led to a significant reduction in the amplitude of peak ALO AIMs by 47%, when compared with the dose of 0.001 mg/kg ($P < 0.05$, Tukey's *post hoc* test).

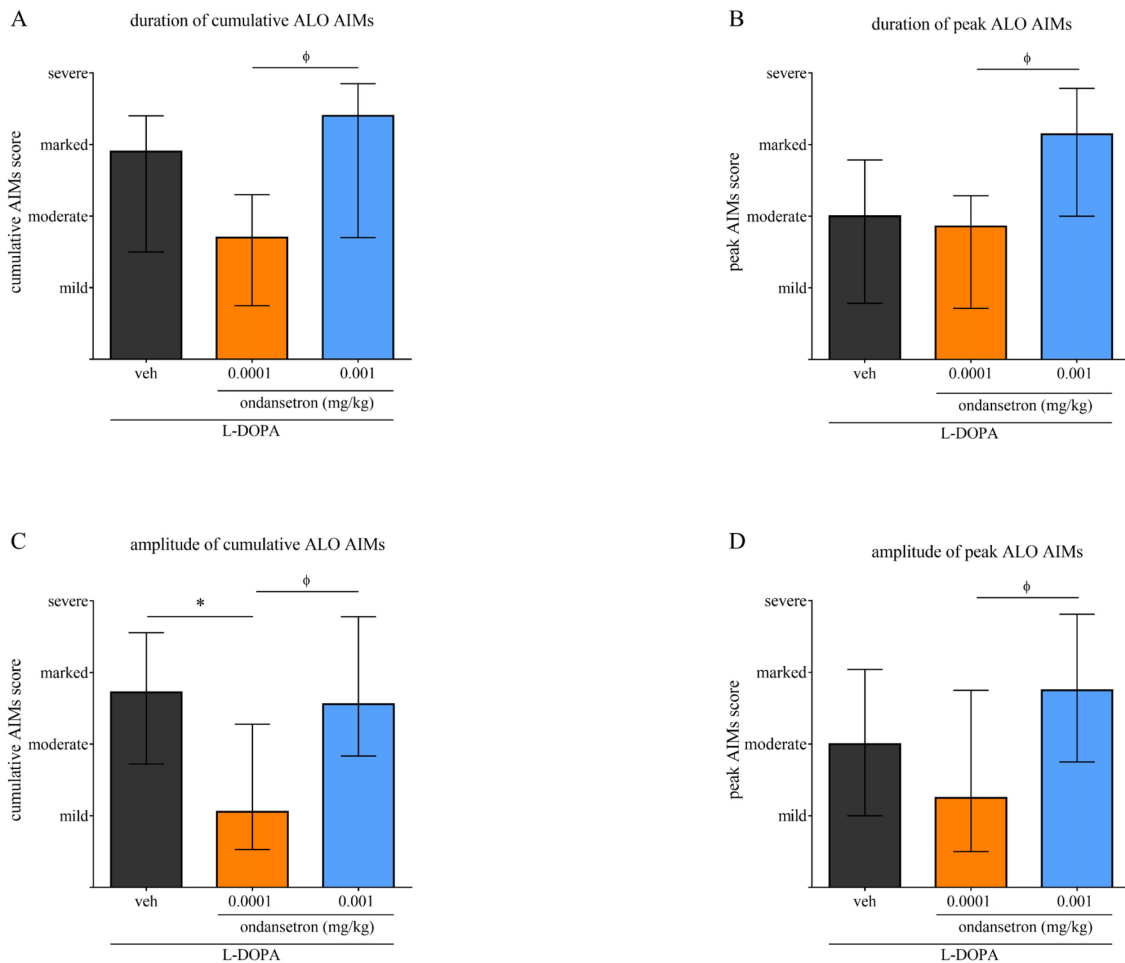


Figure 9: Effect of ondansetron on the duration and amplitude of cumulative and peak AIMs severity during an acute 6 mg/kg L-DOPA challenge. During an acute L-DOPA challenge, rats that were previously administered L-DOPA/ondansetron 0.0001 mg/kg ($n = 9$) during the priming period exhibited similar **A.** cumulative and **B.** peak duration ALO AIMs, when compared with L-DOPA/vehicle ($n = 6$). In combination with L-DOPA, 0.0001 mg/kg ondansetron led to a 32% and 47% decrease in the duration of cumulative and peak ALO AIMs, respectively, when compared with the dose of 0.001 mg/kg ($n = 9$), (both $P < 0.05$, Tukey's *post hoc* test). **C.** In contrast, previous addition of ondansetron 0.0001 mg/kg ondansetron with L-DOPA resulted in a significant reduction in the amplitude of cumulative ALO AIMs, by 33% and 34%, when compared with L-DOPA/vehicle and L-DOPA/0.001 mg/kg ondansetron, respectively (both $P < 0.05$, Tukey's *post hoc* test). **D.** Ondansetron 0.0001 mg/kg led to a significant decrease in the amplitude of peak ALO AIMs by 47%, when compared with the dose of 0.001 mg/kg ($P < 0.05$, Tukey's *post hoc* test). Cumulative and peak AIM scores are expressed as median with interquartile interval. *: $P < 0.05$ vehicle versus ondansetron 0.0001 mg/kg; ϕ : $P < 0.05$ ondansetron 0.0001 mg/kg versus ondansetron 0.001 mg/kg.

Administration of ondansetron does not impair the therapeutic efficacy of L-DOPA in the cylinder test

Following the acute challenges of ondansetron, 6-OHDA-lesioned rats were then subject to a 3 mg/kg L-DOPA challenge to assess whether ondansetron treatment impairs L-DOPA anti-parkinsonian action, as measured by the cylinder test (Figure 10, page 96). As illustrated in Figure 10A (page 96), treatment conditions significantly improved use of the left (lesioned) forepaw in making wall contacts (FS = 28.54, $P < 0.0001$). Although administration of L-DOPA did not alter performance ($P > 0.05$, Dunn's *post hoc* test), ondansetron 0.0001 mg/kg and 1 mg/kg resulted in a significant increase the number of rears using the impaired forepaw when compared to post-surgery performance (both $P < 0.001$, Dunn's *post hoc* test).

As shown in Figure 10B (page 96), administration of L-DOPA alone or in combination with ondansetron led to a decrease in the use of the right (un-lesioned) forepaw (FS = 34.89, $P < 0.0001$). When 6-OHDA-lesioned rats were administered L-DOPA, there was a significant decrease in the number of rears using the un-lesioned side by 34% ($P < 0.01$, Dunn's *post hoc* test). This decrease in rears with the un-lesioned forepaw remained present when ondansetron 0.0001, 0.001, 0.01, 0.1 or 1 mg/kg was combined with L-DOPA by 49%, 38%, 41%, 61% and 41%, respectively (each, $P < 0.001$, $P < 0.01$, $P < 0.001$, $P < 0.0001$ and $P < 0.0001$, Dunn's *post hoc* test). There was no difference between the number of rears using the un-lesioned side between L-DOPA/vehicle and L-DOPA/ondansetron, regardless of the dose of ondansetron (each $P > 0.05$, Dunn's *post hoc* test).

As illustrated in Figure 10C (page 96), 6-OHDA-lesioned animals that received L-DOPA or L-DOPA/ondansetron, demonstrated a significant increase in rears using both forepaws (FS = 21.52, $P < 0.0001$). Following administration of L-DOPA, there was a significant increase in rears using both forepaws, by 156%, compared to drug-naïve animals ($P < 0.05$, Dunn's *post hoc* test). In combination with L-DOPA, ondansetron 0.0001, 0.01 and 0.1 mg/kg increased the use of both forepaws by 114%, 114% and 163%, respectively ($P < 0.05$, $P < 0.05$ and $P < 0.001$, respectively, Dunn's *post hoc* test). There was no difference between the number of rears using both forepaws to rear between L-DOPA/vehicle and L-DOPA/ondansetron, regardless of the dose of ondansetron (each $P > 0.05$, Dunn's *post hoc* test).

Results of the effect size calculated by Glass' delta, which accounts for the variation in standard deviations across treatment conditions in Figures 10B and 10C, are presented in Tables V and VI (Appendices III and IV).

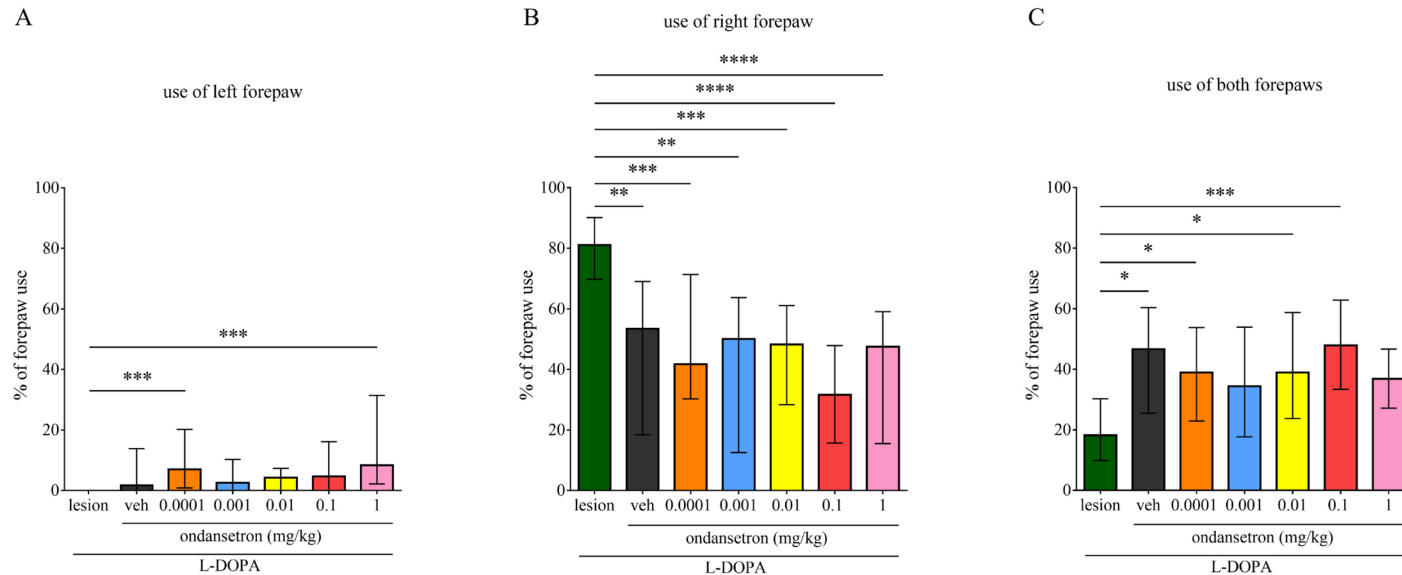


Figure 10: Use of forepaws across treatment conditions. **A.** Left forepaw use across treatments. Drug-naïve 6-OHDA-lesioned rats ($n = 18$) did not use the left (lesioned) forepaw during rearing. Following administration of L-DOPA (3/15 mg/kg), there was no significant change in the number of rears with the lesioned forepaw. The addition of ondansetron 0.0001 and 1 mg/kg to L-DOPA significantly improved use of the lesioned forepaw, when compared with post-surgery performance. **B.** Right forepaw use across treatments. 6-OHDA-lesioned rats ($n = 18$) used the right (un-lesioned) forepaw in 83% of rears. When 6-OHDA-lesioned rats were administered L-DOPA (3/15 mg/kg), there was a significant decrease in the number of rears using the un-lesioned side by 40%. This decrease in rears with the un-lesioned forepaw remained present when ondansetron 0.0001, 0.001, 0.01, 0.1 or 1 mg/kg was combined with L-DOPA by 48%, 39%, 46%, 57% and 51%, respectively. **C.** Use of both forepaws across treatments. 6-OHDA-lesioned rats ($n = 18$) used both forepaws during 17% of rears. Administration of L-DOPA (3/15 mg/kg) led to a significant increase in rears using both forepaws by 118%. In combination with L-DOPA, ondansetron 0.0001, 0.01 and 0.1 mg/kg increased the use of both forepaws by 113%, 121% and 159%, respectively. Data are graphed as median with interquartile range. *: $P < 0.05$; **: $P < 0.01$, ***: $P < 0.001$, ****: $P < 0.0001$.

IV. Discussion

These results demonstrate that ondansetron significantly diminishes the severity of established L-DOPA-induced AIMs compared to L-DOPA alone, without compromising the anti-parkinsonian action of L-DOPA. Furthermore, administration of ondansetron, when begun concurrently with L-DOPA, attenuates the development of AIMs compared to L-DOPA alone. Taken together, these results suggest that 5-HT₃ receptor antagonism represents a novel and effective therapeutic strategy to reduce LID in PD. The following discussion will focus on how 5-HT₃ receptor blockade may be an effective approach to reduce the severity of LID.

Limitations and future directions

In light of the findings reported in the present thesis, it is also important to highlight the limitations of the experiments. First, the pharmacokinetic study characterized peak ondansetron levels, previously unknown, in the rat with the doses used. However, determining brain levels of ondansetron in human is hardly feasible for ethical reasons, so it is not possible to compare or correlate plasma levels of ondansetron with brain levels. Further studies are thus needed to determine the plasma and brain concentration profile of ondansetron and brain ondansetron levels in the brain associated with maximum anti-dyskinetic activity. Second, the primary endpoint of this Master's thesis was to assess the effect of ondansetron on the severity of dyskinesia in a PD animal model. This preclinical study modelled the administration of ondansetron in the clinic as an adjunct to L-DOPA and demonstrated that it achieved its effect on dyskinesia without impairing the therapeutic efficacy of L-DOPA. It would be interesting to study the effect of ondansetron on basal PD disability in a subsequent set of experiments. Third, in the 6-OHDA-lesioned rat, an increase in SERT levels is reported in the striatum of dyskinetic animals and, furthermore, levels of SERT correlate with AIMs severity (989, 990). In addition, following 2-3 weeks of either low or high doses of L-DOPA, SERT levels also increase in the rat striatum (990). Interestingly, the increase in striatal SERT levels appears to be a dose-dependent effect of L-DOPA-induced axonal sprouting (990). Similarly, in the MPTP-lesioned macaque model, there is an increase in the number of 5-HT axon varicosities in the striatum, which is particularly pronounced where DA denervation is most severe (991). Although these studies may be in accordance with the agreed upon role of serotonergic terminals in the pathophysiology of dyskinesia (736), further anatomical characterization in the 6-OHDA-lesioned rat model is needed before inferring their action on raphe-striatal pathways. Fourth, the

acute challenges of ondansetron study demonstrated the acute suppression of L-DOPA induced AIMs in the 6-OHDA lesioned rat model. Whether this effect would be maintained over the long-term remains unknown, and a chronic ondansetron study, where ondansetron would be administered over several days, with regular assessments of AIMs severity, could determine whether the anti-dyskinetic efficacy of ondansetron is maintained or if tolerance develops. Fifth, most of the thesis was a behavioural pharmacology study and is not informative on the mechanism whereby 5-HT₃ receptor blockade reduces the severity of established and prevents the development of dyskinesia. Thus, we are actively seeking answers on elucidating the mechanism that underlies the action of 5-HT₃ blockade, notably by conducting studies that will shed light on the brain areas involved in mediating the anti-dyskinetic effect of antagonising 5-HT₃ receptors. Sixth, several highly-selective 5-HT₃ receptor antagonists are clinically available with differing affinity for the receptor and duration of action, and it would be of further interest to determine whether the pharmacodynamics or pharmacokinetic profiles of a drug influence its anti-dyskinetic efficacy.

Pharmacokinetic study of clinically relevant doses of ondansetron

The preliminary dose-finding PK study, which aimed at assessing whether there was a ceiling effect to anti-dyskinetic efficacy of higher doses of ondansetron, found that ondansetron 0.01 mg/kg led to a C_{max} of 2.31 ng/mL. Clinical studies have described oral administration of ondansetron in healthy volunteers and a single oral dose of 8 mg measured a C_{max} of 19.9 – 31.2 ng/mL (992, 993). The dose of 0.0001 mg/kg ondansetron that conferred the therapeutic benefit in the behavioural studies is likely well below plasma levels in humans and thus, well tolerated in humans. On the other hand, the higher doses of 0.1 and 1 mg/kg ondansetron likely led to 10-fold to 100-fold higher than well tolerated plasma levels in humans, which limits the translational potential of administering higher doses of ondansetron in the clinic, especially if the dosing regimen requires exceeding maximum tolerated doses of ondansetron. As the PK of subcutaneous ondansetron administration has not been assessed in humans, it is not possible to make a valid comparison to assess whether the dose of 0.01 mg/kg ondansetron is clinically relevant (993). It is thus warranted to conduct PK studies of subcutaneous oral administration in primates to confirm anti-dyskinetic doses of ondansetron are clinically relevant.

5-HT₃ blockade with ondansetron alleviates previously established AIMs without impairing the anti-parkinsonian efficacy of L- DOPA

In the present study, during the course of treatment, ondansetron at 0.0001 mg/kg consistently produced the lowest AIMs severity whereas the vehicle produced the most severe AIMs treatment, except for the orolingual AIMs. Accordingly, the duration and amplitude ranking time courses consistently show that the cumulative and peak AIMs with ondansetron 0.0001 mg/kg are significantly reduced, when compared to vehicle. Importantly, ondansetron reduced the severity of cumulative AIMs as well as the peak severity of AIMs, which coincides with the peak L-DOPA concentration, suggesting that ondansetron may alleviate dyskinesia throughout the time period they are expressed, regardless of their intensity, which has important therapeutic implications.

The dose of 0.0001 mg/kg ondansetron was the most effective at alleviating established AIMs, particularly the axial and limbs components, compared to the more cumulative measures of AL and ALO AIMs. Orolingual AIMs are often more difficult to score compared to the other dyskinetic parameters and, as they are more subtle, they may be overestimated or overlooked (965). In the investigators' experience, milder severity levels present with subtle differences while at higher severity, more prominent AIMs of the neck and upper body as well as the forelimb can mask the appearance of orolingual AIMs. Indeed, treatment with ondansetron did not produce a significant reduction on the duration or the amplitude of orolingual AIMs. Thus, while ALO AIMs evaluate the anti-dyskinetic effect of a treatment on the sum of the three components, it is also of interest to express the data as AL AIMs, which are arguably relatively more disabling than orolingual AIMs. Interestingly, despite the lack of effect on the severity and temporal profile of orolingual AIMs, ondansetron appeared to have a more pronounced effect on the severity of ALO than AL AIMs, suggesting that the efficacy of the anti-dyskinetic effect is maintained on cumulative dyskinetic parameters.

A criticism that could be raised on the scale used here is that while it evaluates the severity of ALO AIMs across both duration and amplitude, it does not differentiate between dystonic and hyperkinetic dyskinesia, which could be correlates of dystonia and chorea,

respectively. Steece-Collier and collaborators have introduced a modification to the scale that scores dystonic and hyperkinetic axial and limbs AIMs separately (994), and might allow the detection of differential effects of treatments on these subtypes of dyskinesia (965), and could possibly have enabled us to make predictions as to whether 5-HT₃ blockade might be more effective, in clinical settings, to reduce dystonia, chorea, or both.

The bell-shaped dose-response curve is often ascribed to 5-HT₃ receptor antagonists for indications in preclinical and clinical studies including anxiety (995), depression (996, 997), drug addiction (998) and migraines (883). In general, the maximum response is already observed in the microgram dose range, while higher doses are ineffective (848). However, the mechanism underlying the dose-response curve still lacks satisfactory explanation and it remains to be determined whether this is due to the pharmacology of individual compounds or a group-specific characteristic (818). Thus, in our experiments, lower doses of ondansetron produced the greatest relief of dyskinesia. The most favoured mechanism proposes that, at high concentrations of 5-HT₃ antagonists, there is mutual steric hindrance at the receptor, which refers to an inappropriate interaction of a ligand to its receptor that results in a conformational change in the receptor that prevents the binding of ligands to the receptor” (999-1001) or, more speculatively, additional effects due to low-affinity binding to other receptors. Although ondansetron binds with low affinity to the 5-HT_{1A}, 5-HT_{1B}, α -adrenergic and opioid receptors, its binding to high-affinity 5-HT₃ receptor sites is about 250- and 500-fold higher than that of the other receptors (786, 870, 1002). Alternatively, as the density of 5-HT₃ receptors varies between different brain regions with one density type being completely inhibited at low concentrations and the other type only at high concentrations of 5-HT receptor antagonists, which could explain contrary effects (818), *e.g.* a therapeutic effect, triggered by blockade of 5-HT₃ receptors within one brain area, could be offset when 5-HT₃ receptors from another brain region are completely antagonised (882), which could explain the lack of efficacy of higher doses of ondansetron.

According to the classic model of BG circuitry, dyskinesia may arise as the result of overactivity of the direct pathway and/or underactivity of the indirect pathway (299, 572, 580). Some autoradiographic studies report the expression of 5-HT₃ receptors on GABAergic MSNs of the direct and indirect pathways (638). In view of the anti-dyskinetic results described above, it is possible that, at lower doses of ondansetron, the compound antagonizes inhibitory GABAergic projection neurons of the striatum, and may preferentially block pre-synaptic 5-

HT₃ receptors on MSNs of the direct pathway over MSNs of the indirect pathway. Here, ondansetron would act on the pre-synaptic nerve terminal to prevent the entry of cations into the neurons and their subsequent depolarization followed by release of neurotransmitters (849, 850). As a result, the propagation of inhibitory GABAergic signal to the EP/SNr is reduced, which would diminish hyperactivity of the direct pathway in LID, and may lead to the dampening of AIMs observed with lower doses of the compound.

5-HT₃ receptors are also found on striatal GABAergic interneurons, notably the stomatostatin-/nitric oxide synthase- and calretinin-expressing interneuron subtypes. These two major interneuron populations exhibit high input resistance and persistent low-threshold spiking (PLTS) in response to intracellular depolarization or excitatory synaptic stimulation (1003). Although the output of individual PLTS interneurons is relatively weak and sparse, they may form inhibitory synapses onto distal dendrites of MSNs (1004), which is consistent with SOM+ GABAergic terminals that have been observed on the dendrites of MSNs (1005). Moreover, it is possible that the low connectivity of PLTS interneurons is strengthened under disease states (1006), where they release neuromodulators such as neuropeptide Y, SOM and NO, which more speculatively, may modulate the activity of striatal GABAergic MSNs of the direct pathway (1007-1010).

In addition to their action on MSNs from the direct pathway, antagonism of 5-HT₃ receptors on striatal GABAergic interneurons may exert an inhibitory effect on the activity of the direct pathway that partially restores inhibition of motor cortical areas to physiological levels. Although the presence of AADC expression in striatal interneurons may be controversial (1011, 1012), they could represent another source of L-DOPA derived DA (1013, 1014). Here, ondansetron may prevent extra synaptic DA release by blocking the 5-HT₃ receptor on striatal interneurons, thereby diminishing fluctuations in DA levels associated with the appearance of LIDs (745). In contrast, at higher doses of ondansetron, the compound may also antagonize 5-HT₃ receptors on MSNs of the indirect pathway. This could theoretically result in further disturbance to the equilibrium between the direct and indirect pathways and lead to a greater disinhibition of the motor thalamus and motor cortex, exacerbating existing L-DOPA-induced AIMs.

It should be noted though, that such mechanisms have yet to be demonstrated experimentally, which is further emphasised by the fact that there is currently no consensus in

the literature on the distribution of 5-HT₃ receptors in the rodent BG. In the rodent striatum, studies have reported a variety of expression patterns ranging from weak to moderate expression of the 5-HT₃ receptors. However, a recent immunostaining study in the mouse brain has attempted to address the limitations of these binding studies by using 5-HT_{3A} receptor-green fluorescent protein transgenic mice to map the 5-HT_{3A} receptor subunit in the mouse brain (897). The authors reported moderately high expression of the 5-HT_{3A} receptor subunit in the striatum, slight expression in the thalamus but no expression in the GP and SN. Thus, based upon our results, assuming that the striatum is the structure that mediates, at least in part, the anti-dyskinetic effect of 5-HT₃ blockade, these weak-moderate expression levels of the 5-HT_{3A} receptor are probably sufficient to mediate a behavioural effect. Another possibility is that 5-HT₃ expression is altered in the dyskinetic state, but studies on the expression of 5-HT₃ in the dyskinetic state have yet to be performed.

Nevertheless, 5-HT₃ receptors appear to be well situated to modulate the release of neurotransmitters within the BG, which may be responsible for their anti-dyskinetic effect. Thus, anatomical, electrophysiological and behavioural studies have suggested an important functional crosstalk between 5-HT and DA pathways in the BG. In particular, it has been demonstrated that the 5-HT₃ receptor mediates changes in striatal dopamine release *in vitro* (1015) and *in vivo* (911, 912, 1016). In fact, a microdialysis study reported that intrastriatal injection of the 5-HT₃ antagonists 3-tropanyl-indole-3-carboxylate, MDL-72222 or ondansetron attenuated 5-HT or morphine-induced striatal DA release, which suggests that 5-HT acts at 5-HT₃ receptors to facilitate DA release in the striatum (1017, 1018). Consistent with these results, systemic administration of the 5-HT₃ antagonists ondansetron and MDL 72222 did not affect basal DA efflux in the striatum (1018, 1019). Taken together, these results suggest that the 5-HT₃ receptor regulates evoked nigrostriatal DA release (1020), which is dependent on depolarization and the concomitant elevation of both DA and 5-HT tones (1021). Furthermore, in rat striatal slices, application of the selective 5-HT₃ agonists 2-methyl-5-HT and phenylbiguanide increased endogenous release of DA (1022), and DA release induced by 5-HT and the 5-HT₃ agonist 2-methyl-5-HT was attenuated by the 5-HT₃ antagonist ICS 205-930 (1023). In addition, in the dyskinetic state, where 5-HT fibres mediate DA release; administration of ondansetron may reduce the excitatory 5-HT innervation to the striatum, and this would lead to a reduction of DA release, which would translate behaviourally by a reduction

of dyskinesia. Lastly, it has been suggested that 5-HT₃ receptor may reduce the number of electrically-active and firing neurons within the SN and VTA and produce reductions in evoked DA release within the striatum and nucleus accumbens (1024); here again, this reduction of DA release might translate, at the behavioural level, by a reduction of dyskinesia. Furthermore, behavioural experiments have also suggested that pharmacological modulation of the 5-HT₃ receptor modifies nigrostriatal DA-induced motor responses, for instance, intrastriatal injection of the 5-HT₃ agonists SR-57227A or 2-methyl-5-HT in mice led to contralateral rotations that were abolished with systemic administration of the 5-HT₃ antagonist ondansetron (863). Taken together, these studies implicate the involvement of the 5-HT₃ receptor in DA release in the striatum, and it can be inferred that administration of ondansetron may have attenuated this excessive release of DA and led to the reduction in the severity of ALO AIMs.

L-DOPA derived DA release also occurs outside the striatum and other brain regions that receive 5-HT innervation display increased extrasynaptic DA release that may contribute to the development of LID (1025). Volume transmission of DA and its overflow past its release site can lead to the interaction of DA with multiple synapses (1026), and this volume increases even further with the degeneration of DA axons and loss of DAT activity in PD (1027). Given that extrasynaptic DA receptors are activated further from release sites (1027), pharmacological modulation of DA release in the striatum may exert effects on other BG nuclei including the STN, the entopeduncular nucleus (EP), the rodent homologue to the primate GPi, the SNr as well as the thalamus. The STN is considered the major driving force in the BG circuitry (554, 1028); thus, alterations in its activity by the 5-HT₃ receptor could represent an important site of action to reduce dyskinesia. Intrasubthalamic injection of the 5-HT₃ agonist mCPBG in rats induced contralateral turning behaviour, which was suppressed by lesion of the SNr (1029), which suggests that contralateral rotations could result from decreased excitatory input from the STN to the SNr, which in turn enhanced the activity of the ipsilateral motor thalamus (1029). In spite of the lack of autoradiographic studies on the distribution of the 5-HT₃ receptor in the STN, these findings suggest the presence of functional 5-HT₃ receptors in the STN. In line with the localization of 5-HT₃ receptors to GABAergic neurons, including in the striatum (814, 897) and neocortex (804, 822, 823), blockade of 5-HT₃ receptor may exert an inhibitory effect on subthalamic neurons. The consequent increase in glutamatergic output to the EP/SNr causes the inhibition of the motor thalamus and motor cortex, which might alleviate the severity of

dyskinesia. However, additional autoradiographic and immunohistochemical studies need to be conducted to confirm the expression of 5-HT₃ receptors in the STN.

Effect of ondansetron and ALO AIMs on L-DOPA anti-parkinsonian action

In agreement with previous studies (885, 973, 1030), L-DOPA improved 6-OHDA lesion-induced forelimb use asymmetry at the cylinder test. Importantly, this effect of L-DOPA was maintained after ondansetron was administered in our experiments, which indicates that ondansetron did not impair L-DOPA anti-parkinsonian effect.

A limitation of the cylinder test is that it can be impaired by AIMs. To circumvent this limitation, we used L-DOPA 3 mg/kg when performing the test, a dose that should theoretically not trigger AIMs. However, studies report that even such small doses of L-DOPA can trigger AIMs after chronic dopaminergic therapy (1031, 1032), and an interference of AIMs in the scoring occurred in our study with higher doses of ondansetron, with which AIMs were not reduced, which rendered difficult accurate rearing assessment. Retrospectively, a lower dose of L-DOPA, might have been sufficient to assess the anti-dyskinetic action of L-DOPA without eliciting AIMs in the animals. Alternatively, other tests of physiological motor behaviour such as the rotarod test (1033), open field test (1034) or stepping test (1035) should be administered in conjunction with the cylinder test as they may show greater sensitivity to detect motor activity, although severe AIMs could theoretically alter the performance at these tests as well.

Effect of ondansetron on the development of ALO AIMs

In the present study, chronic L-DOPA administration for 22 days in 6-OHDA-lesioned rats induced the expression of AIMs and the severity increased over time before reaching a plateau. In contrast, in animals treated with both ondansetron 0.0001 mg/kg and L-DOPA the development of AIMs was significantly attenuated. Animals treated with ondansetron 0.001 mg/kg and L-DOPA showed a similar progression in the development of AIMS as the control-DOPA/vehicle group, as opposed to the modest anti-dyskinetic effect observed in the acute challenge study. Following the 22-day priming phase, animals were subject to an acute L-DOPA challenge to assess whether ondansetron treatment interfered with the priming process that led

to the development of dyskinesia, or if the apparent dyskinesia reduction during the priming phase was due to a symptomatic effect of the compound. If ondansetron indeed attenuates the development of dyskinesia, we would expect the L-DOPA/vehicle group of animals to display more severe AIMs when compared to the L-DOPA/ondansetron groups upon administration of L-DOPA alone after the washout period that followed the priming phase. We found that, upon acute administration of L-DOPA alone after the priming phase, the cumulative ALO AIMs severity was significantly diminished in animals that were primed with L-DOPA/ondansetron 0.0001 mg/kg.

Collectively, our results suggest that 5-HT₃ blockade acutely diminishes AIMs severity and interferes with the development of dyskinesia. Quite interestingly is the fact that, by looking at the different time course figures illustrating the development of AIMs during the *de novo* study, ondansetron appeared to have little symptomatic effect, which is in contradiction with the less severe AIMs when animals were administered L-DOPA alone after the priming phase. A possible explanation may be the development of tachyphylaxis to ondansetron. Further studies are needed to explore this possible tolerance to the therapeutic effect of the drug, as any administration in clinical settings would entail chronic intake of the drug; as such, tachyphylaxis might reduce the translational potential of ondansetron as a treatment for dyskinesia.

It is important to note that during the 22-day study, animals in the vehicle group had relatively moderate AIMs scores and consequently, any effect of treatment appears to be rather subtle. Furthermore, even during the L-DOPA challenge, several vehicle-treated animals had no to minimal dyskinesia, while amongst those that displayed dyskinesia, it was only to a moderate level, despite the fact that they all had significant rearing asymmetry at the cylinder test, indicative of severe nigrostriatal lesion. This finding was not unexpected and is in agreement with the literature (473, 965, 1036), as mentioned before, as several 6-OHDA-lesioned rats do not develop AIMs when exposed to L-DOPA, regardless of the severity of their nigrostriatal DA denervation. However, no or mild dyskinesia in the control group renders it more difficult to detect an effect in the active group, which is a limitation of conducting *de novo* studies in the 6-OHDA-lesioned rat. Here however, this consideration did not prevent us from finding significant results.

Although we have used the term “priming” on several instances in this Thesis, it should be pointed out that the existence of such a process has been debated. Indeed, the definition of

priming as receptor sensitivity (604, 699) or behavioural manifestation (441), depends on the research group. Moreover, several paradigms are used to study priming and some consider that priming occurs after a single administration of a dopaminergic drug whereas others indicate that at least two injections are required (441, 1037). Thus, it was proposed that dyskinesia development could be related to plastic changes induced by dopaminergic denervation in the striatum or accelerate aberrant changes induced by the dopaminergic lesion (440). Furthermore, the authors argue that DA replacement therapy may affect the propensity of treatment to elicit dyskinesia by increasing the likelihood of dyskinesia development, decreasing the threshold dose of drug needed to induce dyskinesia and shifting the dyskinesia dose-response curve to the left (474). Thus, the first intake and subsequent administration of L-DOPA or DA agonists may sensitize to the actual mechanisms that underlie LID but not induce them.

Chronic L-DOPA administration in the 6-OHDA-lesioned rat induces the expression of Δ FosB protein in the DA-denervated striatum of animals that develop LID (1038). Increased ERK1/2 phosphorylation correlates with increased Δ FosB and dyskinesia in the DA-depleted striatum of 6-OHDA-lesioned rats (974, 1039). Therefore, the use of biological molecular markers such as Δ FosB and ERK1/2 phosphorylation may provide a cellular measure of dyskinesia that is complementary to the behavioural assessment of dyskinesia. Further studies are needed to determine if *de novo* administration of 5-HT₃ antagonists interfere with these molecular changes that are associated with the dyskinetic state.

V. Conclusion

There remains many gaps in our understanding of the pathophysiology of LID, and this unmet medical need continues to undermine the quality of life of PD patients. Here, we have used the 5-HT₃ receptor antagonist ondansetron, because it is highly-selective and clinically available. Inasmuch as its high selectivity, the therapeutic benefit conferred by ondansetron is likely to be mediated exclusively through 5-HT₃ receptor blockade. Our results therefore indicate that 5-HT₃ receptor antagonism is a new and promising therapeutic strategy to alleviate established, and prevent the development of, LID in PD. Importantly, the anti-dyskinetic action was achieved without compromising L-DOPA anti-parkinsonian action, a problem that has hindered the development of several potential anti-dyskinetic approaches in the past few years (741, 1040-1042).

Further studies are needed to characterise the potential of this exciting new therapeutic paradigm to alleviate LID. Given the effect of acute challenges of ondansetron 0.0001 mg/kg on the severity of established ALO AIMs, it would be of further interest to investigate whether chronic administration of treatment can maintain this anti-dyskinetic effect. Hence, following the two-week induction priming phase, animals would be administered L-DOPA/ondansetron on a daily basis for three weeks and ALO AIMs severity would be assessed at regular intervals. This would differ from the *de novo* experiments that seek to assess the effect of treatment on the development of AIMs as here, L-DOPA is administered to the animals so that they already exhibit AIMs before L-DOPA/experimental drug is administered. In addition, experiments in the gold standard model of PD, the MPTP-lesioned nonhuman primate model, could validate the efficacy of 5-HT₃ receptor antagonism on dyskinesia as well as another complication of L-DOPA therapy, psychosis. Given the clinical availability of 5-HT₃ receptors, positive outcomes in the MPTP-lesioned primate model could facilitate the testing of these compounds in Phase IIa clinical trials in the context of dyskinesia.

VI. Bibliography

1. Parkinson J. An essay on the shaking palsy. Whittingham and Rowland for Sherwood, Neely and Jones. London 1817.
2. Charcot J. De la paralysie agitante. Lecons sur les maladies du systeme nerveux. Paris: A. Delahaye; 1872. p. 115-88.
3. de Lau LM, Breteler MM. Epidemiology of Parkinson's disease. *The Lancet Neurology*. 2006;5(6):525-35.
4. Alves G, Forsaa EB, Pedersen KF, Dreetz Gjerstad M, Larsen JP. Epidemiology of Parkinson's disease. *J Neurol*. 2008;255 Suppl 5:18-32.
5. Massano J, Bhatia KP. Clinical Approach to Parkinson's Disease: Features, Diagnosis, and Principles of Management. Cold Spring Harbor Perspectives in Medicine. 2012;2(6):a008870.
6. Chaudhuri KR, Naidu Y. Early Parkinson's disease and non-motor issues. *J Neurol*. 2008;255 Suppl 5:33-8.
7. Bernheimer H, Birkmayer W, Hornykiewicz O, Jellinger K, Seitelberger F. Brain dopamine and the syndromes of Parkinson and Huntington. Clinical, morphological and neurochemical correlations. *J Neurol Sci*. 1973;20(4):415-55.
8. Kish SJ, Shannak K, Hornykiewicz O. Uneven pattern of dopamine loss in the striatum of patients with idiopathic Parkinson's disease. Pathophysiologic and clinical implications. *N Engl J Med*. 1988;318(14):876-80.
9. Hornykiewicz O. Biochemical aspects of Parkinson's disease. *Neurology*. 1998;51(2 Suppl 2):S2-9.
10. Pakkenberg B, Moller A, Gundersen HJ, Mouritzen Dam A, Pakkenberg H. The absolute number of nerve cells in substantia nigra in normal subjects and in patients with Parkinson's disease estimated with an unbiased stereological method. *J Neurol Neurosurg Psychiatry*. 1991;54(1):30-3.
11. Mizuno Y, Hattori N, Kubo S, Sato S, Nishioka K, Hatano T, et al. Progress in the pathogenesis and genetics of Parkinson's disease. *Philosophical transactions of the Royal Society of London Series B, Biological sciences*. 2008;363(1500):2215-27.
12. Buddhala C, Loftin SK, Kuley BM, Cairns NJ, Campbell MC, Perlmutter JS, et al. Dopaminergic, serotonergic, and noradrenergic deficits in Parkinson disease. *Ann Clin Transl Neurol*. 2015;2(10):949-59.
13. Dexter DT, Jenner P. Parkinson disease: from pathology to molecular disease mechanisms. *Free Radic Biol Med*. 2013;62:132-44.
14. Kalia LV, Kalia SK, McLean PJ, Lozano AM, Lang AE. alpha-Synuclein oligomers and clinical implications for Parkinson disease. *Ann Neurol*. 2013;73(2):155-69.
15. Pacheco CR, Morales CN, Ramirez AE, Munoz FJ, Gallegos SS, Caviedes PA, et al. Extracellular alpha-synuclein alters synaptic transmission in brain neurons by perforating the neuronal plasma membrane. *J Neurochem*. 2015;132(6):731-41.
16. Kalia LV, Kalia SK. alpha-Synuclein and Lewy pathology in Parkinson's disease. *Curr Opin Neurol*. 2015;28(4):375-81.
17. Dawson TM, Dawson VL. Molecular pathways of neurodegeneration in Parkinson's disease. *Science*. 2003;302(5646):819-22.
18. Hely MA, Morris JG, Traficante R, Reid WG, O'Sullivan DJ, Williamson PM. The sydney multicentre study of Parkinson's disease: progression and mortality at 10 years. *J Neurol Neurosurg Psychiatry*. 1999;67(3):300-7.

19. Hely MA, Morris JG, Reid WG, Trafficante R. Sydney Multicenter Study of Parkinson's disease: non-L-dopa-responsive problems dominate at 15 years. *Mov Disord.* 2005;20(2):190-9.
20. Stocchi F, Olanow CW. Neuroprotection in Parkinson's disease: clinical trials. *Ann Neurol.* 2003;53 Suppl 3:S87-97; discussion S-9.
21. Bastide MF, Meissner WG, Picconi B, Fasano S, Fernagut PO, Feyder M, et al. Pathophysiology of L-dopa-induced motor and non-motor complications in Parkinson's disease. *Prog Neurobiol.* 2015;132:96-168.
22. Wong SL, Gilmour H, Ramage-Morin PL. Parkinson's disease: Prevalence, diagnosis and impact. *Health Rep.* 2014;25(11):10-4.
23. Pringsheim T, Jette N, Frolkis A, Steeves TD. The prevalence of Parkinson's disease: a systematic review and meta-analysis. *Mov Disord.* 2014;29(13):1583-90.
24. Hirsch L, Jette N, Frolkis A, Steeves T, Pringsheim T. The Incidence of Parkinson's Disease: A Systematic Review and Meta-Analysis. *Neuroepidemiology.* 2016;46(4):292-300.
25. Elbaz A, Carcaillon L, Kab S, Moisan F. Epidemiology of Parkinson's disease. *Revue neurologique.* 2016;172(1):14-26.
26. Huang Z, de la Fuente-Fernandez R, Stoessl AJ. Etiology of Parkinson's disease. *Can J Neurol Sci.* 2003;30 Suppl 1:S10-8.
27. Tysnes OB, Storstein A. Epidemiology of Parkinson's disease. *Journal of neural transmission (Vienna, Austria : 1996).* 2017;124(8):901-5.
28. von Campenhausen S, Bornschein B, Wick R, Botzel K, Sampaio C, Poewe W, et al. Prevalence and incidence of Parkinson's disease in Europe. *Eur Neuropsychopharmacol.* 2005;15(4):473-90.
29. de Rijk MC, Tzourio C, Breteler MM, Dartigues JF, Amaducci L, Lopez-Pousa S, et al. Prevalence of parkinsonism and Parkinson's disease in Europe: the EUROPARKINSON Collaborative Study. European Community Concerted Action on the Epidemiology of Parkinson's disease. *J Neurol Neurosurg Psychiatry.* 1997;62(1):10-5.
30. Okubadejo NU, Bower JH, Rocca WA, Maraganore DM. Parkinson's disease in Africa: A systematic review of epidemiologic and genetic studies. *Mov Disord.* 2006;21(12):2150-6.
31. Schoenberg BS, Anderson DW, Haerer AF. Prevalence of Parkinson's disease in the biracial population of Copiah County, Mississippi. *Neurology.* 1985;35(6):841-5.
32. Jendroska K, Olasode BJ, Daniel SE, Elliott L, Ogunniyi AO, Aghadiuno PU, et al. Incidental Lewy body disease in black Africans. *Lancet.* 1994;344(8926):882-3.
33. Zhang ZX, Roman GC. Worldwide occurrence of Parkinson's disease: an updated review. *Neuroepidemiology.* 1993;12(4):195-208.
34. Wang SJ, Fuh JL, Teng EL, Liu CY, Lin KP, Chen HM, et al. A door-to-door survey of Parkinson's disease in a Chinese population in Kinmen. *Arch Neurol.* 1996;53(1):66-71.
35. Muangpaisan W, Hori H, Brayne C. Systematic review of the prevalence and incidence of Parkinson's disease in Asia. *J Epidemiol.* 2009;19(6):281-93.
36. Chen RC, Chang SF, Su CL, Chen TH, Yen MF, Wu HM, et al. Prevalence, incidence, and mortality of PD: a door-to-door survey in Ilan county, Taiwan. *Neurology.* 2001;57(9):1679-86.
37. Zhang ZX, Anderson DW, Huang JB, Li H, Hong X, Wei J, et al. Prevalence of Parkinson's disease and related disorders in the elderly population of greater Beijing, China. *Mov Disord.* 2003;18(7):764-72.

38. Van Den Eeden SK, Tanner CM, Bernstein AL, Fross RD, Leimpeter A, Bloch DA, et al. Incidence of Parkinson's disease: variation by age, gender, and race/ethnicity. *Am J Epidemiol.* 2003;157(11):1015-22.
39. Chen S-Y, Tsai S-T. The Epidemiology of Parkinson's Disease. *Tzu Chi Medical Journal.* 2010;22(2):73-81.
40. Morens DM, Davis JW, Grandinetti A, Ross GW, Popper JS, White LR. Epidemiologic observations on Parkinson's disease: incidence and mortality in a prospective study of middle-aged men. *Neurology.* 1996;46(4):1044-50.
41. Mayeux R. Epidemiology of neurodegeneration. *Annu Rev Neurosci.* 2003;26:81-104.
42. Benito-Leon J, Bermejo-Pareja F, Rodriguez J, Molina JA, Gabriel R, Morales JM, et al. Prevalence of PD and other types of parkinsonism in three elderly populations of central Spain. *Mov Disord.* 2003;18(3):267-74.
43. Claveria LE, Duarte J, Sevillano MD, Perez-Sempere A, Cabezas C, Rodriguez F, et al. Prevalence of Parkinson's disease in Cantalejo, Spain: a door-to-door survey. *Mov Disord.* 2002;17(2):242-9.
44. Mayeux R, Marder K, Cote LJ, Denaro J, Hemenegildo N, Mejia H, et al. The frequency of idiopathic Parkinson's disease by age, ethnic group, and sex in northern Manhattan, 1988-1993. *Am J Epidemiol.* 1995;142(8):820-7.
45. Fall PA, Axelson O, Fredriksson M, Hansson G, Lindvall B, Olsson JE, et al. Age-standardized incidence and prevalence of Parkinson's disease in a Swedish community. *J Clin Epidemiol.* 1996;49(6):637-41.
46. Wooten GF, Currie LJ, Bovbjerg VE, Lee JK, Patrie J. Are men at greater risk for Parkinson's disease than women? *J Neurol Neurosurg Psychiatry.* 2004;75(4):637-9.
47. Elbaz A, Bower JH, Maraganore DM, McDonnell SK, Peterson BJ, Ahlskog JE, et al. Risk tables for parkinsonism and Parkinson's disease. *J Clin Epidemiol.* 2002;55(1):25-31.
48. Taylor KS, Cook JA, Counsell CE. Heterogeneity in male to female risk for Parkinson's disease. *J Neurol Neurosurg Psychiatry.* 2007;78(8):905-6.
49. de Rijk MC, Breteler MM, Graveland GA, Ott A, Grobbee DE, van der Meche FG, et al. Prevalence of Parkinson's disease in the elderly: the Rotterdam Study. *Neurology.* 1995;45(12):2143-6.
50. Tison F, Dartigues JF, Dubes L, Zuber M, Alperovitch A, Henry P. Prevalence of Parkinson's disease in the elderly: a population study in Gironde, France. *Acta Neurol Scand.* 1994;90(2):111-5.
51. de Lau LM, Giesbergen PC, de Rijk MC, Hofman A, Koudstaal PJ, Breteler MM. Incidence of parkinsonism and Parkinson disease in a general population: the Rotterdam Study. *Neurology.* 2004;63(7):1240-4.
52. Rajput AH, Offord KP, Beard CM, Kurland LT. Epidemiology of parkinsonism: incidence, classification, and mortality. *Ann Neurol.* 1984;16(3):278-82.
53. Bower JH, Maraganore DM, McDonnell SK, Rocca WA. Incidence and distribution of parkinsonism in Olmsted County, Minnesota, 1976-1990. *Neurology.* 1999;52(6):1214-20.
54. Saunders-Pullman R. Estrogens and Parkinson disease: neuroprotective, symptomatic, neither, or both? *Endocrine.* 2003;21(1):81-7.
55. Verstraeten A, Theuns J, Van Broeckhoven C. Progress in unraveling the genetic etiology of Parkinson disease in a genomic era. *Trends in genetics : TIG.* 2015;31(3):140-9.
56. Calne DB, Langston JW. Aetiology of Parkinson's disease. *Lancet.* 1983;2(8365-66):1457-9.

57. Schapira AH, Jenner P. Etiology and pathogenesis of Parkinson's disease. *Mov Disord.* 2011;26(6):1049-55.
58. Kalia LV, Lang AE. Parkinson's disease. *Lancet.* 2015;386(9996):896-912.
59. Bezard E, Przedborski S. A tale on animal models of Parkinson's disease. *Mov Disord.* 2011;26(6):993-1002.
60. Tanner CM, Aston DA. Epidemiology of Parkinson's disease and akinetic syndromes. *Curr Opin Neurol.* 2000;13(4):427-30.
61. Greener M. Pesticides and Parkinson's disease pathogenesis: the controversy continues. *Progress in Neurology and Psychiatry.* 2013;17(1):20-1.
62. Betarbet R, Sherer TB, MacKenzie G, Garcia-Osuna M, Panov AV, Greenamyre JT. Chronic systemic pesticide exposure reproduces features of Parkinson's disease. *Nat Neurosci.* 2000;3(12):1301-6.
63. Brooks AI, Chadwick CA, Gelbard HA, Cory-Slechta DA, Federoff HJ. Paraquat elicited neurobehavioral syndrome caused by dopaminergic neuron loss. *Brain Res.* 1999;823(1-2):1-10.
64. McCormack AL, Mak SK, Shenasa M, Langston WJ, Forno LS, Di Monte DA. Pathologic modifications of alpha-synuclein in 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-treated squirrel monkeys. *Journal of neuropathology and experimental neurology.* 2008;67(8):793-802.
65. Thiruchelvam M, McCormack A, Richfield EK, Baggs RB, Tank AW, Di Monte DA, et al. Age-related irreversible progressive nigrostriatal dopaminergic neurotoxicity in the paraquat and maneb model of the Parkinson's disease phenotype. *Eur J Neurosci.* 2003;18(3):589-600.
66. Ascherio A, Chen H, Weisskopf MG, O'Reilly E, McCullough ML, Calle EE, et al. Pesticide exposure and risk for Parkinson's disease. *Ann Neurol.* 2006;60(2):197-203.
67. Ferraz HB, Andrade LA, Tumas V, Calia LC, Borges V. Rural or urban living and Parkinson's disease. *Arquivos de neuro-psiquiatria.* 1996;54(1):37-41.
68. Gorell JM, Johnson CC, Rybicki BA, Peterson EL, Richardson RJ. The risk of Parkinson's disease with exposure to pesticides, farming, well water, and rural living. *Neurology.* 1998;50(5):1346-50.
69. Rybicki BA, Johnson CC, Uman J, Gorell JM. Parkinson's disease mortality and the industrial use of heavy metals in Michigan. *Mov Disord.* 1993;8(1):87-92.
70. Winkel R, Kuhn W, Przuntek H. Chronic intoxication with lead- and sulfur compounds may produce Parkinson's disease. *Journal of neural transmission Supplementum.* 1995;46:183-7.
71. Gorell JM, Johnson CC, Rybicki BA, Peterson EL, Kortsha GX, Brown GG, et al. Occupational exposure to manganese, copper, lead, iron, mercury and zinc and the risk of Parkinson's disease. *Neurotoxicology.* 1999;20(2-3):239-47.
72. Harvey C. Are there any consistent environmental risk factors for Parkinson's disease? *Occupational and Environmental Medicine.* 2011;68(Suppl 1):A1.
73. Dick FD, De Palma G, Ahmadi A, Scott NW, Prescott GJ, Bennett J, et al. Environmental risk factors for Parkinson's disease and parkinsonism: the Geoparkinson study. *Occup Environ Med.* 2007;64(10):666-72.
74. Langston JW, Ballard P, Tetrud JW, Irwin I. Chronic Parkinsonism in humans due to a product of meperidine-analog synthesis. *Science.* 1983;219(4587):979-80.
75. Heikkila RE, Hess A, Duvoisin RC. Dopaminergic neurotoxicity of 1-methyl-4-phenyl-1,2,5,6-tetrahydropyridine in mice. *Science.* 1984;224(4656):1451-3.

76. Schneider JS, Markham CH. Neurotoxic effects of N-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) in the cat. Tyrosine hydroxylase immunohistochemistry. *Brain Res.* 1986;373(1-2):258-67.
77. Arai N, Misugi K, Goshima Y, Misu Y. Evaluation of a 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-treated C57 black mouse model for parkinsonism. *Brain Res.* 1990;515(1-2):57-63.
78. Hantraye P, Brouillet E, Ferrante R, Palfi S, Dolan R, Matthews RT, et al. Inhibition of neuronal nitric oxide synthase prevents MPTP-induced parkinsonism in baboons. *Nat Med.* 1996;2(9):1017-21.
79. Araki T, Kumagai T, Tanaka K, Matsubara M, Kato H, Itoyama Y, et al. Neuroprotective effect of riluzole in MPTP-treated mice. *Brain Res.* 2001;918(1-2):176-81.
80. Araki T, Mikami T, Tanji H, Matsubara M, Imai Y, Mizugaki M, et al. Biochemical and immunohistological changes in the brain of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-treated mouse. *Eur J Pharm Sci.* 2001;12(3):231-8.
81. Watanabe Y, Himeda T, Araki T. Mechanisms of MPTP toxicity and their implications for therapy of Parkinson's disease. *Med Sci Monit.* 2005;11(1):RA17-23.
82. Tipton KF, Singer TP. Advances in our understanding of the mechanisms of the neurotoxicity of MPTP and related compounds. *J Neurochem.* 1993;61(4):1191-206.
83. Gainetdinov RR, Fumagalli F, Jones SR, Caron MG. Dopamine transporter is required for in vivo MPTP neurotoxicity: evidence from mice lacking the transporter. *J Neurochem.* 1997;69(3):1322-5.
84. Nicklas WJ, Youngster SK, Kindt MV, Heikkila RE. MPTP, MPP⁺ and mitochondrial function. *Life Sci.* 1987;40(8):721-9.
85. Hasegawa E, Takeshige K, Oishi T, Murai Y, Minakami S. 1-Methyl-4-phenylpyridinium (MPP⁺) induces NADH-dependent superoxide formation and enhances NADH-dependent lipid peroxidation in bovine heart submitochondrial particles. *Biochem Biophys Res Commun.* 1990;170(3):1049-55.
86. Chiueh CC, Miyake H, Peng MT. Role of dopamine autoxidation, hydroxyl radical generation, and calcium overload in underlying mechanisms involved in MPTP-induced parkinsonism. *Adv Neurol.* 1993;60:251-8.
87. Chiueh CC, Rauhala P. Free radicals and MPTP-induced selective destruction of substantia nigra compacta neurons. *Adv Pharmacol.* 1998;42:796-800.
88. Beal MF. Experimental models of Parkinson's disease. *Nat Rev Neurosci.* 2001;2(5):325-34.
89. Shimoji M, Zhang L, Mandir AS, Dawson VL, Dawson TM. Absence of inclusion body formation in the MPTP mouse model of Parkinson's disease. *Brain Res Mol Brain Res.* 2005;134(1):103-8.
90. Bellou V, Belbasis L, Tzoulaki I, Evangelou E, Ioannidis JP. Environmental risk factors and Parkinson's disease: An umbrella review of meta-analyses. *Parkinsonism Relat Disord.* 2016;23:1-9.
91. Hamer M, Chida Y. Physical activity and risk of neurodegenerative disease: a systematic review of prospective evidence. *Psychol Med.* 2009;39(1):3-11.
92. Xu Q, Park Y, Huang X, Hollenbeck A, Blair A, Schatzkin A, et al. Physical activities and future risk of Parkinson disease. *Neurology.* 2010;75(4):341-8.
93. Savica R, Rocca WA, Ahlskog JE. When does Parkinson disease start? *Arch Neurol.* 2010;67(7):798-801.

94. Braak H, de Vos RA, Bohl J, Del Tredici K. Gastric alpha-synuclein immunoreactive inclusions in Meissner's and Auerbach's plexuses in cases staged for Parkinson's disease-related brain pathology. *Neurosci Lett.* 2006;396(1):67-72.
95. Beach TG, Adler CH, Sue LI, Vedders L, Lue L, White Iii CL, et al. Multi-organ distribution of phosphorylated alpha-synuclein histopathology in subjects with Lewy body disorders. *Acta Neuropathol.* 2010;119(6):689-702.
96. Jafari S, Etminan M, Aminzadeh F, Samii A. Head injury and risk of Parkinson disease: a systematic review and meta-analysis. *Mov Disord.* 2013;28(9):1222-9.
97. Noyce AJ, Bestwick JP, Silveira-Moriyama L, Hawkes CH, Giovannoni G, Lees AJ, et al. Meta-analysis of early nonmotor features and risk factors for Parkinson disease. *Ann Neurol.* 2012;72(6):893-901.
98. Shen L, Ji HF. Low uric acid levels in patients with Parkinson's disease: evidence from meta-analysis. *BMJ Open.* 2013;3(11):e003620.
99. Zhang D, Jiang H, Xie J. Alcohol intake and risk of Parkinson's disease: a meta-analysis of observational studies. *Mov Disord.* 2014;29(6):819-22.
100. Ioannidis JPA. Clarifications on the application and interpretation of the test for excess significance and its extensions. *Journal of Mathematical Psychology.* 2013;57(5):184-7.
101. van der Mark M, Brouwer M, Kromhout H, Nijssen P, Huss A, Vermeulen R. Is pesticide use related to Parkinson disease? Some clues to heterogeneity in study results. *Environ Health Perspect.* 2012;120(3):340-7.
102. Gillies GE, Pienaar IS, Vohra S, Qamhawi Z. Sex differences in Parkinson's disease. *Frontiers in Neuroendocrinology.* 2014;35(3):370-84.
103. Moisan F, Kab S, Mohamed F, Canonico M, Le Guern M, Quintin C, et al. Parkinson disease male-to-female ratios increase with age: French nationwide study and meta-analysis. *Journal of Neurology, Neurosurgery & Psychiatry.* 2015.
104. Tanner CM, Goldman SM. Epidemiology of Parkinson's disease. *Neurologic clinics.* 1996;14(2):317-35.
105. Sveinbjornsdottir S, Hicks AA, Jonsson T, Petursson H, Gugmundsson G, Frigge ML, et al. Familial aggregation of Parkinson's disease in Iceland. *N Engl J Med.* 2000;343(24):1765-70.
106. Polymeropoulos MH, Lavedan C, Leroy E, Ide SE, Dehejia A, Dutra A, et al. Mutation in the alpha-synuclein gene identified in families with Parkinson's disease. *Science.* 1997;276(5321):2045-7.
107. Kitada T, Asakawa S, Hattori N, Matsumine H, Yamamura Y, Minoshima S, et al. Mutations in the parkin gene cause autosomal recessive juvenile parkinsonism. *Nature.* 1998;392(6676):605-8.
108. Gasser T, Muller-Myhsok B, Wszolek ZK, Oehlmann R, Calne DB, Bonifati V, et al. A susceptibility locus for Parkinson's disease maps to chromosome 2p13. *Nat Genet.* 1998;18(3):262-5.
109. Kruger R, Kuhn W, Muller T, Woitalla D, Graeber M, Kosel S, et al. Ala30Pro mutation in the gene encoding alpha-synuclein in Parkinson's disease. *Nat Genet.* 1998;18(2):106-8.
110. Calne S, Schoenberg B, Martin W, Uitti RJ, Spencer P, Calne DB. Familial Parkinson's disease: possible role of environmental factors. *Can J Neurol Sci.* 1987;14(3):303-5.
111. de la Fuente-Fernandez R, Calne DB. Familial aggregation of Parkinson's disease. *N Engl J Med.* 2001;344(15):1168.

112. Tanner CM, Ottman R, Goldman SM, Ellenberg J, Chan P, Mayeux R, et al. Parkinson disease in twins: an etiologic study. *JAMA*. 1999;281(4):341-6.
113. Piccini P, Burn DJ, Ceravolo R, Maraganore D, Brooks DJ. The role of inheritance in sporadic Parkinson's disease: evidence from a longitudinal study of dopaminergic function in twins. *Ann Neurol*. 1999;45(5):577-82.
114. Hardy J. Genetic analysis of pathways to Parkinson disease. *Neuron*. 2010;68(2):201-6.
115. Martin I, Dawson VL, Dawson TM. Recent advances in the genetics of Parkinson's disease. *Annu Rev Genomics Hum Genet*. 2011;12:301-25.
116. Klein C, Westenberger A. Genetics of Parkinson's disease. *Cold Spring Harb Perspect Med*. 2012;2(1):a008888.
117. Thomas B, Beal MF. Parkinson's disease. *Human molecular genetics*. 2007;16 Spec No. 2:R183-94.
118. Vilarino-Guell C, Wider C, Ross OA, Dachsel JC, Kachergus JM, Lincoln SJ, et al. VPS35 mutations in Parkinson disease. *Am J Hum Genet*. 2011;89(1):162-7.
119. Zimprich A, Benet-Pages A, Struhal W, Graf E, Eck SH, Offman MN, et al. A mutation in VPS35, encoding a subunit of the retromer complex, causes late-onset Parkinson disease. *Am J Hum Genet*. 2011;89(1):168-75.
120. Chartier-Harlin MC, Dachsel JC, Vilarino-Guell C, Lincoln SJ, Lepretre F, Hulihan MM, et al. Translation initiator EIF4G1 mutations in familial Parkinson disease. *Am J Hum Genet*. 2011;89(3):398-406.
121. Maraganore DM, de Andrade M, Elbaz A, Farrer MJ, Ioannidis JP, Kruger R, et al. Collaborative analysis of alpha-synuclein gene promoter variability and Parkinson disease. *JAMA*. 2006;296(6):661-70.
122. Zabetian CP, Yamamoto M, Lopez AN, Ujike H, Mata IF, Izumi Y, et al. LRRK2 mutations and risk variants in Japanese patients with Parkinson's disease. *Mov Disord*. 2009;24(7):1034-41.
123. Goris A, Williams-Gray CH, Clark GR, Foltynie T, Lewis SJ, Brown J, et al. Tau and alpha-synuclein in susceptibility to, and dementia in, Parkinson's disease. *Ann Neurol*. 2007;62(2):145-53.
124. Sidransky E, Samadpour T, Tayebi N. Mutations in GBA are associated with familial Parkinson disease susceptibility and age at onset. *Neurology*. 2009;73(17):1424-5, author reply 5-6.
125. Spencer CC, Plagnol V, Strange A, Gardner M, Paisan-Ruiz C, Band G, et al. Dissection of the genetics of Parkinson's disease identifies an additional association 5' of SNCA and multiple associated haplotypes at 17q21. *Human molecular genetics*. 2011;20(2):345-53.
126. Xiromerisiou G, Dardiotis E, Tsimourtou V, Kountouras PM, Paterakis KN, Kapsalaki EZ, et al. Genetic basis of Parkinson disease. *Neurosurgical focus*. 2010;28(1):E7.
127. Tan EK, Khajavi M, Thornby JI, Nagamitsu S, Jankovic J, Ashizawa T. Variability and validity of polymorphism association studies in Parkinson's disease. *Neurology*. 2000;55(4):533-8.
128. Maraganore DM, Farrer MJ, McDonnell SK, Elbaz A, Schaid DJ, Hardy JA, et al. Case-control study of estrogen receptor gene polymorphisms in Parkinson's disease. *Mov Disord*. 2002;17(3):509-12.
129. Healy D, Abou-Sleiman P, Lees A, Casas J, Quinn N, Bhatia K, et al. Tau gene and Parkinson's disease: a case-control study and meta-analysis. *Journal of Neurology, Neurosurgery, and Psychiatry*. 2004;75(7):962-5.

130. Huang X, Chen PC, Poole C. APOE-[epsilon]2 allele associated with higher prevalence of sporadic Parkinson disease. *Neurology*. 2004;62(12):2198-202.
131. Houlden H, Singleton AB. The genetics and neuropathology of Parkinson's disease. *Acta Neuropathol*. 2012;124(3):325-38.
132. Grabowski GA. Phenotype, diagnosis, and treatment of Gaucher's disease. *Lancet*. 2008;372(9645):1263-71.
133. Neumann J, Bras J, Deas E, O'Sullivan SS, Parkkinen L, Lachmann RH, et al. Glucocerebrosidase mutations in clinical and pathologically proven Parkinson's disease. *Brain*. 2009;132(Pt 7):1783-94.
134. Sidransky E, Nalls MA, Aasly JO, Aharon-Peretz J, Annesi G, Barbosa ER, et al. Multicenter analysis of glucocerebrosidase mutations in Parkinson's disease. *N Engl J Med*. 2009;361(17):1651-61.
135. Lesage S, Anheim M, Condroyer C, Pollak P, Durif F, Dupuits C, et al. Large-scale screening of the Gaucher's disease-related glucocerebrosidase gene in Europeans with Parkinson's disease. *Human molecular genetics*. 2011;20(1):202-10.
136. McNeill A, Duran R, Hughes DA, Mehta A, Schapira AH. A clinical and family history study of Parkinson's disease in heterozygous glucocerebrosidase mutation carriers. *J Neurol Neurosurg Psychiatry*. 2012;83(8):853-4.
137. Kumar KR, Ramirez A, Gobel A, Kresojevic N, Svetel M, Lohmann K, et al. Glucocerebrosidase mutations in a Serbian Parkinson's disease population. *Eur J Neurol*. 2013;20(2):402-5.
138. Anheim M, Elbaz A, Lesage S, Durr A, Condroyer C, Viallet F, et al. Penetrance of Parkinson disease in glucocerebrosidase gene mutation carriers. *Neurology*. 2012;78(6):417-20.
139. Spatola M, Wider C. Genetics of Parkinson's disease: the yield. *Parkinsonism Relat Disord*. 2014;20 Suppl 1:S35-8.
140. Velayati A, Yu WH, Sidransky E. The role of glucocerebrosidase mutations in Parkinson disease and Lewy body disorders. *Curr Neurol Neurosci Rep*. 2010;10(3):190-8.
141. Du TT, Wang L, Duan CL, Lu LL, Zhang JL, Gao G, et al. GBA deficiency promotes SNCA/alpha-synuclein accumulation through autophagic inhibition by inactivated PPP2A. *Autophagy*. 2015;11(10):1803-20.
142. Spillantini MG, Schmidt ML, Lee VM, Trojanowski JQ, Jakes R, Goedert M. Alpha-synuclein in Lewy bodies. *Nature*. 1997;388(6645):839-40.
143. Spencer B, Potkar R, Trejo M, Rockenstein E, Patrick C, Gindi R, et al. Beclin 1 gene transfer activates autophagy and ameliorates the neurodegenerative pathology in alpha-synuclein models of Parkinson's and Lewy body diseases. *J Neurosci*. 2009;29(43):13578-88.
144. Wong K, Sidransky E, Verma A, Mixon T, Sandberg GD, Wakefield LK, et al. Neuropathology provides clues to the pathophysiology of Gaucher disease. *Molecular genetics and metabolism*. 2004;82(3):192-207.
145. Cullen V, Sardi SP, Ng J, Xu YH, Sun Y, Tomlinson JJ, et al. Acid beta-glucosidase mutants linked to Gaucher disease, Parkinson disease, and Lewy body dementia alter alpha-synuclein processing. *Ann Neurol*. 2011;69(6):940-53.
146. Mazzulli JR, Xu YH, Sun Y, Knight AL, McLean PJ, Caldwell GA, et al. Gaucher disease glucocerebrosidase and alpha-synuclein form a bidirectional pathogenic loop in synucleinopathies. *Cell*. 2011;146(1):37-52.
147. Lee S-J, Desplats P, Lee H-J, Spencer B, Masliah E. Cell-to-Cell Transmission of α -Synuclein Aggregates. *Methods in molecular biology (Clifton, NJ)*. 2012;849:347-59.

148. Guo JL, Lee VMY. Cell-to-cell transmission of pathogenic proteins in neurodegenerative diseases. *Nature medicine*. 2014;20(2):130-8.
149. Moussaud S, Jones DR, Moussaud-Lamodière EL, Delenclos M, Ross OA, McLean PJ. Alpha-synuclein and tau: teammates in neurodegeneration? *Molecular neurodegeneration*. 2014;9:43.
150. Wider C, Vilarino-Guell C, Jasinska-Myga B, Heckman MG, Soto-Ortolaza AI, Cobb SA, et al. Association of the MAPT locus with Parkinson's disease. *Eur J Neurol*. 2010;17(3):483-6.
151. Martin ER, Scott WK, Nance MA, Watts RL, Hubble JP, Koller WC, et al. Association of single-nucleotide polymorphisms of the tau gene with late-onset Parkinson disease. *Jama*. 2001;286(18):2245-50.
152. Singleton A, Myers A, Hardy J. The law of mass action applied to neurodegenerative disease: a hypothesis concerning the etiology and pathogenesis of complex diseases. *Human molecular genetics*. 2004;13 Spec No 1:R123-6.
153. Jellinger KA. Absence of alpha-synuclein pathology in postencephalitic parkinsonism. *Acta Neuropathol*. 2009;118(3):371-9.
154. Lee VM, Giasson BI, Trojanowski JQ. More than just two peas in a pod: common amyloidogenic properties of tau and alpha-synuclein in neurodegenerative diseases. *Trends in neurosciences*. 2004;27(3):129-34.
155. Clinton LK, Blurton-Jones M, Myczek K, Trojanowski JQ, LaFerla FM. Synergistic Interactions between Abeta, tau, and alpha-synuclein: acceleration of neuropathology and cognitive decline. *J Neurosci*. 2010;30(21):7281-9.
156. Lai HJ, Lin CH, Wu RM. Early-onset autosomal-recessive parkinsonian-pyramidal syndrome. *Acta Neurol Taiwan*. 2012;21(3):99-107.
157. Dachsel JC, Farrer MJ. LRRK2 and Parkinson disease. *Arch Neurol*. 2010;67(5):542-7.
158. Nichols WC, Pankratz N, Hernandez D, Paisan-Ruiz C, Jain S, Halter CA, et al. Genetic screening for a single common LRRK2 mutation in familial Parkinson's disease. *Lancet*. 2005;365(9457):410-2.
159. Di Fonzo A, Rohe CF, Ferreira J, Chien HF, Vacca L, Stocchi F, et al. A frequent LRRK2 gene mutation associated with autosomal dominant Parkinson's disease. *Lancet*. 2005;365(9457):412-5.
160. Kachergus J, Mata IF, Hulihan M, Taylor JP, Lincoln S, Aasly J, et al. Identification of a novel LRRK2 mutation linked to autosomal dominant parkinsonism: evidence of a common founder across European populations. *Am J Hum Genet*. 2005;76(4):672-80.
161. Ozelius LJ, Senthil G, Saunders-Pullman R, Ohmann E, Deligtisch A, Tagliati M, et al. LRRK2 G2019S as a cause of Parkinson's disease in Ashkenazi Jews. *N Engl J Med*. 2006;354(4):424-5.
162. Lesage S, Durr A, Tazir M, Lohmann E, Leutenegger AL, Janin S, et al. LRRK2 G2019S as a cause of Parkinson's disease in North African Arabs. *N Engl J Med*. 2006;354(4):422-3.
163. Gasser T. Mendelian forms of Parkinson's disease. *Biochim Biophys Acta*. 2009;1792(7):587-96.
164. Gaig C, Marti MJ, Ezquerra M, Rey MJ, Cardozo A, Tolosa E. G2019S LRRK2 mutation causing Parkinson's disease without Lewy bodies. *J Neurol Neurosurg Psychiatry*. 2007;78(6):626-8.

165. Marti-Masso JF, Ruiz-Martinez J, Bolano MJ, Ruiz I, Gorostidi A, Moreno F, et al. Neuropathology of Parkinson's disease with the R1441G mutation in LRRK2. *Mov Disord.* 2009;24(13):1998-2001.
166. Mori H, Kondo T, Yokochi M, Matsumine H, Nakagawa-Hattori Y, Miyake T, et al. Pathologic and biochemical studies of juvenile parkinsonism linked to chromosome 6q. *Neurology.* 1998;51(3):890-2.
167. Takahashi H, Ohama E, Suzuki S, Horikawa Y, Ishikawa A, Morita T, et al. Familial juvenile parkinsonism: clinical and pathologic study in a family. *Neurology.* 1994;44(3 Pt 1):437-41.
168. Greten-Harrison B, Polydoro M, Morimoto-Tomita M, Diao L, Williams AM, Nie EH, et al. alphasynuclein triple knockout mice reveal age-dependent neuronal dysfunction. *Proc Natl Acad Sci U S A.* 2010;107(45):19573-8.
169. Smith WW, Pei Z, Jiang H, Dawson VL, Dawson TM, Ross CA. Kinase activity of mutant LRRK2 mediates neuronal toxicity. *Nat Neurosci.* 2006;9(10):1231-3.
170. Lee BD, Shin JH, VanKampen J, Petrucelli L, West AB, Ko HS, et al. Inhibitors of leucine-rich repeat kinase-2 protect against models of Parkinson's disease. *Nat Med.* 2010;16(9):998-1000.
171. Hernandez DG, Reed X, Singleton AB. Genetics in Parkinson disease: Mendelian vs. non-Mendelian inheritance. *Journal of neurochemistry.* 2016;139(Suppl 1):59-74.
172. Nuytemans K, Theuns J, Cruts M, Van Broeckhoven C. Genetic etiology of Parkinson disease associated with mutations in the SNCA, PARK2, PINK1, PARK7, and LRRK2 genes: a mutation update. *Hum Mutat.* 2010;31(7):763-80.
173. Moore DJ, West AB, Dawson VL, Dawson TM. Molecular pathophysiology of Parkinson's disease. *Annu Rev Neurosci.* 2005;28:57-87.
174. Burre J, Sharma M, Tsetsenis T, Buchman V, Etherton MR, Sudhof TC. Alpha-synuclein promotes SNARE-complex assembly in vivo and in vitro. *Science.* 2010;329(5999):1663-7.
175. Chandra S, Gallardo G, Fernandez-Chacon R, Schluter OM, Sudhof TC. Alpha-synuclein cooperates with CSPalpha in preventing neurodegeneration. *Cell.* 2005;123(3):383-96.
176. Bonifacino JS, Hurley JH. Retromer. *Curr Opin Cell Biol.* 2008;20(4):427-36.
177. Kumar KR, Weissbach A, Heldmann M, Kasten M, Tunc S, Sue CM, et al. Frequency of the D620N mutation in VPS35 in Parkinson disease. *Arch Neurol.* 2012;69(10):1360-4.
178. Tang FL, Liu W, Hu JX, Erion JR, Ye J, Mei L, et al. VPS35 Deficiency or Mutation Causes Dopaminergic Neuronal Loss by Impairing Mitochondrial Fusion and Function. *Cell Rep.* 2015;12(10):1631-43.
179. Tsika E, Glauser L, Moser R, Fiser A, Daniel G, Sheerin UM, et al. Parkinson's disease-linked mutations in VPS35 induce dopaminergic neurodegeneration. *Human molecular genetics.* 2014;23(17):4621-38.
180. Wang W, Wang X, Fujioka H, Hoppel C, Whone AL, Caldwell MA, et al. Parkinson's disease-associated mutant VPS35 causes mitochondrial dysfunction by recycling DLP1 complexes. *Nat Med.* 2016;22(1):54-63.
181. Williams ET, Chen X, Moore DJ. VPS35, the Retromer Complex and Parkinson's Disease. *Journal of Parkinson's Disease.* 2017;7(2):219-33.
182. Castelo-Branco G, Wagner J, Rodriguez FJ, Kele J, Sousa K, Rawal N, et al. Differential regulation of midbrain dopaminergic neuron development by Wnt-1, Wnt-3a, and Wnt-5a. *Proc Natl Acad Sci U S A.* 2003;100(22):12747-52.

183. Port F, Kuster M, Herr P, Furger E, Banziger C, Hausmann G, et al. Wingless secretion promotes and requires retromer-dependent cycling of Wntless. *Nat Cell Biol.* 2008;10(2):178-85.
184. Deng H, Gao K, Jankovic J. The VPS35 gene and Parkinson's disease. *Mov Disord.* 2013;28(5):569-75.
185. Tabuchi M, Yanatori I, Kawai Y, Kishi F. Retromer-mediated direct sorting is required for proper endosomal recycling of the mammalian iron transporter DMT1. *J Cell Sci.* 2010;123(Pt 5):756-66.
186. Miura E, Hasegawa T, Konno M, Suzuki M, Sugeno N, Fujikake N, et al. VPS35 dysfunction impairs lysosomal degradation of alpha-synuclein and exacerbates neurotoxicity in a *Drosophila* model of Parkinson's disease. *Neurobiol Dis.* 2014;71:1-13.
187. MacLeod DA, Rhinn H, Kuwahara T, Zolin A, Di Paolo G, McCabe BD, et al. RAB7L1 interacts with LRRK2 to modify intraneuronal protein sorting and Parkinson's disease risk. *Neuron.* 2013;77(3):425-39.
188. Linhart R, Wong SA, Cao J, Tran M, Huynh A, Ardrey C, et al. Vacuolar protein sorting 35 (Vps35) rescues locomotor deficits and shortened lifespan in *Drosophila* expressing a Parkinson's disease mutant of Leucine-Rich Repeat Kinase 2 (LRRK2). *Molecular neurodegeneration.* 2014;9:23.
189. Dhungel N, Eleuteri S, Li LB, Kramer NJ, Chartron JW, Spencer B, et al. Parkinson's disease genes VPS35 and EIF4G1 interact genetically and converge on alpha-synuclein. *Neuron.* 2015;85(1):76-87.
190. Malik BR, Godena VK, Whitworth AJ. VPS35 pathogenic mutations confer no dominant toxicity but partial loss of function in *Drosophila* and genetically interact with parkin. *Human molecular genetics.* 2015;24(21):6106-17.
191. Lucking CB, Durr A, Bonifati V, Vaughan J, De Michele G, Gasser T, et al. Association between early-onset Parkinson's disease and mutations in the parkin gene. *N Engl J Med.* 2000;342(21):1560-7.
192. Abbas N, Lucking CB, Ricard S, Durr A, Bonifati V, De Michele G, et al. A wide variety of mutations in the parkin gene are responsible for autosomal recessive parkinsonism in Europe. French Parkinson's Disease Genetics Study Group and the European Consortium on Genetic Susceptibility in Parkinson's Disease. *Human molecular genetics.* 1999;8(4):567-74.
193. Mullin S, Schapira AH. Pathogenic mechanisms of neurodegeneration in Parkinson disease. *Neurologic clinics.* 2015;33(1):1-17.
194. Ramirez A, Heimbach A, Grundemann J, Stiller B, Hampshire D, Cid LP, et al. Hereditary parkinsonism with dementia is caused by mutations in ATP13A2, encoding a lysosomal type 5 P-type ATPase. *Nat Genet.* 2006;38(10):1184-91.
195. Djarmati A, Hagenah J, Reetz K, Winkler S, Behrens MI, Pawlack H, et al. ATP13A2 variants in early-onset Parkinson's disease patients and controls. *Mov Disord.* 2009;24(14):2104-11.
196. Gitler AD, Chesi A, Geddie ML, Strathearn KE, Hamamichi S, Hill KJ, et al. [alpha]-Synuclein is part of a diverse and highly conserved interaction network that includes PARK9 and manganese toxicity. *Nat Genet.* 2009;41(3):308-15.
197. Di Fonzo A, Dekker MC, Montagna P, Baruzzi A, Yonova EH, Correia Guedes L, et al. FBXO7 mutations cause autosomal recessive, early-onset parkinsonian-pyramidal syndrome. *Neurology.* 2009;72(3):240-5.

198. Paisan-Ruiz C, Guevara R, Federoff M, Hanagasi H, Sina F, Elahi E, et al. Early-onset L-dopa-responsive parkinsonism with pyramidal signs due to ATP13A2, PLA2G6, FBXO7 and spatacsin mutations. *Mov Disord.* 2010;25(12):1791-800.
199. Morgan NV, Westaway SK, Morton JE, Gregory A, Gissen P, Sonek S, et al. PLA2G6, encoding a phospholipase A2, is mutated in neurodegenerative disorders with high brain iron. *Nat Genet.* 2006;38(7):752-4.
200. Doherty KM, Silveira-Moriyama L, Parkkinen L, Healy DG, Farrell M, Mencacci NE, et al. Parkin disease: a clinicopathologic entity? *JAMA neurology.* 2013;70(5):571-9.
201. Paisan-Ruiz C, Bhatia KP, Li A, Hernandez D, Davis M, Wood NW, et al. Characterization of PLA2G6 as a locus for dystonia-parkinsonism. *Ann Neurol.* 2009;65(1):19-23.
202. Sina F, Shojaee S, Elahi E, Paisan-Ruiz C. R632W mutation in PLA2G6 segregates with dystonia-parkinsonism in a consanguineous Iranian family. *Eur J Neurol.* 2009;16(1):101-4.
203. Yoshino H, Tomiyama H, Tachibana N, Ogaki K, Li Y, Funayama M, et al. Phenotypic spectrum of patients with PLA2G6 mutation and PARK14-linked parkinsonism. *Neurology.* 2010;75(15):1356-61.
204. Lesage S, Brice A. Parkinson's disease: from monogenic forms to genetic susceptibility factors. *Human molecular genetics.* 2009;18(R1):R48-59.
205. Braak H, Rüb U, Gai WP, Del Tredici K. Idiopathic Parkinson's disease: possible routes by which vulnerable neuronal types may be subject to neuroinvasion by an unknown pathogen. *Journal of Neural Transmission.* 2003;110(5):517-36.
206. Braak H, Tredici KD, Rüb U, de Vos RAI, Jansen Steur ENH, Braak E. Staging of brain pathology related to sporadic Parkinson's disease. *Neurobiology of Aging.* 2003;24(2):197-211.
207. Visanji NP, Brooks PL, Hazrati L-N, Lang AE. The prion hypothesis in Parkinson's disease: Braak to the future. *Acta Neuropathologica Communications.* 2013;1:2-.
208. Braak H, Rub U, Jansen Steur EN, Del Tredici K, de Vos RA. Cognitive status correlates with neuropathologic stage in Parkinson disease. *Neurology.* 2005;64(8):1404-10.
209. Luk KC, Song C, O'Brien P, Stieber A, Branch JR, Brunden KR, et al. Exogenous alpha-synuclein fibrils seed the formation of Lewy body-like intracellular inclusions in cultured cells. *Proc Natl Acad Sci U S A.* 2009;106(47):20051-6.
210. Desplats P, Lee HJ, Bae EJ, Patrick C, Rockenstein E, Crews L, et al. Inclusion formation and neuronal cell death through neuron-to-neuron transmission of alpha-synuclein. *Proc Natl Acad Sci U S A.* 2009;106(31):13010-5.
211. Hansen C, Angot E, Bergstrom AL, Steiner JA, Pieri L, Paul G, et al. alpha-Synuclein propagates from mouse brain to grafted dopaminergic neurons and seeds aggregation in cultured human cells. *J Clin Invest.* 2011;121(2):715-25.
212. Mougnot AL, Nicot S, Bencsik A, Morignat E, Verchere J, Lakhdar L, et al. Prion-like acceleration of a synucleinopathy in a transgenic mouse model. *Neurobiol Aging.* 2012;33(9):2225-8.
213. Bloch A, Probst A, Bissig H, Adams H, Tolnay M. α -Synuclein pathology of the spinal and peripheral autonomic nervous system in neurologically unimpaired elderly subjects. *Neuropathology and Applied Neurobiology.* 2006;32(3):284-95.
214. Ross GW, Petrovitch H, Abbott RD, Tanner CM, Popper J, Masaki K, et al. Association of olfactory dysfunction with risk for future Parkinson's disease. *Ann Neurol.* 2008;63(2):167-73.

215. Jellinger KA. A critical reappraisal of current staging of Lewy-related pathology in human brain. *Acta Neuropathol.* 2008;116(1):1-16.
216. Zaccai J, Brayne C, McKeith I, Matthews F, Ince PG. Patterns and stages of alpha-synucleinopathy: Relevance in a population-based cohort. *Neurology.* 2008;70(13):1042-8.
217. Beach TG, Adler CH, Lue L, Sue LI, Bachalakuri J, Henry-Watson J, et al. Unified staging system for Lewy body disorders: correlation with nigrostriatal degeneration, cognitive impairment and motor dysfunction. *Acta Neuropathol.* 2009;117(6):613-34.
218. Kalaitzakis ME, Graeber MB, Gentleman SM, Pearce RKB. Evidence against a reliable staging system of α -synuclein pathology in Parkinson's disease. *Neuropathology and Applied Neurobiology.* 2009;35(1):125-6.
219. Kalaitzakis ME, Graeber MB, Gentleman SM, Pearce RKB. Controversies over the staging of α -synuclein pathology in Parkinson's disease. *Acta Neuropathologica.* 2008;116(1):125.
220. Rietdijk CD, Perez-Pardo P, Garssen J, van Wezel RJA, Kraneveld AD. Exploring Braak's Hypothesis of Parkinson's Disease. *Frontiers in Neurology.* 2017;8(37).
221. McNaught KS, Olanow CW, Halliwell B, Isacson O, Jenner P. Failure of the ubiquitin-proteasome system in Parkinson's disease. *Nat Rev Neurosci.* 2001;2(8):589-94.
222. Olanow CW, McNaught KS. Ubiquitin-proteasome system and Parkinson's disease. *Mov Disord.* 2006;21(11):1806-23.
223. Lim K-L, Tan JMM. Role of the ubiquitin proteasome system in Parkinson's disease. *BMC Biochemistry.* 2007;8(Suppl 1):S13-S.
224. Ross CA, Pickart CM. The ubiquitin-proteasome pathway in Parkinson's disease and other neurodegenerative diseases. *Trends Cell Biol.* 2004;14(12):703-11.
225. Giasson BI, Lee VMY. Are Ubiquitination Pathways Central to Parkinson's Disease? *Cell.* 2003;114(1):1-8.
226. McNaught KS, Perl DP, Brownell AL, Olanow CW. Systemic exposure to proteasome inhibitors causes a progressive model of Parkinson's disease. *Ann Neurol.* 2004;56(1):149-62.
227. Cook C, Petrucelli L. A critical evaluation of the ubiquitin-proteasome system in Parkinson's disease. *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease.* 2009;1792(7):664-75.
228. Walden H, Muqit MMK. Ubiquitin and Parkinson's disease through the looking glass of genetics. *Biochemical Journal.* 2017;474(9):1439-51.
229. Spillantini MG, Crowther RA, Jakes R, Hasegawa M, Goedert M. α -Synuclein in filamentous inclusions of Lewy bodies from Parkinson's disease and dementia with Lewy bodies. *Proceedings of the National Academy of Sciences of the United States of America.* 1998;95(11):6469-73.
230. Engelender S. Ubiquitination of alpha-synuclein and autophagy in Parkinson's disease. *Autophagy.* 2008;4(3):372-4.
231. Taylor JM, Song YJ, Huang Y, Farrer MJ, Delatycki MB, Halliday GM, et al. Parkin Co-Regulated Gene (PACRG) is regulated by the ubiquitin-proteasomal system and is present in the pathological features of Parkinsonian diseases. *Neurobiol Dis.* 2007;27(2):238-47.
232. Michel Patrick P, Hirsch Etienne C, Hunot S. Understanding Dopaminergic Cell Death Pathways in Parkinson Disease. *Neuron.* 2016;90(4):675-91.
233. Cookson MR, van der Brug M. Cell systems and the toxic mechanism(s) of alpha-synuclein. *Exp Neurol.* 2008;209(1):5-11.

234. Marques O, Outeiro TF. Alpha-synuclein: from secretion to dysfunction and death. *Cell Death Dis.* 2012;3:e350.
235. Martin LJ, Pan Y, Price AC, Sterling W, Copeland NG, Jenkins NA, et al. Parkinson's disease alpha-synuclein transgenic mice develop neuronal mitochondrial degeneration and cell death. *J Neurosci.* 2006;26(1):41-50.
236. Devi L, Raghavendran V, Prabhu BM, Avadhani NG, Anandatheerthavarada HK. Mitochondrial import and accumulation of alpha-synuclein impair complex I in human dopaminergic neuronal cultures and Parkinson disease brain. *J Biol Chem.* 2008;283(14):9089-100.
237. Shavali S, Brown-Borg HM, Ebadi M, Porter J. Mitochondrial localization of alpha-synuclein protein in alpha-synuclein overexpressing cells. *Neurosci Lett.* 2008;439(2):125-8.
238. Subramaniam M, Althof D, Gispert S, Schwenk J, Auburger G, Kulik A, et al. Mutant alpha-synuclein enhances firing frequencies in dopamine substantia nigra neurons by oxidative impairment of A-type potassium channels. *J Neurosci.* 2014;34(41):13586-99.
239. Schipper HM, Liberman A, Stopa EG. Neural heme oxygenase-1 expression in idiopathic Parkinson's disease. *Exp Neurol.* 1998;150(1):60-8.
240. Paxinou E, Chen Q, Weisse M, Giasson BI, Norris EH, Rueter SM, et al. Induction of alpha-synuclein aggregation by intracellular nitrative insult. *J Neurosci.* 2001;21(20):8053-61.
241. Ischiropoulos H, Beckman JS. Oxidative stress and nitration in neurodegeneration: cause, effect, or association? *J Clin Invest.* 2003;111(2):163-9.
242. Hashimoto M, Hsu LJ, Xia Y, Takeda A, Sisk A, Sundsmo M, et al. Oxidative stress induces amyloid-like aggregate formation of NACP/alpha-synuclein in vitro. *Neuroreport.* 1999;10(4):717-21.
243. Conway KA, Harper JD, Lansbury PT. Accelerated in vitro fibril formation by a mutant alpha-synuclein linked to early-onset Parkinson disease. *Nat Med.* 1998;4(11):1318-20.
244. Tsigelny IF, Crews L, Desplats P, Shaked GM, Sharikov Y, Mizuno H, et al. Mechanisms of hybrid oligomer formation in the pathogenesis of combined Alzheimer's and Parkinson's diseases. *PLoS One.* 2008;3(9):e3135.
245. Oueslati A, Fournier M, Lashuel HA. Role of post-translational modifications in modulating the structure, function and toxicity of alpha-synuclein: implications for Parkinson's disease pathogenesis and therapies. *Prog Brain Res.* 2010;183:115-45.
246. Taschenberger G, Garrido M, Tereshchenko Y, Bahr M, Zweckstetter M, Kugler S. Aggregation of alphaSynuclein promotes progressive in vivo neurotoxicity in adult rat dopaminergic neurons. *Acta Neuropathol.* 2012;123(5):671-83.
247. Galvin JE, Lee VM, Trojanowski JQ. Synucleinopathies: clinical and pathological implications. *Arch Neurol.* 2001;58(2):186-90.
248. Meyer-Luehmann M, Coomaraswamy J, Bolmont T, Kaeser S, Schaefer C, Kilger E, et al. Exogenous induction of cerebral beta-amyloidogenesis is governed by agent and host. *Science.* 2006;313(5794):1781-4.
249. Clavaguera F, Bolmont T, Crowther RA, Abramowski D, Frank S, Probst A, et al. Transmission and spreading of tauopathy in transgenic mouse brain. *Nat Cell Biol.* 2009;11(7):909-13.
250. Aguzzi A, Rajendran L. The transcellular spread of cytosolic amyloids, prions, and prionoids. *Neuron.* 2009;64(6):783-90.

251. Decressac M, Mattsson B, Lundblad M, Weikop P, Björklund A. Progressive neurodegenerative and behavioural changes induced by AAV-mediated overexpression of α -synuclein in midbrain dopamine neurons. *Neurobiology of Disease*. 2012;45(3):939-53.
252. Koprach JB, Johnston TH, Huot P, Reyes MG, Espinosa M, Brotchie JM. Progressive neurodegeneration or endogenous compensation in an animal model of Parkinson's disease produced by decreasing doses of alpha-synuclein. *PLoS One*. 2011;6(3):e17698.
253. Kirik D, Annett LE, Burger C, Muzyczka N, Mandel RJ, Björklund A. Nigrostriatal alpha-synucleinopathy induced by viral vector-mediated overexpression of human alpha-synuclein: a new primate model of Parkinson's disease. *Proc Natl Acad Sci U S A*. 2003;100(5):2884-9.
254. Koprach JB, Johnston TH, Reyes G, Omana V, Brotchie JM. Towards a Non-Human Primate Model of Alpha-Synucleinopathy for Development of Therapeutics for Parkinson's Disease: Optimization of AAV1/2 Delivery Parameters to Drive Sustained Expression of Alpha Synuclein and Dopaminergic Degeneration in Macaque. *PLOS ONE*. 2016;11(11):e0167235.
255. Koprach JB, Kalia LV, Brotchie JM. Animal models of α -synucleinopathy for Parkinson disease drug development. *Nat Rev Neurosci*. 2017;18:515.
256. Gray MT, Munoz DG, Gray DA, Schlossmacher MG, Woulfe JM. Alpha-synuclein in the appendiceal mucosa of neurologically intact subjects. *Mov Disord*. 2014;29(8):991-8.
257. Visanji NP, Marras C, Kern DS, Al Dakheel A, Gao A, Liu LW, et al. Colonic mucosal α -synuclein lacks specificity as a biomarker for Parkinson disease. *Neurology*. 2015;84(6):609-16.
258. Antunes L, Frasilho S, Ostaszewski M, Weber J, Longhino L, Antony P, et al. Similar alpha-Synuclein staining in the colon mucosa in patients with Parkinson's disease and controls. *Mov Disord*. 2016;31(10):1567-70.
259. Dias V, Junn E, Mouradian MM. The role of oxidative stress in Parkinson's disease. *J Parkinsons Dis*. 2013;3(4):461-91.
260. Zhu J, Chu CT. Mitochondrial dysfunction in Parkinson's disease. *J Alzheimers Dis*. 2010;20 Suppl 2:S325-34.
261. Jenner P, Olanow CW. The pathogenesis of cell death in Parkinson's disease. *Neurology*. 2006;66(10 Suppl 4):S24-36.
262. Beal MF. Mitochondria take center stage in aging and neurodegeneration. *Ann Neurol*. 2005;58(4):495-505.
263. Hastings TG. The role of dopamine oxidation in mitochondrial dysfunction: implications for Parkinson's disease. *J Bioenerg Biomembr*. 2009;41(6):469-72.
264. Gotz ME, Double K, Gerlach M, Youdim MB, Riederer P. The relevance of iron in the pathogenesis of Parkinson's disease. *Annals of the New York Academy of Sciences*. 2004;1012:193-208.
265. Dryanovski DI, Guzman JN, Xie Z, Galteri DJ, Volpicelli-Daley LA, Lee VM, et al. Calcium entry and alpha-synuclein inclusions elevate dendritic mitochondrial oxidant stress in dopaminergic neurons. *J Neurosci*. 2013;33(24):10154-64.
266. Wallace DC. Mitochondrial DNA mutations in disease and aging. *Environ Mol Mutagen*. 2010;51(5):440-50.
267. Jenner P. Oxidative stress in Parkinson's disease. *Ann Neurol*. 2003;53 Suppl 3:S26-36; discussion S-8.
268. Yoritaka A, Hattori N, Uchida K, Tanaka M, Stadtman ER, Mizuno Y. Immunohistochemical detection of 4-hydroxynonenal protein adducts in Parkinson disease.

- Proceedings of the National Academy of Sciences of the United States of America. 1996;93(7):2696-701.
269. Floor E, Wetzel MG. Increased protein oxidation in human substantia nigra pars compacta in comparison with basal ganglia and prefrontal cortex measured with an improved dinitrophenylhydrazine assay. *J Neurochem*. 1998;70(1):268-75.
270. Alam ZI, Jenner A, Daniel SE, Lees AJ, Cairns N, Marsden CD, et al. Oxidative DNA damage in the parkinsonian brain: an apparent selective increase in 8-hydroxyguanine levels in substantia nigra. *J Neurochem*. 1997;69(3):1196-203.
271. Zhang J, Perry G, Smith MA, Robertson D, Olson SJ, Graham DG, et al. Parkinson's disease is associated with oxidative damage to cytoplasmic DNA and RNA in substantia nigra neurons. *Am J Pathol*. 1999;154(5):1423-9.
272. Bové J, Prou D, Perier C, Przedborski S. Toxin-Induced Models of Parkinson's Disease. *NeuroRx*. 2005;2(3):484-94.
273. Turrens JF, Boveris A. Generation of superoxide anion by the NADH dehydrogenase of bovine heart mitochondria. *Biochem J*. 1980;191(2):421-7.
274. Barja G, Herrero A. Localization at complex I and mechanism of the higher free radical production of brain nonsynaptic mitochondria in the short-lived rat than in the longevous pigeon. *J Bioenerg Biomembr*. 1998;30(3):235-43.
275. Turrens JF. Mitochondrial formation of reactive oxygen species. *J Physiol*. 2003;552(Pt 2):335-44.
276. Schapira AH, Cooper JM, Dexter D, Clark JB, Jenner P, Marsden CD. Mitochondrial complex I deficiency in Parkinson's disease. *J Neurochem*. 1990;54(3):823-7.
277. Shults CW, Haas RH, Passov D, Beal MF. Coenzyme Q10 levels correlate with the activities of complexes I and II/III in mitochondria from parkinsonian and nonparkinsonian subjects. *Ann Neurol*. 1997;42(2):261-4.
278. Keeney PM, Xie J, Capaldi RA, Bennett JP, Jr. Parkinson's disease brain mitochondrial complex I has oxidatively damaged subunits and is functionally impaired and misassembled. *J Neurosci*. 2006;26(19):5256-64.
279. Elstner M, Morris CM, Heim K, Bender A, Mehta D, Jaros E, et al. Expression analysis of dopaminergic neurons in Parkinson's disease and aging links transcriptional dysregulation of energy metabolism to cell death. *Acta Neuropathol*. 2011;122(1):75-86.
280. Schneider L. Chapter 1 - Anatomy and Physiology of Normal Sleep A2 - Miglis, Mitchell G. *Sleep and Neurologic Disease*. San Diego: Academic Press; 2017. p. 1-28.
281. Anden NE, Carlsson A, Dahlstrom A, Fuxe K, Hillarp NA, Larsson K. DEMONSTRATION AND MAPPING OUT OF NIGRO-NEOSTRIATAL DOPAMINE NEURONS. *Life sciences (1962)*. 1964;3:523-30.
282. Dahlstrom A, Fuxe K. Localization of monoamines in the lower brain stem. *Experientia*. 1964;20(7):398-9.
283. Dauer W, Przedborski S. Parkinson's disease: mechanisms and models. *Neuron*. 2003;39(6):889-909.
284. Hudzik TJ, Markgraf CG. Chapter 1 - Nonclinical Assessment of Abuse Potential for New Pharmaceuticals in a Regulatory Space. *Nonclinical Assessment of Abuse Potential for New Pharmaceuticals*. Boston: Academic Press; 2015. p. 1-7.
285. Hudepohl NS, Nasrallah HA. Chapter 39 - Antipsychotic drugs. In: Aminoff MJ, Boller F, Swaab DF, editors. *Handbook of Clinical Neurology*. 106: Elsevier; 2012. p. 657-67.

286. Beninger RJ. The role of dopamine in locomotor activity and learning. *Brain Res.* 1983;287(2):173-96.
287. Meiser J, Weindl D, Hiller K. Complexity of dopamine metabolism. *Cell Commun Signal.* 2013;11(1):34.
288. Fuxe K, Goldstein M, Hokfelt T, Joh TH. Immunohistochemical localization of dopamine-*-*hydroxylase in the peripheral and central nervous system. *Res Commun Chem Pathol Pharmacol.* 1970;1(5):627-36.
289. Beaulieu JM, Gainetdinov RR. The physiology, signaling, and pharmacology of dopamine receptors. *Pharmacol Rev.* 2011;63(1):182-217.
290. Beaulieu JM, Gainetdinov RR, Caron MG. The Akt-GSK-3 signaling cascade in the actions of dopamine. *Trends in pharmacological sciences.* 2007;28(4):166-72.
291. Civelli O, Bunzow JR, Grandy DK. Molecular diversity of the dopamine receptors. *Annu Rev Pharmacol Toxicol.* 1993;33:281-307.
292. Gingrich JA, Caron MG. Recent advances in the molecular biology of dopamine receptors. *Annu Rev Neurosci.* 1993;16:299-321.
293. Keabian JW, Calne DB. Multiple receptors for dopamine. *Nature.* 1979;277(5692):93-6.
294. Stoof JC, Keabian JW. Opposing roles for D-1 and D-2 dopamine receptors in efflux of cyclic AMP from rat neostriatum. *Nature.* 1981;294(5839):366-8.
295. Mercuri NB, Saiardi A, Bonci A, Picetti R, Calabresi P, Bernardi G, et al. Loss of autoreceptor function in dopaminergic neurons from dopamine D2 receptor deficient mice. *Neuroscience.* 1997;79(2):323-7.
296. Hurd YL, Suzuki M, Sedvall GC. D1 and D2 dopamine receptor mRNA expression in whole hemisphere sections of the human brain. *J Chem Neuroanat.* 2001;22(1-2):127-37.
297. Sokoloff P, Schwartz JC. Novel dopamine receptors half a decade later. *Trends in pharmacological sciences.* 1995;16(8):270-5.
298. Meador-Woodruff JH, Damask SP, Wang J, Haroutunian V, Davis KL, Watson SJ. Dopamine receptor mRNA expression in human striatum and neocortex. *Neuropsychopharmacology.* 1996;15(1):17-29.
299. DeLong MR, Wichmann T. Circuits and circuit disorders of the basal ganglia. *Arch Neurol.* 2007;64(1):20-4.
300. Aubert I, Ghorayeb I, Normand E, Bloch B. Phenotypical characterization of the neurons expressing the D1 and D2 dopamine receptors in the monkey striatum. *The Journal of comparative neurology.* 2000;418(1):22-32.
301. Koprach JB, Johnston TH, Huot P, Fox SH, Brotchie JM. New insights into the organization of the basal ganglia. *Curr Neurol Neurosci Rep.* 2009;9(4):298-304.
302. Cepeda C, Hurst RS, Altemus KL, Flores-Hernandez J, Calvert CR, Jokel ES, et al. Facilitated glutamatergic transmission in the striatum of D2 dopamine receptor-deficient mice. *J Neurophysiol.* 2001;85(2):659-70.
303. Wang H, Pickel VM. Dopamine D2 receptors are present in prefrontal cortical afferents and their targets in patches of the rat caudate-putamen nucleus. *The Journal of comparative neurology.* 2002;442(4):392-404.
304. Bezard E, Ferry S, Mach U, Stark H, Leriche L, Boraud T, et al. Attenuation of levodopa-induced dyskinesia by normalizing dopamine D3 receptor function. *Nat Med.* 2003;9(6):762-7.

305. Murray AM, Ryoo HL, Gurevich E, Joyce JN. Localization of dopamine D3 receptors to mesolimbic and D2 receptors to mesostriatal regions of human forebrain. *Proceedings of the National Academy of Sciences of the United States of America*. 1994;91(23):11271-5.
306. Suzuki M, Hurd YL, Sokoloff P, Schwartz JC, Sedvall G. D3 dopamine receptor mRNA is widely expressed in the human brain. *Brain Res*. 1998;779(1-2):58-74.
307. Gurevich EV, Himes JW, Joyce JN. Developmental regulation of expression of the D3 dopamine receptor in rat nucleus accumbens and islands of Calleja. *J Pharmacol Exp Ther*. 1999;289(1):587-98.
308. Cenci MA. Presynaptic Mechanisms of 1-DOPA-Induced Dyskinesia: The Findings, the Debate, and the Therapeutic Implications. *Front Neurol*. 2014;5:242.
309. Postuma RB, Berg D. Advances in markers of prodromal Parkinson disease. *Nat Rev Neurol*. 2016;12(11):622-34.
310. Kakkar AK, Dahiya N. Management of Parkinsons disease: Current and future pharmacotherapy. *Eur J Pharmacol*. 2015;750:74-81.
311. Marras C, Lang A. Parkinson's disease subtypes: lost in translation? *Journal of Neurology, Neurosurgery & Psychiatry*. 2013;84(4):409.
312. Berardelli A, Rothwell JC, Thompson PD, Hallett M. Pathophysiology of bradykinesia in Parkinson's disease. *Brain*. 2001;124(Pt 11):2131-46.
313. Evarts EV, Teravainen H, Calne DB. Reaction time in Parkinson's disease. *Brain*. 1981;104(Pt 1):167-86.
314. Jahanshahi M, Brown RG, Marsden CD. Simple and choice reaction time and the use of advance information for motor preparation in Parkinson's disease. *Brain*. 1992;115 (Pt 2):539-64.
315. Bloem BR, Hausdorff JM, Visser JE, Giladi N. Falls and freezing of gait in Parkinson's disease: a review of two interconnected, episodic phenomena. *Mov Disord*. 2004;19(8):871-84.
316. Michalowska M, Fiszer U, Krygowska-Wajs A, Owczarek K. Falls in Parkinson's disease. Causes and impact on patients' quality of life. *Functional neurology*. 2005;20(4):163-8.
317. Morris ME. Locomotor training in people with Parkinson disease. *Physical therapy*. 2006;86(10):1426-35.
318. Wielinski CL, Erickson-Davis C, Wichmann R, Walde-Douglas M, Parashos SA. Falls and injuries resulting from falls among patients with Parkinson's disease and other parkinsonian syndromes. *Mov Disord*. 2005;20(4):410-5.
319. Hallett M. Clinical neurophysiology of akinesia. *Revue neurologique*. 1990;146(10):585-90.
320. Fahn S. Description of Parkinson's disease as a clinical syndrome. *Annals of the New York Academy of Sciences*. 2003;991:1-14.
321. Mazzoni P, Shabbott B, Cortés JC. Motor Control Abnormalities in Parkinson's Disease. *Cold Spring Harbor Perspectives in Medicine*. 2012;2(6):a009282.
322. Martinez-Martin P, Rodriguez-Blazquez C, Kurtis MM, Chaudhuri KR, Group NV. The impact of non-motor symptoms on health-related quality of life of patients with Parkinson's disease. *Mov Disord*. 2011;26(3):399-406.
323. Chaudhuri KR, Odin P. The challenge of non-motor symptoms in Parkinson's disease. *Prog Brain Res*. 2010;184:325-41.
324. Trenkwalder C, Kies B, Rudzinska M, Fine J, Nikl J, Honczarenko K, et al. Rotigotine effects on early morning motor function and sleep in Parkinson's disease: a double-blind, randomized, placebo-controlled study (RECOVER). *Mov Disord*. 2011;26(1):90-9.

325. Honig H, Antonini A, Martinez-Martin P, Forgacs I, Faye GC, Fox T, et al. Intrajejunal levodopa infusion in Parkinson's disease: a pilot multicenter study of effects on nonmotor symptoms and quality of life. *Mov Disord.* 2009;24(10):1468-74.
326. Chaudhuri KR, Martinez-Martin P, Odin P, Antonini A. *Handbook of Non-Motor Symptoms in Parkinson's Disease*: Springer Healthcare Limited; 2012.
327. Barone P, Antonini A, Colosimo C, Marconi R, Morgante L, Avarello TP, et al. The PRIAMO study: A multicenter assessment of nonmotor symptoms and their impact on quality of life in Parkinson's disease. *Mov Disord.* 2009;24(11):1641-9.
328. Guo X, Song W, Chen K, Chen X, Zheng Z, Cao B, et al. Gender and onset age-related features of non-motor symptoms of patients with Parkinson's disease--a study from Southwest China. *Parkinsonism Relat Disord.* 2013;19(11):961-5.
329. Spica V, Pekmezovic T, Svetel M, Kostic VS. Prevalence of non-motor symptoms in young-onset versus late-onset Parkinson's disease. *J Neurol.* 2013;260(1):131-7.
330. Gallagher DA, Lees AJ, Schrag A. What are the most important nonmotor symptoms in patients with Parkinson's disease and are we missing them? *Mov Disord.* 2010;25(15):2493-500.
331. Schrag A, Jahanshahi M, Quinn N. What contributes to quality of life in patients with Parkinson's disease? *J Neurol Neurosurg Psychiatry.* 2000;69(3):308-12.
332. Chaudhuri KR, Martinez-Martin P, Brown RG, Sethi K, Stocchi F, Odin P, et al. The metric properties of a novel non-motor symptoms scale for Parkinson's disease: Results from an international pilot study. *Mov Disord.* 2007;22(13):1901-11.
333. Doty RL. Olfactory dysfunction in Parkinson disease. *Nature reviews Neurology.* 2012;8(6):329-39.
334. Berg D, Marek K, Ross GW, Poewe W. Defining at-risk populations for Parkinson's disease: lessons from ongoing studies. *Mov Disord.* 2012;27(5):656-65.
335. Haehner A, Hummel T, Hummel C, Sommer U, Junghanns S, Reichmann H. Olfactory loss may be a first sign of idiopathic Parkinson's disease. *Mov Disord.* 2007;22(6):839-42.
336. Ross GW, Petrovitch H, Abbott RD, Tanner CM, Popper J, Masaki K, et al. Association of olfactory dysfunction with risk for future Parkinson's disease. *Ann Neurol.* 2008;63(2):167-73.
337. Ponsen MM, Stoffers D, Twisk JW, Wolters E, Berendse HW. Hyposmia and executive dysfunction as predictors of future Parkinson's disease: a prospective study. *Mov Disord.* 2009;24(7):1060-5.
338. Postuma RB, Gagnon JF, Vendette M, Desjardins C, Montplaisir JY. Olfaction and color vision identify impending neurodegeneration in rapid eye movement sleep behavior disorder. *Ann Neurol.* 2011;69(5):811-8.
339. Mahlknecht P, Iranzo A, Hogg B, Frauscher B, Muller C, Santamaria J, et al. Olfactory dysfunction predicts early transition to a Lewy body disease in idiopathic RBD. *Neurology.* 2015;84(7):654-8.
340. Jennings D, Stern M, Siderowf A, Marek K. Longitudinal imaging and phenoconversion in the PARS prodromal cohort. *Neurodegener Dis.* 2015;15(Suppl. 1):242.
341. Schenck CH, Montplaisir JY, Frauscher B, Hogg B, Gagnon JF, Postuma R, et al. Rapid eye movement sleep behavior disorder: devising controlled active treatment studies for symptomatic and neuroprotective therapy--a consensus statement from the International Rapid Eye Movement Sleep Behavior Disorder Study Group. *Sleep Med.* 2013;14(8):795-806.

342. Postuma RB, Gagnon JF, Bertrand JA, Genier Marchand D, Montplaisir JY. Parkinson risk in idiopathic REM sleep behavior disorder: preparing for neuroprotective trials. *Neurology*. 2015;84(11):1104-13.
343. Postuma RB, Iranzo A, Hogl B, Arnulf I, Ferini-Strambi L, Manni R, et al. Risk factors for neurodegeneration in idiopathic rapid eye movement sleep behavior disorder: a multicenter study. *Ann Neurol*. 2015;77(5):830-9.
344. Schenck CH, Boeve BF, Mahowald MW. Delayed emergence of a parkinsonian disorder or dementia in 81% of older men initially diagnosed with idiopathic rapid eye movement sleep behavior disorder: a 16-year update on a previously reported series. *Sleep Med*. 2013;14(8):744-8.
345. Iranzo A, Fernandez-Arcos A, Tolosa E, Serradell M, Molinuevo JL, Valldeoriola F, et al. Neurodegenerative disorder risk in idiopathic REM sleep behavior disorder: study in 174 patients. *PLoS One*. 2014;9(2):e89741.
346. Wing YK, Li SX, Mok V, Lam SP, Tsoh J, Chan A, et al. Prospective outcome of rapid eye movement sleep behaviour disorder: psychiatric disorders as a potential early marker of Parkinson's disease. *J Neurol Neurosurg Psychiatry*. 2012;83(4):470-2.
347. Arnulf I, Neutel D, Herlin B, Golmard JL, Leu-Semenescu S, Cochen de Cock V, et al. Sleepiness in Idiopathic REM Sleep Behavior Disorder and Parkinson Disease. *Sleep*. 2015;38(10):1529-35.
348. Boeve BF, Silber MH, Ferman TJ, Lin SC, Benarroch EE, Schmeichel AM, et al. Clinicopathologic correlations in 172 cases of rapid eye movement sleep behavior disorder with or without a coexisting neurologic disorder. *Sleep Med*. 2013;14(8):754-62.
349. Luk KC, Kehm V, Carroll J, Zhang B, O'Brien P, Trojanowski JQ, et al. Pathological alpha-synuclein transmission initiates Parkinson-like neurodegeneration in nontransgenic mice. *Science*. 2012;338(6109):949-53.
350. Svensson E, Horvath-Puho E, Thomsen RW, Djurhuus JC, Pedersen L, Borghammer P, et al. Vagotomy and subsequent risk of Parkinson's disease. *Ann Neurol*. 2015;78(4):522-9.
351. Schrag A, Horsfall L, Walters K, Noyce A, Petersen I. Prediagnostic presentations of Parkinson's disease in primary care: a case-control study. *The Lancet Neurology*. 2015;14(1):57-64.
352. Leentjens AF, Van den Akker M, Metsemakers JF, Lousberg R, Verhey FR. Higher incidence of depression preceding the onset of Parkinson's disease: a register study. *Mov Disord*. 2003;18(4):414-8.
353. Leentjens AF, Driessen G, Weber W, Drukker M, van Os J. Mental health care use in Parkinson's disease: a record linkage study. *Neuroepidemiology*. 2008;30(2):71-5.
354. Fang F, Xu Q, Park Y, Huang X, Hollenbeck A, Blair A, et al. Depression and the subsequent risk of Parkinson's disease in the NIH-AARP Diet and Health Study. *Mov Disord*. 2010;25(9):1157-62.
355. Gustafsson H, Nordstrom A, Nordstrom P. Depression and subsequent risk of Parkinson disease: A nationwide cohort study. *Neurology*. 2015;84(24):2422-9.
356. Alonso A, Rodriguez LA, Logroscino G, Hernan MA. Use of antidepressants and the risk of Parkinson's disease: a prospective study. *J Neurol Neurosurg Psychiatry*. 2009;80(6):671-4.
357. Poletti M, Frosini D, Ceravolo R, Bonuccelli U. Mild cognitive impairment in De Novo Parkinson's disease according to movement disorder guidelines. *Mov Disord*. 2012;27(13):1706; author reply 7.

358. Schapira AHV, Chaudhuri KR, Jenner P. Non-motor features of Parkinson disease. *2017*;18:435.
359. AlDakheel A, Kalia LV, Lang AE. Pathogenesis-targeted, disease-modifying therapies in Parkinson disease. *Neurotherapeutics*. 2014;11(1):6-23.
360. Jankovic J. Levodopa strengths and weaknesses. *Neurology*. 2002;58(4 Suppl 1):S19-32.
361. Jankovic J, Aguilar LG. Current approaches to the treatment of Parkinson's disease. *Neuropsychiatric Disease and Treatment*. 2008;4(4):743-57.
362. Fahn S. The history of dopamine and levodopa in the treatment of Parkinson's disease. *Mov Disord*. 2008;23 Suppl 3:S497-508.
363. Funk C. LXV.-Synthesis of dl-3 : 4-dihydroxyphenylalanine. *Journal of the Chemical Society, Transactions*. 1911;99(0):554-7.
364. Ehringer H, Hornykiewicz O. [Distribution of noradrenaline and dopamine (3-hydroxytyramine) in the human brain and their behavior in diseases of the extrapyramidal system]. *Klin Wochenschr*. 1960;38:1236-9.
365. Hornykiewicz O. Dopamine (3-hydroxytyramine) and brain function. *Pharmacol Rev*. 1966;18(2):925-64.
366. Hornykiewicz O. L-DOPA: from a biologically inactive amino acid to a successful therapeutic agent. *Amino Acids*. 2002;23(1-3):65-70.
367. Deleu D, Northway MG, Hanssens Y. Clinical pharmacokinetic and pharmacodynamic properties of drugs used in the treatment of Parkinson's disease. *Clin Pharmacokinet*. 2002;41(4):261-309.
368. Barbeau A, Sourkes TL, GF M. Les catécholamines dans la maladie de Parkinson. In: J dA, editor. *Monoamines et système nerveux central*. Geneva: Georg & Cie SA; 1962. p. 247-62.
369. Cotzias GC. Parkinsonism and Dopa. *J Chronic Dis*. 1969;22(5):297-301.
370. Cotzias GC, Papavasiliou PS, Gellene R. L-dopa in parkinson's syndrome. *N Engl J Med*. 1969;281(5):272.
371. Cotzias GC, Papavasiliou PS, Gellene R. Modification of Parkinsonism--chronic treatment with L-dopa. *N Engl J Med*. 1969;280(7):337-45.
372. Barbeau A, Mars H, Botez MI, Joubert M. Levodopa combined with peripheral decarboxylase inhibition in Parkinson's disease. *Canadian Medical Association Journal*. 1972;106(11):1169-74.
373. Sweet RD, McDowell FH, Wasterlain CG, Stern PH. Treatment of "on-off effect" with a dopa decarboxylase inhibitor. *Arch Neurol*. 1975;32(8):560-3.
374. Nutt JG, Woodward WR, Anderson JL. The effect of carbidopa on the pharmacokinetics of intravenously administered levodopa: the mechanism of action in the treatment of parkinsonism. *Ann Neurol*. 1985;18(5):537-43.
375. Hauser RA. Levodopa: past, present, and future. *European neurology*. 2009;62(1):1-8.
376. Editorial: Dopa decarboxylase inhibitors. *British Medical Journal*. 1974;4(5939):250-1.
377. LeWitt PA. Levodopa therapeutics: new treatment strategies. *Neurology*. 1993;43(12 Suppl 6):S31-7.
378. Wade DN, Mearrick PT, Morris JL. Active Transport of L-Dopa in the Intestine. *Nature*. 1973;242(5398):463-5.

379. Kageyama T, Nakamura M, Matsuo A, Yamasaki Y, Takakura Y, Hashida M, et al. The 4F2hc/LAT1 complex transports L-DOPA across the blood-brain barrier. *Brain Res.* 2000;879(1-2):115-21.
380. Pardridge WM. Transport of small molecules through the blood-brain barrier: biology and methodology. *Advanced Drug Delivery Reviews.* 1995;15(1):5-36.
381. Mena I, Cotzias GC. Protein Intake and Treatment of Parkinson's Disease with Levodopa. *New England Journal of Medicine.* 1975;292(4):181-4.
382. Frankel JP, Kempster PA, Bovingdon M, Webster R, Lees AJ, Stern GM. The effects of oral protein on the absorption of intraduodenal levodopa and motor performance. *J Neurol Neurosurg Psychiatry.* 1989;52(9):1063-7.
383. Cedarbaum JM. Clinical pharmacokinetics of anti-parkinsonian drugs. *Clin Pharmacokinet.* 1987;13(3):141-78.
384. Gancher ST, Nutt JG, Woodward WR. Peripheral pharmacokinetics of levodopa in untreated, stable, and fluctuating parkinsonian patients. *Neurology.* 1987;37(6):940-4.
385. Hauser RA, Ellenbogen AL, Metman LV, Hsu A, O'Connell MJ, Modi NB, et al. Crossover comparison of IPX066 and a standard levodopa formulation in advanced Parkinson's disease. *Mov Disord.* 2011;26(12):2246-52.
386. Nagai M, Kubo M, Nishikawa N, Nomoto M. Fluctuation in plasma entacapone concentrations in accordance with variable plasma levodopa concentrations. *Parkinsonism Relat Disord.* 2010;16(10):697-9.
387. Nutt JG, Woodward WR, Beckner RM, Stone CK, Berggren K, Carter JH, et al. Effect of peripheral catechol-O-methyltransferase inhibition on the pharmacokinetics and pharmacodynamics of levodopa in parkinsonian patients. *Neurology.* 1994;44(5):913-9.
388. Tohgi H, Abe T, Yamazaki K, Saheki M, Takahashi S, Tsukamoto Y. Effects of the catechol-O-methyltransferase inhibitor tolcapone in Parkinson's disease: correlations between concentrations of dopaminergic substances in the plasma and cerebrospinal fluid and clinical improvement. *Neuroscience Letters.* 1995;192(3):165-8.
389. Olanow CW, Gauger LL, Cedarbaum JM. Temporal relationships between plasma and cerebrospinal fluid pharmacokinetics of levodopa and clinical effect in Parkinson's disease. *Ann Neurol.* 1991;29(5):556-9.
390. Woodward WR, Olanow CW, Beckner RM, Hauser RA, Gauger LL, Cedarbaum JM, et al. The effect of L-dopa infusions with and without phenylalanine challenges in parkinsonian patients: Plasma and ventricular CSF L-dopa levels and clinical responses. *Neurology.* 1993;43(9):1704.
391. Napolitano A, Del Dotto P, Petrozzi L, Dell'Agnello G, Bellini G, Gambaccini G, et al. Pharmacokinetics and pharmacodynamics of L-Dopa after acute and 6-week tolcapone administration in patients with Parkinson's disease. *Clinical neuropharmacology.* 1999;22(1):24-9.
392. Scott LJ. Opicapone: A Review in Parkinson's Disease. *Drugs.* 2016;76(13):1293-300.
393. Beltramo M, Krieger M, Calas A, Franzoni MF, Thibault J. Aromatic amino acid decarboxylase (AADC) immunohistochemistry in vertebrate brainstem with an antiserum raised against AADC made in *E. coli*. *Brain Res Bull.* 1993;32(2):123-32.
394. Arai R, Karasawa N, Nagatsu I. Aromatic L-amino acid decarboxylase is present in serotonergic fibers of the striatum of the rat. A double-labeling immunofluorescence study. *Brain Res.* 1996;706(1):177-9.

395. Karasawa N, Hayashi M, Yamada K, Nagatsu I, Iwasa M, Takeuchi T, et al. Tyrosine Hydroxylase (TH)- and Aromatic-L-Amino Acid Decarboxylase (AADC)-Immunoreactive Neurons of the Common Marmoset (*Callithrix jacchus*) Brain: An Immunohistochemical Analysis. *Acta Histochemica et Cytochemica*. 2007;40(3):83-92.
396. Weihe E, Depboylu C, Schütz B, Schäfer MKH, Eiden LE. Three Types of Tyrosine Hydroxylase-Positive CNS Neurons Distinguished by Dopa Decarboxylase and VMAT2 Co-Expression. *Cellular and molecular neurobiology*. 2006;26(0):659-78.
397. Meredith GE, Farrell T, Kellaghan P, Tan Y, Zahm DS, Totterdell S. Immunocytochemical characterization of catecholaminergic neurons in the rat striatum following dopamine-depleting lesions. *Eur J Neurosci*. 1999;11(10):3585-96.
398. Huot P, Parent A. Dopaminergic neurons intrinsic to the striatum. *J Neurochem*. 2007;101(6):1441-7.
399. Huot P, Johnston TH, Koprach JB, Fox SH, Brotchie JM. The pharmacology of L-DOPA-induced dyskinesia in Parkinson's disease. *Pharmacol Rev*. 2013;65(1):171-222.
400. Jankovic J. Motor fluctuations and dyskinesias in Parkinson's disease: clinical manifestations. *Mov Disord*. 2005;20 Suppl 11:S11-6.
401. Marras C, Lang AE. Measuring motor complications in clinical trials for early Parkinson's disease. *J Neurol Neurosurg Psychiatry*. 2003;74(2):143-6.
402. Tan EK. Dopamine agonists and their role in Parkinson's disease treatment. *Expert Rev Neurother*. 2003;3(6):805-10.
403. Fiszer U. [Adverse effects of dopamine agonists]. *Neurol Neurochir Pol*. 2007;41(2 Suppl 1):S34-9.
404. Broussolle E, Cinotti L, Pollak P, Landais P, Le Bars D, Galy G, et al. Relief of akinesia by apomorphine and cerebral metabolic changes in Parkinson's disease. *Mov Disord*. 1993;8(4):459-62.
405. Krishna R, Ali M, Moustafa AA. Effects of combined MAO-B inhibitors and levodopa vs. monotherapy in Parkinson's disease. *Frontiers in Aging Neuroscience*. 2014;6:180.
406. Lyytinen J, Kaakkola S, Ahtila S, Tuomainen P, Teravainen H. Simultaneous MAO-B and COMT inhibition in L-Dopa-treated patients with Parkinson's disease. *Mov Disord*. 1997;12(4):497-505.
407. Assal F, Spahr L, Hadengue A, Rubbia-Brandt L, Burkhard PR. Tolcapone and fulminant hepatitis. *Lancet*. 1998;352(9132):958.
408. Olanow CW. Tolcapone and hepatotoxic effects. Tasmar Advisory Panel. *Arch Neurol*. 2000;57(2):263-7.
409. Schrag A. Entacapone in the treatment of Parkinson's disease. *The Lancet Neurology*. 2005;4(6):366-70.
410. Pahwa R, Factor SA, Lyons KE, Ondo WG, Gronseth G, Bronte-Stewart H, et al. Practice Parameter: Treatment of Parkinson disease with motor fluctuations and dyskinesia (an evidence-based review): Report of the Quality Standards Subcommittee of the American Academy of Neurology. *Neurology*. 2006;66(7):983-95.
411. Hickey P, Stacy M. Deep Brain Stimulation: A Paradigm Shifting Approach to Treat Parkinson's Disease. *Front Neurosci*. 2016;10:173.
412. Martinez-Ramirez D, Hu W, Bona AR, Okun MS, Wagle Shukla A. Update on deep brain stimulation in Parkinson's disease. *Transl Neurodegener*. 2015;4:12.
413. Kalia SK, Sankar T, Lozano AM. Deep brain stimulation for Parkinson's disease and other movement disorders. *Curr Opin Neurol*. 2013;26(4):374-80.

414. Deuschl G, Schupbach M, Knudsen K, Pinski MO, Cornu P, Rau J, et al. Stimulation of the subthalamic nucleus at an earlier disease stage of Parkinson's disease: concept and standards of the EARLYSTIM-study. *Parkinsonism Relat Disord*. 2013;19(1):56-61.
415. Okun MS, Fernandez HH, Wu SS, Kirsch-Darrow L, Bowers D, Bova F, et al. Cognition and mood in Parkinson's disease in subthalamic nucleus versus globus pallidus interna deep brain stimulation: the COMPARE trial. *Ann Neurol*. 2009;65(5):586-95.
416. Weaver FM, Follett K, Stern M, Hur K, Harris C, Marks WJ, et al. Bilateral Deep Brain Stimulation vs Best Medical Therapy for Patients With Advanced Parkinson Disease: A Randomized Controlled Trial. *JAMA : the journal of the American Medical Association*. 2009;301(1):63.
417. Williams A, Gill S, Varma T, Jenkinson C, Quinn N, Mitchell R, et al. Deep brain stimulation plus best medical therapy versus best medical therapy alone for advanced Parkinson's disease (PD SURG trial): a randomised, open-label trial. *Lancet Neurology*. 2010;9(6):581-91.
418. Limousin P, Pollak P, Benazzouz A, Hoffmann D, Broussolle E, Perret JE, et al. Bilateral subthalamic nucleus stimulation for severe Parkinson's disease. *Mov Disord*. 1995;10(5):672-4.
419. Benazzouz A, Gao DM, Ni ZG, Piallat B, Bouali-Benazzouz R, Benabid AL. Effect of high-frequency stimulation of the subthalamic nucleus on the neuronal activities of the substantia nigra pars reticulata and ventrolateral nucleus of the thalamus in the rat. *Neuroscience*. 2000;99(2):289-95.
420. Zis P, Chaudhuri KR, Samuel M. Phenomenology of Levodopa-Induced Dyskinesia. In: Fox SH, Brotchie JM, editors. *Levodopa-Induced Dyskinesia in Parkinson's Disease*. London: Springer London; 2014. p. 1-16.
421. Poewe WH, Lees AJ, Stern GM. Dystonia in Parkinson's disease: clinical and pharmacological features. *Ann Neurol*. 1988;23(1):73-8.
422. Marconi R, Lefebvre-Caparros D, Bonnet AM, Vidailhet M, Dubois B, Agid Y. Levodopa-induced dyskinesias in Parkinson's disease phenomenology and pathophysiology. *Mov Disord*. 1994;9(1):2-12.
423. Rascol O, Brooks DJ, Korczyn AD, De Deyn PP, Clarke CE, Lang AE. A five-year study of the incidence of dyskinesia in patients with early Parkinson's disease who were treated with ropinirole or levodopa. *N Engl J Med*. 2000;342(20):1484-91.
424. Constantinescu R, Romer M, McDermott MP, Kamp C, Kiebertz K. Impact of pramipexole on the onset of levodopa-related dyskinesias. *Mov Disord*. 2007;22(9):1317-9.
425. Martinez-Martin P, O'Brien CF. Extending levodopa action: COMT inhibition. *Neurology*. 1998;50(6 Suppl 6):S27-32; discussion S44-8.
426. Nutt JG. On-off phenomenon: relation to levodopa pharmacokinetics and pharmacodynamics. *Ann Neurol*. 1987;22(4):535-40.
427. Zesiewicz TA, Sullivan KL, Hauser RA. Levodopa-induced dyskinesia in Parkinson's disease: epidemiology, etiology, and treatment. *Curr Neurol Neurosci Rep*. 2007;7(4):302-10.
428. Fearnley JM, Lees AJ. Ageing and Parkinson's disease: substantia nigra regional selectivity. *Brain*. 1991;114 (Pt 5):2283-301.
429. Schapira AHV, Hartmann A, Agid Y. Parkinsonian disorders in clinical practice. 2009.
430. Guridi J, Gonz, #xe1, lez-Redondo R, Obeso JA. Clinical Features, Pathophysiology, and Treatment of Levodopa-Induced Dyskinesias in Parkinson's Disease. *Parkinson's Disease*. 2012;2012:15.

431. Hametner E, Seppi K, Poewe W. The clinical spectrum of levodopa-induced motor complications. *J Neurol*. 2010;257(Suppl 2):S268-75.
432. Melamed E. Early-morning dystonia. A late side effect of long-term levodopa therapy in Parkinson's disease. *Arch Neurol*. 1979;36(5):308-10.
433. Quinn NP. Classification of fluctuations in patients with Parkinson's disease. *Neurology*. 1998;51(2 Suppl 2):S25-9.
434. Lees AJ. A sustained-release formulation of L-dopa (Madopar HBS) in the treatment of nocturnal and early-morning disabilities in Parkinson's disease. *European neurology*. 1987;27 Suppl 1:126-34.
435. Muentner MD, Sharpless NS, Tyce GM, Darley FL. Patterns of dystonia ("I-D-I" and "D-I-D") in response to l-dopa therapy for Parkinson's disease. *Mayo Clinic proceedings*. 1977;52(3):163-74.
436. Alegre M, Lopez-Azcarate J, Alonso-Frech F, Rodriguez-Oroz MC, Valencia M, Guridi J, et al. Subthalamic activity during diphasic dyskinesias in Parkinson's disease. *Mov Disord*. 2012;27(9):1178-81.
437. Obeso JA, Grandas F, Vaamonde J, Luquin MR, Artieda J, Lera G, et al. Motor complications associated with chronic levodopa therapy in Parkinson's disease. *Neurology*. 1989;39(11 Suppl 2):11-9.
438. Luquin MR, Scipioni O, Vaamonde J, Gershanik O, Obeso JA. Levodopa-induced dyskinesias in Parkinson's disease: clinical and pharmacological classification. *Mov Disord*. 1992;7(2):117-24.
439. Jenner P. Molecular mechanisms of L-DOPA-induced dyskinesia. *Nat Rev Neurosci*. 2008;9(9):665-77.
440. Nadjar A, Gerfen CR, Bezard E. Priming for l-dopa-induced dyskinesia in Parkinson's disease: a feature inherent to the treatment or the disease? *Prog Neurobiol*. 2009;87(1):1-9.
441. Di Chiara G, Morelli M, Barone P, Pontieri F. Priming as a model of behavioural sensitization. *Developmental pharmacology and therapeutics*. 1992;18(3-4):223-7.
442. Nutt JG. Continuous dopaminergic stimulation: Is it the answer to the motor complications of Levodopa? *Mov Disord*. 2007;22(1):1-9.
443. Cenci MA. Chapter 43 - Molecular Mechanisms of l-DOPA-Induced Dyskinesia. In: Steiner H, Tseng KY, editors. *Handbook of Behavioral Neuroscience*. 24: Elsevier; 2017. p. 857-71.
444. Munhoz RP, Cerasa A, Okun MS. Surgical Treatment of Dyskinesia in Parkinson's Disease. *Frontiers in Neurology*. 2014;5(65).
445. Thanvi B, Lo N, Robinson T. Levodopa-induced dyskinesia in Parkinson's disease: clinical features, pathogenesis, prevention and treatment. *Postgrad Med J*. 2007;83(980):384-8.
446. Jenner P. Molecular mechanisms of L-DOPA-induced dyskinesia. *Nat Rev Neurosci*. 2008;9(9):665-77.
447. Jenner P, McCreary AC, Scheller DKA. Continuous drug delivery in early- and late-stage Parkinson's disease as a strategy for avoiding dyskinesia induction and expression. *Journal of Neural Transmission*. 2011;118(12):1691-702.
448. Muentner MD, Tyce GM. L-dopa therapy of Parkinson's disease: plasma L-dopa concentration, therapeutic response, and side effects. *Mayo Clinic proceedings*. 1971;46(4):231-9.
449. Nutt JG. Pharmacokinetics and pharmacodynamics of levodopa. *Mov Disord*. 2008;23 Suppl 3:S580-4.

450. Jenner P, McCreary AC, Scheller DK. Continuous drug delivery in early- and late-stage Parkinson's disease as a strategy for avoiding dyskinesia induction and expression. *J Neural Transm (Vienna)*. 2011;118(12):1691-702.
451. Kvernmo T, Härtter S, Burger E. A review of the receptor-binding and pharmacokinetic properties of dopamine agonists. *Clinical Therapeutics*. 2006;28(8):1065-78.
452. Olanow CW, Obeso JA, Stocchi F. Drug insight: Continuous dopaminergic stimulation in the treatment of Parkinson's disease. *Nat Clin Pract Neurol*. 2006;2(7):382-92.
453. Rascol O. Drugs and drug delivery in PD: optimizing control of symptoms with pramipexole prolonged-release. *Eur J Neurol*. 2011;18 Suppl 1:3-10.
454. De Keyser J, De Backer J-P, Wilczak N, Herroelen L. Dopamine agonists used in the treatment of Parkinson's disease and their selectivity for the D1, D2, and D3 dopamine receptors in human striatum. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*. 1995;19(7):1147-54.
455. Cassarino DS, Fall CP, Smith TS, Bennett JP, Jr. Pramipexole reduces reactive oxygen species production in vivo and in vitro and inhibits the mitochondrial permeability transition produced by the parkinsonian neurotoxin methylpyridinium ion. *J Neurochem*. 1998;71(1):295-301.
456. Trugman JM, James CL, Wooten GF. D1/D2 dopamine receptor stimulation by L-dopa. A [¹⁴C]-2-deoxyglucose autoradiographic study. *Brain*. 1991;114 (Pt 3):1429-40.
457. Taylor JL, Bishop C, Walker PD. Dopamine D1 and D2 receptor contributions to L-DOPA-induced dyskinesia in the dopamine-depleted rat. *Pharmacol Biochem Behav*. 2005;81(4):887-93.
458. Berthet A, Porrás G, Doudnikoff E, Stark H, Cador M, Bezard E, et al. Pharmacological Analysis Demonstrates Dramatic Alteration of D₁ Dopamine Receptor Neuronal Distribution in the Rat Analog of ^{sc}-DOPA-Induced Dyskinesia. *The Journal of Neuroscience*. 2009;29(15):4829-35.
459. Vanover KE, Betz AJ, Weber SM, Bibbiani F, Kielaitė A, Weiner DM, et al. A 5-HT_{2A} receptor inverse agonist, ACP-103, reduces tremor in a rat model and levodopa-induced dyskinesias in a monkey model. *Pharmacology, biochemistry, and behavior*. 2008;90(4):540-4.
460. Huot P, Fox SH. The serotonergic system in motor and non-motor manifestations of Parkinson's disease. *Exp Brain Res*. 2013;230(4):463-76.
461. Huot P. Pharmacological Properties of Levodopa. *Levodopa-Induced Dyskinesia in Parkinson's Disease* 2014. p. 147-69.
462. Stocchi F, Vacca L, Ruggieri S, Olanow CW. Intermittent vs continuous levodopa administration in patients with advanced Parkinson disease: a clinical and pharmacokinetic study. *Arch Neurol*. 2005;62(6):905-10.
463. Puente V, De Fabregues O, Oliveras C, Ribera G, Pont-Sunyer C, Vivanco R, et al. Eighteen month study of continuous intraduodenal levodopa infusion in patients with advanced Parkinson's disease: impact on control of fluctuations and quality of life. *Parkinsonism Relat Disord*. 2010;16(3):218-21.
464. Olanow CW, Kieburtz K, Odin P, Espay AJ, Standaert DG, Fernandez HH, et al. Continuous intrajejunal infusion of levodopa-carbidopa intestinal gel for patients with advanced Parkinson's disease: a randomised, controlled, double-blind, double-dummy study. *The Lancet Neurology*. 2014;13(2):141-9.
465. Cenci MA, Lundblad M. Post- versus presynaptic plasticity in L-DOPA-induced dyskinesia. *J Neurochem*. 2006;99(2):381-92.

466. Bargiotas P, Konitsiotis S. Levodopa-induced dyskinesias in Parkinson's disease: emerging treatments. *Neuropsychiatr Dis Treat*. 2013;9:1605-17.
467. Olanow CW, Agid Y, Mizuno Y, Albanese A, Bonuccelli U, Damier P, et al. Levodopa in the treatment of Parkinson's disease: current controversies. *Mov Disord*. 2004;19(9):997-1005.
468. Ahlskog JE, Muentner MD. Frequency of levodopa-related dyskinesias and motor fluctuations as estimated from the cumulative literature. *Mov Disord*. 2001;16(3):448-58.
469. Holloway RG, Shoulson I, Fahn S, Kieburtz K, Lang A, Marek K, et al. Pramipexole vs levodopa as initial treatment for Parkinson disease: a 4-year randomized controlled trial. *Arch Neurol*. 2004;61(7):1044-53.
470. Manson A, Stirpe P, Schrag A. Levodopa-induced-dyskinesias clinical features, incidence, risk factors, management and impact on quality of life. *J Parkinsons Dis*. 2012;2(3):189-98.
471. Pearce RK, Heikkila M, Linden IB, Jenner P. L-dopa induces dyskinesia in normal monkeys: behavioural and pharmacokinetic observations. *Psychopharmacology (Berl)*. 2001;156(4):402-9.
472. Togasaki DM, Tan L, Protell P, Di Monte DA, Quik M, Langston JW. Levodopa induces dyskinesias in normal squirrel monkeys. *Ann Neurol*. 2001;50(2):254-7.
473. Picconi B, Paille V, Ghiglieri V, Bagetta V, Barone I, Lindgren HS, et al. l-DOPA dosage is critically involved in dyskinesia via loss of synaptic depotentiation. *Neurobiol Dis*. 2008;29(2):327-35.
474. Mouradian MM, Heuser IJE, Baronti F, Fabbrini G, Juncos JL, Chase TN. Pathogenesis of dyskinesias in parkinson's disease. *Annals of Neurology*. 1989;25(5):523-6.
475. Del Sorbo F, Albanese A. Levodopa-induced dyskinesias and their management. *J Neurol*. 2008;255 Suppl 4:32-41.
476. Hashim HZ, Norlinah MI, Nafisah WY, Tan HJ, Raymond AA, Tamil AM. Risk factors and predictors of levodopa-induced dyskinesia among multiethnic Malaysians with Parkinson's disease. *Int J Neurosci*. 2014;124(3):187-91.
477. Parkinson Study Group. Impact of deprenyl and tocopherol treatment on Parkinson's disease in DATATOP patients requiring levodopa. *Ann Neurol*. 1996;39(1):37-45.
478. Blanchet PJ, Allard P, Gregoire L, Tardif F, Bedard PJ. Risk factors for peak dose dyskinesia in 100 levodopa-treated parkinsonian patients. *Can J Neurol Sci*. 1996;23(3):189-93.
479. Lyons KE, Hubble JP, Tröster AI, Pahwa R, Koller WC. Gender differences in Parkinson's disease. *Clinical neuropharmacology*. 1998;21(2):118-21.
480. Zappia M, Annesi G, Nicoletti G, et al. Sex differences in clinical and genetic determinants of levodopa peak-dose dyskinesias in parkinson disease: An exploratory study. *Archives of Neurology*. 2005;62(4):601-5.
481. Martinelli P, Contin M, Scaglione C, Riva R, Albani F, Baruzzi A. Levodopa pharmacokinetics and dyskinesias: are there sex-related differences? *Neurological sciences : official journal of the Italian Neurological Society and of the Italian Society of Clinical Neurophysiology*. 2003;24(3):192-3.
482. Arabia G, Zappia M, Bosco D, Crescibene L, Bagala A, Bastone L, et al. Body weight, levodopa pharmacokinetics and dyskinesia in Parkinson's disease. *Neurological sciences : official journal of the Italian Neurological Society and of the Italian Society of Clinical Neurophysiology*. 2002;23 Suppl 2:S53-4.

483. Sharma JC, Ross IN, Rascol O, Brooks D. Relationship between weight, levodopa and dyskinesia: the significance of levodopa dose per kilogram body weight. *Eur J Neurol*. 2008;15(5):493-6.
484. Lee J-Y, Jeon BS. Risk Factors for Levodopa-Induced Dyskinesia. In: Fox SH, Brotchie JM, editors. *Levodopa-Induced Dyskinesia in Parkinson's Disease*. London: Springer London; 2014. p. 51-68.
485. Hauser RA, Hsu A, Kell S, Espay AJ, Sethi K, Stacy M, et al. Extended-release carbidopa-levodopa (IPX066) compared with immediate-release carbidopa-levodopa in patients with Parkinson's disease and motor fluctuations: a phase 3 randomised, double-blind trial. *The Lancet Neurology*. 2013;12(4):346-56.
486. Ahlskog JE, Muenter MD, McManis PG, Bell GN, Bailey PA. Controlled-release Sinemet (CR-4): a double-blind crossover study in patients with fluctuating Parkinson's disease. *Mayo Clinic proceedings*. 1988;63(9):876-86.
487. Hutton JT, Morris JL, Bush DF, Smith ME, Liss CL, Reines S. Multicenter controlled study of Sinemet CR vs Sinemet (25/100) in advanced Parkinson's disease. *Neurology*. 1989;39(11 Suppl 2):67-72; discussion -3.
488. Koller WC, Hutton JT, Tolosa E, Capilldeo R. Immediate-release and controlled-release carbidopa/levodopa in PD: a 5-year randomized multicenter study. *Carbidopa/Levodopa Study Group. Neurology*. 1999;53(5):1012-9.
489. Cedarbaum JM, Hoey M, McDowell FH. A double-blind crossover comparison of Sinemet CR4 and standard Sinemet 25/100 in patients with Parkinson's disease and fluctuating motor performance. *J Neurol Neurosurg Psychiatry*. 1989;52(2):207-12.
490. Sage JI, Mark MH. Comparison of controlled-release Sinemet (CR4) and standard Sinemet (25 mg/100 mg) in advanced Parkinson's disease: a double-blind, crossover study. *Clinical neuropharmacology*. 1988;11(2):174-9.
491. Deleu D, Jacques M, Michotte Y, Ebinger G. Controlled-release carbidopa/levodopa (CR) in parkinsonian patients with response fluctuations on standard levodopa treatment: clinical and pharmacokinetic observations. *Neurology*. 1989;39(11 Suppl 2):88-92; discussion 5.
492. Stocchi F, Quinn NP, Barbato L, Patsalos PN, O'Connell MT, Ruggieri S, et al. Comparison between a fast and a slow release preparation of levodopa and a combination of the two: a clinical and pharmacokinetic study. *Clinical neuropharmacology*. 1994;17(1):38-44.
493. Quinn N, Marsden CD, Parkes JD. Complicated response fluctuations in Parkinson's disease: response to intravenous infusion of levodopa. *Lancet*. 1982;2(8295):412-5.
494. Schuh LA, Bennett JP, Jr. Suppression of dyskinesias in advanced Parkinson's disease. I. Continuous intravenous levodopa shifts dose response for production of dyskinesias but not for relief of parkinsonism in patients with advanced Parkinson's disease. *Neurology*. 1993;43(8):1545-50.
495. Schrag A, Quinn N. Dyskinesias and motor fluctuations in Parkinson's disease. A community-based study. *Brain*. 2000;123 (Pt 11):2297-305.
496. Sossi V, de la Fuente-Fernandez R, Schulzer M, Adams J, Stoessl J. Age-related differences in levodopa dynamics in Parkinson's: implications for motor complications. *Brain*. 2006;129(Pt 4):1050-8.
497. Sossi V, de la Fuente-Fernandez R, Schulzer M, Troiano AR, Ruth TJ, Stoessl AJ. Dopamine transporter relation to dopamine turnover in Parkinson's disease: a positron emission tomography study. *Ann Neurol*. 2007;62(5):468-74.

498. Cerasa A, Salsone M, Morelli M, Pugliese P, Arabia G, Gioia CM, et al. Age at onset influences neurodegenerative processes underlying PD with levodopa-induced dyskinesias. *Parkinsonism Relat Disord*. 2013;19(10):883-8.
499. group Ps. Impact of deprenyl and tocopherol treatment on Parkinson's disease in DATATOP patients requiring levodopa. Parkinson Study Group. *Ann Neurol*. 1996;39(1):37-45.
500. Schrag A, Ben-Shlomo Y, Brown R, Marsden CD, Quinn N. Young-onset Parkinson's disease revisited--clinical features, natural history, and mortality. *Mov Disord*. 1998;13(6):885-94.
501. Quinn N, Critchley P, Marsden CD. Young onset Parkinson's disease. *Mov Disord*. 1987;2(2):73-91.
502. Papapetropoulos S, Argyriou AA, Ellul J, Chroni E. Comparison of motor fluctuations and L-dopa-induced dyskinesias in patients with familial and sporadic Parkinson's disease. *Eur J Neurol*. 2004;11(2):115-9.
503. Khan NL, Katzenschlager R, Watt H, Bhatia KP, Wood NW, Quinn N, et al. Olfaction differentiates parkin disease from early-onset parkinsonism and Parkinson disease. *Neurology*. 2004;62(7):1224-6.
504. Khan NL, Graham E, Critchley P, Schrag AE, Wood NW, Lees AJ, et al. Parkin disease: a phenotypic study of a large case series. *Brain*. 2003;126(Pt 6):1279-92.
505. Dekker M, Bonifati V, van Swieten J, Leenders N, Galjaard R-J, Snijders P, et al. Clinical features and neuroimaging of PARK7-linked parkinsonism. *Movement Disorders*. 2003;18(7):751-7.
506. Nishioka K, Kefi M, Jasinska-Myga B, Wider C, Vilarino-Guell C, Ross OA, et al. A comparative study of LRRK2, PINK1 and genetically undefined familial Parkinson's disease. *J Neurol Neurosurg Psychiatry*. 2010;81(4):391-5.
507. Marras C, Schule B, Munhoz RP, Rogaeva E, Langston JW, Kasten M, et al. Phenotype in parkinsonian and nonparkinsonian LRRK2 G2019S mutation carriers. *Neurology*. 2011;77(4):325-33.
508. Lohmann E, Thobois S, Lesage S, Broussolle E, du Montcel ST, Ribeiro MJ, et al. A multidisciplinary study of patients with early-onset PD with and without parkin mutations. *Neurology*. 2009;72(2):110-6.
509. Gilgun-Sherki Y, Djaldetti R, Melamed E, Offen D. Polymorphism in candidate genes: implications for the risk and treatment of idiopathic Parkinson's disease. *Pharmacogenomics J*. 2004;4(5):291-306.
510. Kaiser R, Hofer A, Grapengiesser A, Gasser T, Kupsch A, Roots I, et al. L-Dopa-induced adverse effects in PD and dopamine transporter gene polymorphism. *Neurology*. 2003;60(11):1750-5.
511. Oliveri RL, Annesi G, Zappia M, Civitelli D, Montesanti R, Branca D, et al. Dopamine D2 receptor gene polymorphism and the risk of levodopa-induced dyskinesias in PD. *Neurology*. 1999;53(7):1425-30.
512. Colosimo C, Martinez-Martin P, Fabbrini G, Hauser RA, Merello M, Miyasaki J, et al. Task force report on scales to assess dyskinesia in Parkinson's disease: critique and recommendations. *Mov Disord*. 2010;25(9):1131-42.
513. Colosimo C, Fabbrini G, Berardelli A. Drug Insight: new drugs in development for Parkinson's disease. *Nat Clin Pract Neurol*. 2006;2(11):600-10.

514. Parkinson Study G. Evaluation of dyskinesias in a pilot, randomized, placebo-controlled trial of remacemide in advanced Parkinson disease. *Arch Neurol.* 2001;58(10):1660-8.
515. Katzenschlager R, Schrag A, Evans A, Manson A, Carroll CB, Ottaviani D, et al. Quantifying the impact of dyskinesias in PD: the PDYS-26: a patient-based outcome measure. *Neurology.* 2007;69(6):555-63.
516. Guy W, National Institute of Mental H, Psychopharmacology Research B, Early Clinical Drug Evaluation P. ECDEU assessment manual for psychopharmacology. Rockville, Md.: U.S. Dept. of Health, Education, and Welfare, Public Health Service, Alcohol, Drug Abuse, and Mental Health Administration, National Institute of Mental Health, Psychopharmacology Research Branch, Division of Extramural Research Programs; 1976.
517. Goetz CG, Stebbins GT, Shale HM, Lang AE, Chernik DA, Chmura TA, et al. Utility of an objective dyskinesia rating scale for Parkinson's disease: inter- and intrarater reliability assessment. *Mov Disord.* 1994;9(4):390-4.
518. Goetz CG, Colosimo C. Dyskinesia Rating Scales in Parkinson's Disease. In: Fox SH, Brotchie JM, editors. *Levodopa-Induced Dyskinesia in Parkinson's Disease.* London: Springer London; 2014. p. 17-31.
519. Goetz CG, Nutt JG, Stebbins GT. The Unified Dyskinesia Rating Scale: presentation and clinimetric profile. *Mov Disord.* 2008;23(16):2398-403.
520. Goetz CG, Nutt JG, Stebbins GT, Chmura TA. Teaching program for the Unified Dyskinesia Rating Scale. *Mov Disord.* 2009;24(9):1296-8.
521. Goetz CG, Stebbins GT, Theeuwes A, Stocchi F, Ferreira JJ, van de Witte S, et al. Temporal stability of the Unified Dyskinesia Rating Scale. *Mov Disord.* 2011;26(14):2556-9.
522. Katzenschlager R. Pharmacological Treatment Options for Levodopa-Induced Dyskinesia. In: Fox SH, Brotchie JM, editors. *Levodopa-Induced Dyskinesia in Parkinson's Disease.* London: Springer London; 2014. p. 69-88.
523. Verhagen Metman L, Del Dotto P, van den Munckhof P, Fang J, Mouradian MM, Chase TN. Amantadine as treatment for dyskinesias and motor fluctuations in Parkinson's disease. *Neurology.* 1998;50(5):1323-6.
524. Wolf E, Seppi K, Katzenschlager R, Hochschorner G, Ransmayr G, Schwingenschuh P, et al. Long-term antidyskinetic efficacy of amantadine in Parkinson's disease. *Mov Disord.* 2010;25(10):1357-63.
525. Crosby N, Deane KH, Clarke CE. Amantadine in Parkinson's disease. *Cochrane Database Syst Rev.* 2003(1):CD003468.
526. Schmid CL, Streicher JM, Meltzer HY, Bohn LM. Clozapine Acts as an Agonist at Serotonin 2A Receptors to Counter MK-801-Induced Behaviors through a β Arrestin2-Independent Activation of Akt. *Neuropsychopharmacology.* 2014;39(8):1902-13.
527. Bennett JP, Landow ER, Schuh LA. Suppression of dyskinesias in advanced Parkinson's disease: II. Increasing daily clozapine doses suppress dyskinesias and improve parkinsonism symptoms. *Neurology.* 1993;43(8):1551.
528. Durif F, Debilly B, Galitzky M, Morand D, Viallet F, Borg M, et al. Clozapine improves dyskinesias in Parkinson disease: a double-blind, placebo-controlled study. *Neurology.* 2004;62(3):381-8.
529. Alvir JMJ, Lieberman JA, Safferman AZ, Schwimmer JL, Schaaf JA. Clozapine-Induced Agranulocytosis -- Incidence and Risk Factors in the United States. *New England Journal of Medicine.* 1993;329(3):162-7.

530. Rajagopal S. Clozapine, agranulocytosis, and benign ethnic neutropenia. *Postgraduate Medical Journal*. 2005;81(959):545-6.
531. Follett KA. Comparison of pallidal and subthalamic deep brain stimulation for the treatment of levodopa-induced dyskinesias. *Neurosurgical focus*. 2004;17(1):E3.
532. Benabid AL, Pollak P, Louveau A, Henry S, de Rougemont J. Combined (thalamotomy and stimulation) stereotactic surgery of the VIM thalamic nucleus for bilateral Parkinson disease. *Appl Neurophysiol*. 1987;50(1-6):344-6.
533. Anderson JC, Costantino MM, Stratford T. Basal ganglia: anatomy, pathology, and imaging characteristics. *Curr Probl Diagn Radiol*. 2004;33(1):28-41.
534. Guridi J, Lozano AM. A Brief History of Pallidotomy. *Neurosurgery*. 1997;41(5):1169-83.
535. Gillingham J. Forty-Five Years of Stereotactic Surgery for Parkinson's Disease: A Review. *Stereotactic and functional neurosurgery*. 2000;74(3-4):95-8.
536. Toda H, Saiki H, Nishida N, Iwasaki K. Update on Deep Brain Stimulation for Dyskinesia and Dystonia: A Literature Review. *Neurologia medico-chirurgica*. 2016;56(5):236-48.
537. Crossman AR, Neary D, Crossman B. *Neuroanatomy : an illustrated colour text*. 2015.
538. Ko WKD, Bastide M, Bezard E. Basal Ganglia Circuitry Models of Levodopa-Induced Dyskinesia. In: Fox SH, Brotchie JM, editors. *Levodopa-Induced Dyskinesia in Parkinson's Disease*. London: Springer London; 2014. p. 109-34.
539. Marsden CD, Obeso JA. The functions of the basal ganglia and the paradox of stereotaxic surgery in Parkinson's disease. *Brain*. 1994;117 (Pt 4):877-97.
540. Martin KE, Phillips JG, Iansek R, Bradshaw JL. Inaccuracy and instability of sequential movements in Parkinson's disease. *Exp Brain Res*. 1994;102(1):131-40.
541. Haber SN, Calzavara R. The cortico-basal ganglia integrative network: the role of the thalamus. *Brain Res Bull*. 2009;78(2-3):69-74.
542. Lincoln CM, Bello JA, Lui YW. Decoding the Deep Gray: A Review of the Anatomy, Function, and Imaging Patterns Affecting the Basal Ganglia. *Neurographics*. 2012;2(3):92-102.
543. Alexander GE, Crutcher MD. Functional architecture of basal ganglia circuits: neural substrates of parallel processing. *Trends in neurosciences*. 1990;13(7):266-71.
544. Wilson S. Disorders of motility and tone. *The Lancet Neurology*. 1925;1:1-103.
545. Martin JP. HEMICHOREA RESULTING FROM A LOCAL LESION OF THE BRAIN. (THE SYNDROME OF THE BODY OF LUYS. *Brain*. 1927;50(3-4):637-49.
546. Lanciego JL, Luquin N, Obeso JA. Functional Neuroanatomy of the Basal Ganglia. *Cold Spring Harbor Perspectives in Medicine*. 2012;2(12):a009621.
547. Parent A, Hazrati LN. Functional anatomy of the basal ganglia. I. The cortico-basal ganglia-thalamo-cortical loop. *Brain Res Brain Res Rev*. 1995;20(1):91-127.
548. Alexander GE, Crutcher MD, DeLong MR. Basal ganglia-thalamocortical circuits: parallel substrates for motor, oculomotor, "prefrontal" and "limbic" functions. *Prog Brain Res*. 1990;85:119-46.
549. Alexander GE, DeLong MR, Strick PL. Parallel organization of functionally segregated circuits linking basal ganglia and cortex. *Annu Rev Neurosci*. 1986;9:357-81.
550. Hoover JE, Strick PL. Multiple output channels in the basal ganglia. *Science*. 1993;259(5096):819-21.

551. Lanciego JL, Gonzalo N, Castle M, Sanchez-Escobar C, Aymerich MS, Obeso JA. Thalamic innervation of striatal and subthalamic neurons projecting to the rat entopeduncular nucleus. *Eur J Neurosci.* 2004;19(5):1267-77.
552. Wise SP, Murray EA, Gerfen CR. The frontal cortex-basal ganglia system in primates. *Critical reviews in neurobiology.* 1996;10(3-4):317-56.
553. Afifi AK. Topical Review: Basal Ganglia: Functional Anatomy and Physiology. Part 2. *Journal of Child Neurology.* 1994;9(4):352-61.
554. Albin RL, Young AB, Penney JB. The functional anatomy of basal ganglia disorders. *Trends in neurosciences.* 1989;12(10):366-75.
555. Gerfen CR, Wilson CJ. Chapter II The basal ganglia. In: Swanson LW, Björklund A, Hökfelt T, editors. *Handbook of Chemical Neuroanatomy.* 12: Elsevier; 1996. p. 371-468.
556. Groenewegen HJ, Wright CI, Beijer AV. The nucleus accumbens: gateway for limbic structures to reach the motor system? *Prog Brain Res.* 1996;107:485-511.
557. Kita H. Chapter 4 GABAergic circuits of the striatum. In: Arbuthnott GW, Emson PC, editors. *Progress in Brain Research.* 99: Elsevier; 1993. p. 51-72.
558. Haber SN. *Frontiers in Neuroscience Neuroanatomy of Reward: A View from the Ventral Striatum.* In: Gottfried JA, editor. *Neurobiology of Sensation and Reward.* Boca Raton (FL): CRC Press/Taylor & Francis Llc.; 2011.
559. Lavoie B, Smith Y, Parent A. Dopaminergic innervation of the basal ganglia in the squirrel monkey as revealed by tyrosine hydroxylase immunohistochemistry. *The Journal of comparative neurology.* 1989;289(1):36-52.
560. Lavoie B, Parent A. Immunohistochemical study of the serotonergic innervation of the basal ganglia in the squirrel monkey. *The Journal of comparative neurology.* 1990;299(1):1-16.
561. Delfs JM, Zhu Y, Druhan JP, Aston-Jones GS. Origin of noradrenergic afferents to the shell subregion of the nucleus accumbens: anterograde and retrograde tract-tracing studies in the rat. *Brain Res.* 1998;806(2):127-40.
562. Kunzle H. Bilateral projections from precentral motor cortex to the putamen and other parts of the basal ganglia. An autoradiographic study in *Macaca fascicularis.* *Brain Res.* 1975;88(2):195-209.
563. Künzle H. Projections from the primary somatosensory cortex to basal ganglia and thalamus in the monkey. *Experimental Brain Research.* 1977;30(4):481-92.
564. McGeorge AJ, Faull RLM. The organization of the projection from the cerebral cortex to the striatum in the rat. *Neuroscience.* 1989;29(3):503-37.
565. Somogyi P, Smith AD. Projection of neostriatal spiny neurons to the substantia nigra. Application of a combined Golgi-staining and horseradish peroxidase transport procedure at both light and electron microscopic levels. *Brain Res.* 1979;178(1):3-15.
566. Somogyi P, Bolam JP, Totterdell S, Smith AD. Monosynaptic input from the nucleus accumbens--ventral striatum region to retrogradely labelled nigrostriatal neurones. *Brain Res.* 1981;217(2):245-63.
567. Galvan L, André VM, Wang EA, Cepeda C, Levine MS. Functional Differences Between Direct and Indirect Striatal Output Pathways in Huntington's Disease. *Journal of Huntington's disease.* 2012;1(1):17-25.
568. Smith AD, Bolam JP. The neural network of the basal ganglia as revealed by the study of synaptic connections of identified neurones. *Trends in neurosciences.* 1990;13(7):259-65.

569. Kawaguchi Y, Wilson CJ, Augood SJ, Emson PC. Striatal interneurons: chemical, physiological and morphological characterization. *Trends in neurosciences*. 1995;18(12):527-35.
570. Zucca FA, Basso E, Cupaioli FA, Ferrari E, Sulzer D, Casella L, et al. Neuromelanin of the Human Substantia Nigra: An Update. *Neurotoxicity Research*. 2014;25(1):13-23.
571. Grofova I, Deniau JM, Kitai ST. Morphology of the substantia nigra pars reticulata projection neurons intracellularly labeled with HRP. *The Journal of comparative neurology*. 1982;208(4):352-68.
572. DeLong MR. Primate models of movement disorders of basal ganglia origin. *Trends in neurosciences*. 1990;13(7):281-5.
573. Deniau JM, Chevalier G. Synaptic organization of the basal ganglia: an electroanatomical approach in the rat. *Ciba Foundation symposium*. 1984;107:48-63.
574. Nambu A, Tokuno H, Hamada I, Kita H, Imanishi M, Akazawa T, et al. Excitatory cortical inputs to pallidal neurons via the subthalamic nucleus in the monkey. *J Neurophysiol*. 2000;84(1):289-300.
575. Hamani C, Saint-Cyr JA, Fraser J, Kaplitt M, Lozano AM. The subthalamic nucleus in the context of movement disorders. *Brain*. 2004;127(Pt 1):4-20.
576. Jaeger D, Kita H. Functional connectivity and integrative properties of globus pallidus neurons. *Neuroscience*. 2011;198:44-53.
577. Kita H, Kitai ST. Efferent projections of the subthalamic nucleus in the rat: light and electron microscopic analysis with the PHA-L method. *The Journal of comparative neurology*. 1987;260(3):435-52.
578. Parent A, Smith Y. Organization of efferent projections of the subthalamic nucleus in the squirrel monkey as revealed by retrograde labeling methods. *Brain Res*. 1987;436(2):296-310.
579. Carpenter MB, Jayaraman A. Subthalamic Nucleus Afferents: Anatomical and Immunocytochemical Features. In: Bernardi G, Carpenter MB, Di Chiara G, Morelli M, Stanzione P, editors. *The Basal Ganglia III*. Boston, MA: Springer New York; 1991. p. 109-17.
580. Parent A, Sato F, Wu Y, Gauthier J, Levesque M, Parent M. Organization of the basal ganglia: the importance of axonal collateralization. *Trends in neurosciences*. 2000;23(10 Suppl):S20-7.
581. Dudman JT, Gerfen CR. Chapter 17 - The Basal Ganglia A2 - Paxinos, George. *The Rat Nervous System (Fourth Edition)*. San Diego: Academic Press; 2015. p. 391-440.
582. MacLeod NK, James TA, Kilpatrick IC, Starr MS. Evidence for a GABAergic nigrothalamic pathway in the rat. II. Electrophysiological studies. *Exp Brain Res*. 1980;40(1):55-61.
583. Anderson ME, Postupna N, Ruffo M. Effects of high-frequency stimulation in the internal globus pallidus on the activity of thalamic neurons in the awake monkey. *J Neurophysiol*. 2003;89(2):1150-60.
584. Gerfen CR. The neostriatal mosaic: multiple levels of compartmental organization. *Trends in neurosciences*. 1992;15(4):133-9.
585. Kandel ER. *Principles of neural science* 2013.
586. Obeso JA, Rodriguez-Oroz MC, Rodriguez M, DeLong MR, Olanow CW. Pathophysiology of levodopa-induced dyskinesias in Parkinson's disease: problems with the current model. *Ann Neurol*. 2000;47(4 Suppl 1):S22-32; discussion S-4.

587. Bezard E, Crossman AR, Gross CE, Brotchie JM. Structures outside the basal ganglia may compensate for dopamine loss in the presymptomatic stages of Parkinson's disease. *FASEB journal : official publication of the Federation of American Societies for Experimental Biology*. 2001;15(6):1092-4.
588. Crossman AR, Mitchell IJ, Sambrook MA. Regional brain uptake of 2-deoxyglucose in N-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)—induced parkinsonism in the macaque monkey. *Neuropharmacology*. 1985;24(6):587-91.
589. Mitchell IJ, Cross AJ, Sambrook MA, Crossman AR. N-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-induced parkinsonism in the monkey: neurochemical pathology and regional brain metabolism. *Journal of neural transmission Supplementum*. 1986;20:41-6.
590. Mitchell IJ, Clarke CE, Boyce S, Robertson RG, Peggs D, Sambrook MA, et al. Neural mechanisms underlying parkinsonian symptoms based upon regional uptake of 2-deoxyglucose in monkeys exposed to 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine. *Neuroscience*. 1989;32(1):213-26.
591. Fillion M, Tremblay L, Bedard PJ. Effects of dopamine agonists on the spontaneous activity of globus pallidus neurons in monkeys with MPTP-induced parkinsonism. *Brain Res*. 1991;547(1):152-61.
592. Bergman H, Wichmann T, Karmon B, DeLong MR. The primate subthalamic nucleus. II. Neuronal activity in the MPTP model of parkinsonism. *J Neurophysiol*. 1994;72(2):507-20.
593. Soares J, Kliem MA, Betarbet R, Greenamyre JT, Yamamoto B, Wichmann T. Role of external pallidal segment in primate parkinsonism: comparison of the effects of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-induced parkinsonism and lesions of the external pallidal segment. *J Neurosci*. 2004;24(29):6417-26.
594. Miller R. The “Indirect” Pathways from Striatum to Basal Ganglia Output Nuclei, and Their Relation to the “Direct” Pathway. A theory of the basal ganglia and their disorders. Boca Raton: CRC Press; 2008.
595. Crossman AR. A hypothesis on the pathophysiological mechanisms that underlie levodopa- or dopamine agonist-induced dyskinesia in Parkinson's disease: implications for future strategies in treatment. *Mov Disord*. 1990;5(2):100-8.
596. Vidailhet M, Bonnet AM, Marconi R, Durif F, Agid Y. The phenomenology of L-dopa-induced dyskinesias in Parkinson's disease. *Mov Disord*. 1999;14 Suppl 1:13-8.
597. Obeso JA, Rodriguez-Oroz MC, Rodriguez M, Lanciego JL, Artieda J, Gonzalo N, et al. Pathophysiology of the basal ganglia in Parkinson's disease. *Trends in neurosciences*. 2000;23(10):S8-S19.
598. Brooks DJ, Piccini P, Turjanski N, Samuel M. Neuroimaging of dyskinesia. *Ann Neurol*. 2000;47(4 Suppl 1):S154-8; discussion S8-9.
599. Rascol O, Sabatini U, Brefel C, Fabre N, Rai S, Senard JM, et al. Cortical motor overactivation in parkinsonian patients with L-dopa-induced peak-dose dyskinesia. *Brain*. 1998;121 (Pt 3):527-33.
600. Mitchell IJ, Boyce S, Sambrook MA, Crossman AR. A 2-deoxyglucose study of the effects of dopamine agonists on the parkinsonian primate brain. Implications for the neural mechanisms that mediate dopamine agonist-induced dyskinesia. *Brain*. 1992;115 (Pt 3):809-24.
601. Vila M, Levy R, Herrero MT, Ruberg M, Faucheux B, Obeso JA, et al. Consequences of nigrostriatal denervation on the functioning of the basal ganglia in human and nonhuman

- primates: an in situ hybridization study of cytochrome oxidase subunit I mRNA. *J Neurosci.* 1997;17(2):765-73.
602. Cenci MA, Lee CS, Bjorklund A. L-DOPA-induced dyskinesia in the rat is associated with striatal overexpression of prodynorphin- and glutamic acid decarboxylase mRNA. *Eur J Neurosci.* 1998;10(8):2694-706.
603. Calon F, Birdi S, Rajput AH, Hornykiewicz O, Bedard PJ, Di Paolo T. Increase of preproenkephalin mRNA levels in the putamen of Parkinson disease patients with levodopa-induced dyskinesias. *Journal of neuropathology and experimental neurology.* 2002;61(2):186-96.
604. Bezard E, Brotchie JM, Gross CE. Pathophysiology of levodopa-induced dyskinesia: potential for new therapies. *Nat Rev Neurosci.* 2001;2(8):577-88.
605. Papa SM, Desimone R, Fiorani M, Oldfield EH. Internal globus pallidus discharge is nearly suppressed during levodopa-induced dyskinesias. *Annals of Neurology.* 1999;46(5):732-8.
606. Boraud T, Bezard E, Guehl D, Bioulac B, Gross C. Effects of L-DOPA on neuronal activity of the globus pallidus externalis (GPe) and globus pallidus internalis (GPi) in the MPTP-treated monkey. *Brain Res.* 1998;787(1):157-60.
607. Boraud T, Bezard E, Bioulac B, Gross CE. Dopamine agonist-induced dyskinesias are correlated to both firing pattern and frequency alterations of pallidal neurones in the MPTP-treated monkey. *Brain.* 2001;124(Pt 3):546-57.
608. Merello M, Balej J, Delfino M, Cammarota A, Betti O, Leiguarda R. Apomorphine induces changes in GPi spontaneous outflow in patients with parkinson's disease. *Movement Disorders.* 1999;14(1):45-9.
609. Lozano AM, Lang AE, Levy R, Hutchison W, Dostrovsky J. Neuronal recordings in Parkinson's disease patients with dyskinesias induced by apomorphine. *Ann Neurol.* 2000;47(4 Suppl 1):S141-6.
610. Stefani A, Stanzione P, Bassi A, Mazzone P, Vangelista T, Bernardi G. Effects of increasing doses of apomorphine during stereotaxic neurosurgery in Parkinson's disease: clinical score and internal globus pallidus activity. Short communication. *Journal of neural transmission (Vienna, Austria : 1996).* 1997;104(8-9):895-904.
611. Ravenscroft P, Chalon S, Brotchie JM, Crossman AR. Ropinirole versus L-DOPA effects on striatal opioid peptide precursors in a rodent model of Parkinson's disease: implications for dyskinesia. *Exp Neurol.* 2004;185(1):36-46.
612. Henry B, Crossman AR, Brotchie JM. Effect of repeated L-DOPA, bromocriptine, or lisuride administration on preproenkephalin-A and preproenkephalin-B mRNA levels in the striatum of the 6-hydroxydopamine-lesioned rat. *Exp Neurol.* 1999;155(2):204-20.
613. Lundblad M, Picconi B, Lindgren H, Cenci MA. A model of L-DOPA-induced dyskinesia in 6-hydroxydopamine lesioned mice: relation to motor and cellular parameters of nigrostriatal function. *Neurobiol Dis.* 2004;16(1):110-23.
614. Aubert I, Guigoni C, Li Q, Dovero S, Bioulac BH, Gross CE, et al. Enhanced Preproenkephalin-B-Derived Opioid Transmission in Striatum and Subthalamic Nucleus Converges Upon Globus Pallidus Internalis in L-3,4-dihydroxyphenylalanine-Induced Dyskinesia. *Biological Psychiatry.* 2007;61(7):836-44.
615. Rascol O, Nutt JG, Blin O, Goetz CG, Trugman JM, Soubrouillard C, et al. Induction by dopamine D1 receptor agonist ABT-431 of dyskinesia similar to levodopa in patients with Parkinson disease. *Arch Neurol.* 2001;58(2):249-54.

616. Nadjar A, Brotchie JM, Guigoni C, Li Q, Zhou SB, Wang GJ, et al. Phenotype of striatofugal medium spiny neurons in parkinsonian and dyskinetic nonhuman primates: a call for a reappraisal of the functional organization of the basal ganglia. *J Neurosci*. 2006;26(34):8653-61.
617. Barroso-Chinea P, Bezard E. Basal Ganglia circuits underlying the pathophysiology of levodopa-induced dyskinesia. *Front Neuroanat*. 2010;4.
618. Stephens B, Mueller AJ, Shering AF, Hood SH, Taggart P, Arbuthnott GW, et al. Evidence of a breakdown of corticostriatal connections in Parkinson's disease. *Neuroscience*. 2005;132(3):741-54.
619. Ingham CA, Hood SH, Taggart P, Arbuthnott GW. Plasticity of Synapses in the Rat Neostriatum after Unilateral Lesion of the Nigrostriatal Dopaminergic Pathway. *The Journal of Neuroscience*. 1998;18(12):4732-43.
620. Scholz B, Svensson M, Alm H, Sköld K, Fälth M, Kultima K, et al. Striatal Proteomic Analysis Suggests that First L-Dopa Dose Equates to Chronic Exposure. *PLOS ONE*. 2008;3(2):e1589.
621. Villalba RM, Lee H, Smith Y. Dopaminergic Denervation and Spine Loss in the Striatum of MPTP-treated Monkeys. *Experimental neurology*. 2009;215(2):220-7.
622. Obeso JA, Rodriguez MC, DeLong MR. Basal ganglia pathophysiology. A critical review. *Adv Neurol*. 1997;74:3-18.
623. Iravani MM, Costa S, Al-Bargouthy G, Jackson MJ, Zeng B-Y, Kuoppamäki M, et al. Unilateral pallidotomy in 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-treated common marmosets exhibiting levodopa-induced dyskinesia. *European Journal of Neuroscience*. 2005;22(6):1305-18.
624. Lang AE. Surgery for levodopa-induced dyskinesias. *Ann Neurol*. 2000;47(4 Suppl 1):S193-9; discussion S9-202.
625. Parkin SG, Gregory RP, Scott R, Bain P, Silburn P, Hall B, et al. Unilateral and bilateral pallidotomy for idiopathic Parkinson's disease: a case series of 115 patients. *Mov Disord*. 2002;17(4):682-92.
626. McHaffie JG, Stanford TR, Stein BE, Coizet V, Redgrave P. Subcortical loops through the basal ganglia. *Trends in neurosciences*. 2005;28(8):401-7.
627. Graybiel AM. Habits, rituals, and the evaluative brain. *Annu Rev Neurosci*. 2008;31:359-87.
628. Obeso JA, Rodriguez-Oroz MC, Benitez-Temino B, Blesa FJ, Guridi J, Marin C, et al. Functional organization of the basal ganglia: therapeutic implications for Parkinson's disease. *Mov Disord*. 2008;23 Suppl 3:S548-59.
629. Lévesque M, Parent A. The striatofugal fiber system in primates: A reevaluation of its organization based on single-axon tracing studies. *Proceedings of the National Academy of Sciences of the United States of America*. 2005;102(33):11888-93.
630. Obeso JA, Rodriguez-Oroz MC, Javier Blesa F, Guridi J. The globus pallidus pars externa and Parkinson's disease. Ready for prime time? *Exp Neurol*. 2006;202(1):1-7.
631. Kita H. Globus pallidus external segment. *Prog Brain Res*. 2007;160:111-33.
632. Mena-Segovia J, Bolam JP, Magill PJ. Pedunculopontine nucleus and basal ganglia: distant relatives or part of the same family? *Trends in neurosciences*. 2004;27(10):585-8.
633. Haber SN, Fudge JL, McFarland NR. Striatonigrostriatal pathways in primates form an ascending spiral from the shell to the dorsolateral striatum. *J Neurosci*. 2000;20(6):2369-82.

634. Sato F, Lavallee P, Levesque M, Parent A. Single-axon tracing study of neurons of the external segment of the globus pallidus in primate. *The Journal of comparative neurology*. 2000;417(1):17-31.
635. Mallet N, Micklem BR, Henny P, Brown MT, Williams C, Bolam JP, et al. Dichotomous organization of the external globus pallidus. *Neuron*. 2012;74(6):1075-86.
636. Kita H, Tachibana Y, Nambu A, Chiken S. Balance of monosynaptic excitatory and disynaptic inhibitory responses of the globus pallidus induced after stimulation of the subthalamic nucleus in the monkey. *J Neurosci*. 2005;25(38):8611-9.
637. Obeso J, Lanciego J. Past, Present, and Future of the Pathophysiological Model of the Basal Ganglia. *Frontiers in Neuroanatomy*. 2011;5(39).
638. Emson PC, Waldvogel HJ, Faull RLM. Neurotransmitter Receptors in the Basal Ganglia. *HANDBOOK OF BEHAVIORAL NEUROSCIENCE*. 2010;20:75-98.
639. Rylander D. The serotonin system: a potential target for anti-dyskinetic treatments and biomarker discovery. *Parkinsonism & Related Disorders*. 2012;18:S126-S8.
640. Arai R, Karasawa N, Geffard M, Nagatsu I. L-DOPA is converted to dopamine in serotonergic fibers of the striatum of the rat: a double-labeling immunofluorescence study. *Neurosci Lett*. 1995;195(3):195-8.
641. Carta M, Carlsson T, Kirik D, Bjorklund A. Dopamine released from 5-HT terminals is the cause of L-DOPA-induced dyskinesia in parkinsonian rats. *Brain*. 2007;130(Pt 7):1819-33.
642. Carta M, Bezard E. Contribution of pre-synaptic mechanisms to L-DOPA-induced dyskinesia. *Neuroscience*. 2011;198:245-51.
643. Lodish HF. *Molecular cell biology* 2016.
644. Carta M, Carlsson T, Munoz A, Kirik D, Bjorklund A. Serotonin-dopamine interaction in the induction and maintenance of L-DOPA-induced dyskinesias. *Prog Brain Res*. 2008;172:465-78.
645. Sprouse JS, Aghajanian GK. Electrophysiological responses of serotonergic dorsal raphe neurons to 5-HT_{1A} and 5-HT_{1B} agonists. *Synapse*. 1987;1(1):3-9.
646. Kia HK, Miquel MC, Brisorgueil MJ, Daval G, Riad M, El Mestikawy S, et al. Immunocytochemical localization of serotonin_{1A} receptors in the rat central nervous system. *The Journal of comparative neurology*. 1996;365(2):289-305.
647. Göthert M, Schlicker E, Fink K, Classen K. Effects of RU 24969 on serotonin release in rat brain cortex: further support for the identity of serotonin autoreceptors with 5-HT_{1B} sites. *Arch Int Pharmacodyn Ther*. 1987;288(1):31-42.
648. Knobelmann DA, Kung HF, Lucki I. Regulation of extracellular concentrations of 5-hydroxytryptamine (5-HT) in mouse striatum by 5-HT_{1A} and 5-HT_{1B} receptors. *J Pharmacol Exp Ther*. 2000;292(3):1111-7.
649. Riad M, Garcia S, Watkins KC, Jodoin N, Doucet E, Langlois X, et al. Somatodendritic localization of 5-HT_{1A} and preterminal axonal localization of 5-HT_{1B} serotonin receptors in adult rat brain. *The Journal of comparative neurology*. 2000;417(2):181-94.
650. Adell A, Celada P, Artigas F. The role of 5-HT_{1B} receptors in the regulation of serotonin cell firing and release in the rat brain. *J Neurochem*. 2001;79(1):172-82.
651. Arai R, Karasawa N, Geffard M, Nagatsu T, Nagatsu I. Immunohistochemical evidence that central serotonin neurons produce dopamine from exogenous L-DOPA in the rat, with reference to the involvement of aromatic L-amino acid decarboxylase. *Brain Res*. 1994;667(2):295-9.

652. Dimatteo V, Digiovanni G, Pierucci M, Esposito E. Serotonin control of central dopaminergic function: focus on in vivo microdialysis studies. *Serotonin–Dopamine Interaction: Experimental Evidence and Therapeutic Relevance. Progress in Brain Research*2008. p. 7-44.
653. Palkovits M, Brownstein M, Saavedra JM. Serotonin content of the brain stem nuclei in the rat. *Brain Res.* 1974;80(2):237-49.
654. Saavedra JM. Distribution of serotonin and synthesizing enzymes in discrete areas of the brain. *Federation proceedings.* 1977;36(8):2134-41.
655. Miguelez C, Morera-Herreras T, Torrecilla M, Ruiz-Ortega JA, Ugedo L. Interaction between the 5-HT system and the basal ganglia: functional implication and therapeutic perspective in Parkinson's disease. *Frontiers in Neural Circuits.* 2014;8(21).
656. Reed M, Nijhout H, Best J. Computational studies of the role of serotonin in the basal ganglia. *Frontiers in Integrative Neuroscience.* 2013;7(41).
657. Parent M, Wallman MJ, Gagnon D, Parent A. Serotonin innervation of basal ganglia in monkeys and humans. *J Chem Neuroanat.* 2011;41(4):256-65.
658. Delaville C, Chetrit J, Abdallah K, Morin S, Carroit L, De Deurwaerdere P, et al. Emerging dysfunctions consequent to combined monoaminergic depletions in Parkinsonism. *Neurobiol Dis.* 2012;45(2):763-73.
659. Sinton CM, Fallon SL. Electrophysiological evidence for a functional differentiation between subtypes of the 5-HT1 receptor. *European journal of pharmacology.* 1988;157(2-3):173-81.
660. Arborelius L, Chergui K, Murase S, Nomikos GG, Backlund Höök B, Chouvet G, et al. The 5-HT1A receptor selective ligands, (R)-8-OH-DPAT and (S)-UH-301, differentially affect the activity of midbrain dopamine neurons. *Naunyn-Schmiedeberg's archives of pharmacology.* 1993;347(4):353-62.
661. Kelland MD, Freeman AS, Chiodo LA. Serotonergic afferent regulation of the basic physiology and pharmacological responsiveness of nigrostriatal dopamine neurons. *J Pharmacol Exp Ther.* 1990;253(2):803-11.
662. Olpe H-R, Koella WP. The response of striatal cells upon stimulation of the dorsal and median raphe nuclei. *Brain Research.* 1977;122(2):357-60.
663. Davies J, Tongroach P. Neuropharmacological studies on the nigro-striatal and raphe-striatal system in the rat. *Eur J Pharmacol.* 1978;51(2):91-100.
664. Yakel JL, Trussell LO, Jackson MB. Three serotonin responses in cultured mouse hippocampal and striatal neurons. *J Neurosci.* 1988;8(4):1273-85.
665. Gerber R, Altar CA, Liebman JM. Rotational behavior induced by 8-hydroxy-DPAT, a putative 5HT-1A agonist, in 6-hydroxydopamine-lesioned rats. *Psychopharmacology (Berl).* 1988;94(2):178-82.
666. Antonelli T, Fuxe K, Tomasini MC, Bartoszyk GD, Seyfried CA, Tanganelli S, et al. Effects of sarizotan on the corticostriatal glutamate pathways. *Synapse.* 2005;58(3):193-9.
667. Dupre KB, Ostock CY, Eskow Jaunarajs KL, Button T, Savage LM, Wolf W, et al. Local modulation of striatal glutamate efflux by serotonin 1A receptor stimulation in dyskinetic, hemiparkinsonian rats. *Exp Neurol.* 2011;229(2):288-99.
668. Dupre KB, Ostock CY, George JA, Eskow Jaunarajs KL, Hueston CM, Bishop C. Effects of 5-HT(1A) Receptor Stimulation on D1 Receptor Agonist-Induced Striatonigral Activity and Dyskinesia in Hemiparkinsonian Rats. *ACS Chemical Neuroscience.* 2013;4(5):747-60.

669. el Mansari M, Radja F, Ferron A, Reader TA, Molina-Holgado E, Descarries L. Hypersensitivity to serotonin and its agonists in serotonin-hyperinnervated neostriatum after neonatal dopamine denervation. *Eur J Pharmacol.* 1994;261(1-2):171-8.
670. el Mansari M, Blier P. In vivo electrophysiological characterization of 5-HT receptors in the guinea pig head of caudate nucleus and orbitofrontal cortex. *Neuropharmacology.* 1997;36(4-5):577-88.
671. Stanford IM, Kantaria MA, Chahal HS, Loucif KC, Wilson CL. 5-Hydroxytryptamine induced excitation and inhibition in the subthalamic nucleus: action at 5-HT(2C), 5-HT(4) and 5-HT(1A) receptors. *Neuropharmacology.* 2005;49(8):1228-34.
672. Parent M, Wallman M-J, Descarries L. Distribution and ultrastructural features of the serotonin innervation in rat and squirrel monkey subthalamic nucleus. *European Journal of Neuroscience.* 2010;31(7):1233-42.
673. Wallman MJ, Gagnon D, Parent M. Serotonin innervation of human basal ganglia. *Eur J Neurosci.* 2011;33(8):1519-32.
674. Liu J, Chu YX, Zhang QJ, Wang S, Feng J, Li Q. 5,7-Dihydroxytryptamine lesion of the dorsal raphe nucleus alters neuronal activity of the subthalamic nucleus in normal and 6-hydroxydopamine-lesioned rats. *Brain Research.* 2007;1149(Supplement C):216-22.
675. Aristieta A, Morera-Herreras T, Ruiz-Ortega JA, Miguelez C, Vidaurrazaga I, Arrue A, et al. Modulation of the subthalamic nucleus activity by serotonergic agents and fluoxetine administration. *Psychopharmacology (Berl).* 2014;231(9):1913-24.
676. Rav-Acha M, Bergman H, Yarom Y. Pre- and postsynaptic serotonergic excitation of globus pallidus neurons. *J Neurophysiol.* 2008;100(2):1053-66.
677. Delaville C, Navailles S, Benazzouz A. Effects of noradrenaline and serotonin depletions on the neuronal activity of globus pallidus and substantia nigra pars reticulata in experimental parkinsonism. *Neuroscience.* 2012;202:424-33.
678. Chen L, Yung KK, Chan YS, Yung WH. 5-HT excites globus pallidus neurons by multiple receptor mechanisms. *Neuroscience.* 2008;151(2):439-51.
679. Querejeta E, Oviedo-Chávez A, Araujo-Alvarez JM, Quiñones-Cárdenas AR, Delgado A. In vivo effects of local activation and blockade of 5-HT1B receptors on globus pallidus neuronal spiking. *Brain research.* 2005;1043(1-2):186-94.
680. Commins DL, Shaughnessy RA, Axt KJ, Vosmer G, Seiden LS. Variability among brain regions in the specificity of 6-hydroxydopamine (6-OHDA)-induced lesions. *J Neural Transm.* 1989;77(2-3):197-210.
681. Balcioglu A, Zhang K, Tarazi FI. Dopamine depletion abolishes apomorphine- and amphetamine-induced increases in extracellular serotonin levels in the striatum of conscious rats: a microdialysis study. *Neuroscience.* 2003;119(4):1045-53.
682. Breese GR, Baumeister AA, McCown TJ, Emerick SG, Frye GD, Crotty K, et al. Behavioral differences between neonatal and adult 6-hydroxydopamine-treated rats to dopamine agonists: relevance to neurological symptoms in clinical syndromes with reduced brain dopamine. *J Pharmacol Exp Ther.* 1984;231(2):343-54.
683. Aguiar LMV, Nobre HV, Macêdo DS, Oliveira AA, Freitas RM, Vasconcelos SM, et al. Neuroprotective effects of caffeine in the model of 6-hydroxydopamine lesion in rats. *Pharmacology, biochemistry, and behavior.* 2006;84(3):415-9.
684. Frechilla D, Cobreros A, Saldise L, Moratalla R, Insausti R, Luquin MR, et al. Serotonin 5-HT1A receptor expression is selectively enhanced in the striosomal compartment of chronic parkinsonian monkeys. *Synapse.* 2001;39(4):288-96.

685. Jourdain VA, Morin N, Di Paolo T. Dopamine Receptors and Levodopa-Induced Dyskinesia. In: Fox SH, Brotchie JM, editors. *Levodopa-Induced Dyskinesia in Parkinson's Disease*. London: Springer London; 2014. p. 171-97.
686. Aubert I, Guigoni C, Håkansson K, Li Q, Dovero S, Barthe N, et al. Increased D1 dopamine receptor signaling in levodopa-induced dyskinesia. *Annals of Neurology*. 2005;57(1):17-26.
687. Gagnon C, Bedard PJ, Di Paolo T. Effect of chronic treatment of MPTP monkeys with dopamine D-1 and/or D-2 receptor agonists. *Eur J Pharmacol*. 1990;178(1):115-20.
688. Gnanalingham KK, Smith LA, Hunter AJ, Jenner P, Marsden CD. Alterations in striatal and extrastriatal D-1 and D-2 dopamine receptors in the MPTP-treated common marmoset: an autoradiographic study. *Synapse*. 1993;14(2):184-94.
689. Rioux L, Frohna PA, Joyce JN, Schneider JS. The effects of chronic levodopa treatment on pre- and postsynaptic markers of dopaminergic function in striatum of parkinsonian monkeys. *Mov Disord*. 1997;12(2):148-58.
690. Blanchet P, Bedard PJ, Britton DR, Keabian JW. Differential effect of selective D-1 and D-2 dopamine receptor agonists on levodopa-induced dyskinesia in 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine- exposed monkeys. *J Pharmacol Exp Ther*. 1993;267(1):275-9.
691. Goulet M, Madras BK. D(1) dopamine receptor agonists are more effective in alleviating advanced than mild parkinsonism in 1-methyl-4-phenyl-1,2,3, 6-tetrahydropyridine-treated monkeys. *J Pharmacol Exp Ther*. 2000;292(2):714-24.
692. Grondin R, Bedard PJ, Britton DR, Shiosaki K. Potential therapeutic use of the selective dopamine D1 receptor agonist, A-86929: an acute study in parkinsonian levodopa-primed monkeys. *Neurology*. 1997;49(2):421-6.
693. Blanchet PJ, Grondin R, Bedard PJ. Dyskinesia and wearing-off following dopamine D1 agonist treatment in drug-naive 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-lesioned primates. *Mov Disord*. 1996;11(1):91-4.
694. Delfino MA, Stefano AV, Ferrario JE, Taravini IR, Murer MG, Gershanik OS. Behavioral sensitization to different dopamine agonists in a parkinsonian rodent model of drug-induced dyskinesias. *Behav Brain Res*. 2004;152(2):297-306.
695. Westin JE, Vercammen L, Strome EM, Konradi C, Cenci MA. Spatiotemporal Pattern of Striatal ERK1/2 Phosphorylation in a Rat Model of L-DOPA-Induced Dyskinesia and the Role of Dopamine D1 Receptors. *Biological psychiatry*. 2007;62(7):800-10.
696. Darmopil S, Martin AB, De Diego IR, Ares S, Moratalla R. Genetic inactivation of dopamine D1 but not D2 receptors inhibits L-DOPA-induced dyskinesia and histone activation. *Biol Psychiatry*. 2009;66(6):603-13.
697. Playford ED, Brooks DJ. In vivo and in vitro studies of the dopaminergic system in movement disorders. *Cerebrovascular and brain metabolism reviews*. 1992;4(2):144-71.
698. Muriel MP, Orioux G, Hirsch EC. Levodopa but not ropinirole induces an internalization of D1 dopamine receptors in parkinsonian rats. *Mov Disord*. 2002;17(6):1174-9.
699. Muriel MP, Bernard V, Levey AI, Laribi O, Abrous DN, Agid Y, et al. Levodopa induces a cytoplasmic localization of D1 dopamine receptors in striatal neurons in Parkinson's disease. *Ann Neurol*. 1999;46(1):103-11.
700. Fiorentini C, Missale C. Oligomeric assembly of dopamine D1 and glutamate NMDA receptors: molecular mechanisms and functional implications. *Biochem Soc Trans*. 2004;32(Pt 6):1025-8.

701. Fiorentini C, Gardoni F, Spano P, Di Luca M, Missale C. Regulation of dopamine D1 receptor trafficking and desensitization by oligomerization with glutamate N-methyl-D-aspartate receptors. *J Biol Chem.* 2003;278(22):20196-202.
702. Lee FJ, Xue S, Pei L, Vukusic B, Chery N, Wang Y, et al. Dual regulation of NMDA receptor functions by direct protein-protein interactions with the dopamine D1 receptor. *Cell.* 2002;111(2):219-30.
703. Fiorentini C, Rizzetti MC, Busi C, Bontempi S, Collo G, Spano P, et al. Loss of synaptic D1 dopamine/N-methyl-D-aspartate glutamate receptor complexes in L-DOPA-induced dyskinesia in the rat. *Molecular pharmacology.* 2006;69(3):805-12.
704. Krapivinsky G, Krapivinsky L, Manasian Y, Ivanov A, Tyzio R, Pellegrino C, et al. The NMDA receptor is coupled to the ERK pathway by a direct interaction between NR2B and RasGRF1. *Neuron.* 2003;40(4):775-84.
705. Santini E, Sgambato-Faure V, Li Q, Savasta M, Dovero S, Fisone G, et al. Distinct changes in cAMP and extracellular signal-regulated protein kinase signalling in L-DOPA-induced dyskinesia. *PLoS One.* 2010;5(8):e12322.
706. Lindgren HS, Ohlin KE, Cenci MA. Differential involvement of D1 and D2 dopamine receptors in L-DOPA-induced angiogenic activity in a rat model of Parkinson's disease. *Neuropsychopharmacology.* 2009;34(12):2477-88.
707. Schuster S, Nadjar A, Guo JT, Li Q, Ittrich C, Hengerer B, et al. The 3-hydroxy-3-methylglutaryl-CoA reductase inhibitor lovastatin reduces severity of L-DOPA-induced abnormal involuntary movements in experimental Parkinson's disease. *J Neurosci.* 2008;28(17):4311-6.
708. Asin KE, Wirtshafter D, Nikkel A. Amphetamine induces Fos-like immunoreactivity in the striatum of primates. *Brain Res.* 1996;719(1-2):138-42.
709. Graybiel AM, Moratalla R, Robertson HA. Amphetamine and cocaine induce drug-specific activation of the c-fos gene in striosome-matrix compartments and limbic subdivisions of the striatum. *Proc Natl Acad Sci U S A.* 1990;87(17):6912-6.
710. Drake JD, Kibuuka LN, Dimitrov KD, Pollack AE. Abnormal involuntary movement (AIM) expression following D2 dopamine agonist challenge is determined by the nature of prior dopamine receptor stimulation (priming) in 6-hydroxydopamine lesioned rats. *Pharmacol Biochem Behav.* 2013;105:26-33.
711. Dupre KB, Eskow KL, Negron G, Bishop C. The differential effects of 5-HT(1A) receptor stimulation on dopamine receptor-mediated abnormal involuntary movements and rotations in the primed hemiparkinsonian rat. *Brain Res.* 2007;1158:135-43.
712. Fasano S, Bezard E, D'Antoni A, Francardo V, Indrigo M, Qin L, et al. Inhibition of Ras-guanine nucleotide-releasing factor 1 (Ras-GRF1) signaling in the striatum reverts motor symptoms associated with L-dopa-induced dyskinesia. *Proc Natl Acad Sci U S A.* 2010;107(50):21824-9.
713. Samadi P, Gregoire L, Bedard PJ. Opioid antagonists increase the dyskinetic response to dopaminergic agents in parkinsonian monkeys: interaction between dopamine and opioid systems. *Neuropharmacology.* 2003;45(7):954-63.
714. Schapira AH, Barone P, Hauser RA, Mizuno Y, Rascol O, Busse M, et al. Extended-release pramipexole in advanced Parkinson disease: a randomized controlled trial. *Neurology.* 2011;77(8):767-74.

715. Calon F, Goulet M, Blanchet PJ, Martel JC, Piercey MF, Bédard PJ, et al. Levodopa or D2 agonist induced dyskinesia in MPTP monkeys: correlation with changes in dopamine and GABAA_A receptors in the striatopallidal complex. *Brain Research*. 1995;680(1):43-52.
716. Goulet M, Grondin R, Blanchet PJ, Bédard PJ, Di Paolo T. Dyskinesias and tolerance induced by chronic treatment with a D1 agonist administered in pulsatile or continuous mode do not correlate with changes of putaminal D1 receptors in drug-naive MPTP monkeys. *Brain Res*. 1996;719(1-2):129-37.
717. Alexander GM, Brainard DL, Gordon SW, Hichens M, Grothusen JR, Schwartzman RJ. Dopamine receptor changes in untreated and (+)-PHNO-treated MPTP parkinsonian primates. *Brain Res*. 1991;547(2):181-9.
718. Alexander GM, Schwartzman RJ, Grothusen JR, Brainard L, Gordon SW. Changes in brain dopamine receptors in MPTP parkinsonian monkeys following L-dopa treatment. *Brain Res*. 1993;625(2):276-82.
719. Thobois S, Vingerhoets F, Fraix V, Xie-Brustolin J, Mollion H, Costes N, et al. Role of dopaminergic treatment in dopamine receptor down-regulation in advanced Parkinson disease: a positron emission tomographic study. *Arch Neurol*. 2004;61(11):1705-9.
720. Kaasinen V, Ruottinen HM, Nagren K, Lehtikoinen P, Oikonen V, Rinne JO. Upregulation of putaminal dopamine D2 receptors in early Parkinson's disease: a comparative PET study with [¹¹C] raclopride and [¹¹C]N-methylspiperone. *J Nucl Med*. 2000;41(1):65-70.
721. Gagnon C, Gomez-Mancilla B, Markstein R, Bédard PJ, Di Paolo T. Effect of adding the D-1 agonist CY 208-243 to chronic bromocriptine treatment of MPTP-monkeys: regional changes of brain dopamine receptors. *Progress in neuro-psychopharmacology & biological psychiatry*. 1995;19(4):667-76.
722. Gomez-Mancilla B, Boucher R, Gagnon C, Di Paolo T, Markstein R, Bédard PJ. Effect of adding the D1 agonist CY 208-243 to chronic bromocriptine treatment. I: Evaluation of motor parameters in relation to striatal catecholamine content and dopamine receptors. *Mov Disord*. 1993;8(2):144-50.
723. Rinne JO, Laihin A, Rinne UK, Nagren K, Bergman J, Ruotsalainen U. PET study on striatal dopamine D2 receptor changes during the progression of early Parkinson's disease. *Mov Disord*. 1993;8(2):134-8.
724. Antonini A, Schwarz J, Oertel WH, Beer HF, Madeja UD, Leenders KL. [¹¹C]raclopride and positron emission tomography in previously untreated patients with Parkinson's disease: Influence of L-dopa and lisuride therapy on striatal dopamine D2-receptors. *Neurology*. 1994;44(7):1325-9.
725. Morissette M, Goulet M, Calon F, Falardeau P, Blanchet PJ, Bédard PJ, et al. Changes of D1 and D2 dopamine receptor mRNA in the brains of monkeys lesioned with 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine: correction with chronic administration of L-3,4-dihydroxyphenylalanine. *Molecular pharmacology*. 1996;50(5):1073-9.
726. Herrero MT, Augood SJ, Asensi H, Hirsch EC, Agid Y, Obeso JA, et al. Effects of L-DOPA-therapy on dopamine D2 receptor mRNA expression in the striatum of MPTP-intoxicated parkinsonian monkeys. *Brain Res Mol Brain Res*. 1996;42(1):149-55.
727. Goulet M, Morissette M, Calon F, Blanchet PJ, Falardeau P, Bédard PJ, et al. Continuous or pulsatile chronic D2 dopamine receptor agonist (U91356A) treatment of drug-naive 4-phenyl-1,2,3,6-tetrahydropyridine monkeys differentially regulates brain D1 and D2 receptor expression: in situ hybridization histochemical analysis. *Neuroscience*. 1997;79(2):497-507.

728. Twarog BM, Page IH. Serotonin content of some mammalian tissues and urine and a method for its determination. *Am J Physiol.* 1953;175(1):157-61.
729. Bogdanski DF, Pletscher A, Brodie BB, Udenfriend S. Identification and assay of serotonin in brain. *J Pharmacol Exp Ther.* 1956;117(1):82-8.
730. Nichols DE, Nichols CD. Serotonin receptors. *Chem Rev.* 2008;108(5):1614-41.
731. Tronci E, Lisci C, Stancampiano R, Fidalgo C, Collu M, Devoto P, et al. 5-Hydroxytryptophan for the treatment of L-DOPA-induced dyskinesia in the rat Parkinson's disease model. *Neurobiol Dis.* 2013;60:108-14.
732. Jacobs BL, Azmitia EC. Structure and function of the brain serotonin system. *Physiol Rev.* 1992;72(1):165-229.
733. Hornung JP. The human raphe nuclei and the serotonergic system. *J Chem Neuroanat.* 2003;26(4):331-43.
734. Stansley BJ, Yamamoto BK. L-Dopa and Brain Serotonin System Dysfunction. *Toxics.* 2015;3(1):75-88.
735. Descarries L, Audet MA, Doucet GUY, Garcia S, Oleskevich S, SÉGuÉLa P, et al. Morphology of Central Serotonin Neurons. *Annals of the New York Academy of Sciences.* 1990;600(1):81-92.
736. Huot P, Fox SH, Brotchie JM. The serotonergic system in Parkinson's disease. *Prog Neurobiol.* 2011;95(2):163-212.
737. Jacobs BL, Foote SL, Bloom FE. Differential projections of neurons within the dorsal raphe nucleus of the rat: a horseradish peroxidase (HRP) study. *Brain Res.* 1978;147(1):149-53.
738. Lowry CA, Evans AK, Gasser PJ, Hale MW, Staub DR, Shekhar A. Topographic organization and chemoarchitecture of the dorsal raphe nucleus and the median raphe nucleus. In: Monti JM, Pandi-Perumal SR, Jacobs BL, Nutt DJ, editors. *Serotonin and Sleep: Molecular, Functional and Clinical Aspects.* Basel: Birkhäuser Basel; 2008. p. 25-67.
739. Carlsson T, Carta M, Winkler C, Björklund A, Kirik D. Serotonin Neuron Transplants Exacerbate L-DOPA- Induced Dyskinesias in a Rat Model of Parkinson's Disease. *The Journal of Neuroscience.* 2007;27(30):8011-22.
740. Eskow KL, Dupre KB, Barnum CJ, Dickinson SO, Park JY, Bishop C. The role of the dorsal raphe nucleus in the development, expression, and treatment of L-dopa-induced dyskinesia in hemiparkinsonian rats. *Synapse.* 2009;63(7):610-20.
741. Bezard E, Tronci E, Pioli EY, Li Q, Porrás G, Björklund A, et al. Study of the antidyskinetic effect of eltopazine in animal models of levodopa-induced dyskinesia. *Mov Disord.* 2013;28(8):1088-96.
742. Miller DW, Abercrombie ED. Role of high-affinity dopamine uptake and impulse activity in the appearance of extracellular dopamine in striatum after administration of exogenous L-DOPA: studies in intact and 6-hydroxydopamine-treated rats. *J Neurochem.* 1999;72(4):1516-22.
743. Tanaka H, Kannari K, Maeda T, Tomiyama M, Suda T, Matsunaga M. Role of serotonergic neurons in L-DOPA-derived extracellular dopamine in the striatum of 6-OHDA-lesioned rats. *Neuroreport.* 1999;10(3):631-4.
744. Kannari K, Yamato H, Shen H, Tomiyama M, Suda T, Matsunaga M. Activation of 5-HT1A but not 5-HT1B receptors attenuates an increase in extracellular dopamine derived from exogenously administered L-DOPA in the striatum with nigrostriatal denervation. *Journal of Neurochemistry.* 2001;76(5):1346-53.

745. Carta M, Tronci E. Serotonin System Implication in l-DOPA-Induced Dyskinesia: From Animal Models to Clinical Investigations. *Front Neurol.* 2014;5:78.
746. Carta M, Carlsson T, Munoz A, Kirik D, Bjorklund A. Serotonin–dopamine interaction in the induction and maintenance of L-DOPA-induced dyskinesias. *Serotonin–Dopamine Interaction: Experimental Evidence and Therapeutic Relevance. Progress in Brain Research*2008. p. 465-78.
747. Eiden LE, Weihe E. VMAT2: a dynamic regulator of brain monoaminergic neuronal function interacting with drugs of abuse. *Annals of the New York Academy of Sciences.* 2011;1216:86-98.
748. Lopez A, Munoz A, Guerra MJ, Labandeira-Garcia JL. Mechanisms of the effects of exogenous levodopa on the dopamine-denervated striatum. *Neuroscience.* 2001;103(3):639-51.
749. Tronci E, Carta M. 5-HT1 receptor agonists for the treatment of L-DOPA-induced dyskinesia: From animal models to clinical investigation. *Basal Ganglia.* 2013;3(1):9-13.
750. Ulusoy A, Sahin G, Kirik D. Presynaptic dopaminergic compartment determines the susceptibility to L-DOPA–induced dyskinesia in rats. *Proceedings of the National Academy of Sciences.* 2010;107(29):13159-64.
751. Shin E, Garcia J, Winkler C, Bjorklund A, Carta M. Serotonergic and dopaminergic mechanisms in graft-induced dyskinesia in a rat model of Parkinson's disease. *Neurobiol Dis.* 2012;47(3):393-406.
752. de la Fuente-Fernandez R, Schulzer M, Mak E, Calne DB, Stoessl AJ. Presynaptic mechanisms of motor fluctuations in Parkinson's disease: a probabilistic model. *Brain.* 2004;127(Pt 4):888-99.
753. Bishop C, Taylor JL, Kuhn DM, Eskow KL, Park JY, Walker PD. MDMA and fenfluramine reduce L-DOPA-induced dyskinesia via indirect 5-HT1A receptor stimulation. *Eur J Neurosci.* 2006;23(10):2669-76.
754. Muñoz A, Carlsson T, Tronci E, Kirik D, Björklund A, Carta M. Serotonin neuron-dependent and -independent reduction of dyskinesia by 5-HT1A and 5-HT1B receptor agonists in the rat Parkinson model. *Experimental Neurology.* 2009;219(1):298-307.
755. Iderberg H, McCreary AC, Varney MA, Cenci MA, Newman-Tancredi A. Activity of serotonin 5-HT(1A) receptor 'biased agonists' in rat models of Parkinson's disease and L-DOPA-induced dyskinesia. *Neuropharmacology.* 2015;93:52-67.
756. Hashemi P, Dankoski EC, Lama R, Wood KM, Takmakov P, Wightman RM. Brain dopamine and serotonin differ in regulation and its consequences. *Proceedings of the National Academy of Sciences.* 2012;109(29):11510-5.
757. Cenci MA, Konradi C. Maladaptive striatal plasticity in L-DOPA-induced dyskinesia. *Progress in brain research.* 2010;183:209-33.
758. Munoz A, Li Q, Gardoni F, Marcello E, Qin C, Carlsson T, et al. Combined 5-HT1A and 5-HT1B receptor agonists for the treatment of L-DOPA-induced dyskinesia. *Brain.* 2008;131(Pt 12):3380-94.
759. Lindgren HS, Andersson DR, Lagerkvist S, Nissbrandt H, Cenci MA. L-DOPA-induced dopamine efflux in the striatum and the substantia nigra in a rat model of Parkinson's disease: temporal and quantitative relationship to the expression of dyskinesia. *J Neurochem.* 2010;112(6):1465-76.
760. Nahimi A, Høltzermann M, Landau AM, Simonsen M, Jakobsen S, Alstrup AKO, et al. Serotonergic modulation of receptor occupancy in rats treated with l-DOPA after unilateral 6-OHDA lesioning. *Journal of Neurochemistry.* 2012;120(5):806-17.

761. Bibbiani F, Oh JD, Chase TN. Serotonin 5-HT_{1A} agonist improves motor complications in rodent and primate parkinsonian models. *Neurology*. 2001;57(10):1829-34.
762. Bara-Jimenez W, Bibbiani F, Morris MJ, Dimitrova T, Sherzai A, Mouradian MM, et al. Effects of serotonin 5-HT_{1A} agonist in advanced Parkinson's disease. *Mov Disord*. 2005;20(8):932-6.
763. Eskow KL, Gupta V, Alam S, Park JY, Bishop C. The partial 5-HT_{1A} agonist buspirone reduces the expression and development of l-DOPA-induced dyskinesia in rats and improves l-DOPA efficacy. *Pharmacol Biochem Behav*. 2007;87(3):306-14.
764. Goetz CG, Damier P, Hicking C, Laska E, Muller T, Olanow CW, et al. Sarizotan as a treatment for dyskinesias in Parkinson's disease: a double-blind placebo-controlled trial. *Mov Disord*. 2007;22(2):179-86.
765. Bonifati V, Fabrizio E, Cipriani R, Vanacore N, Mecocci G. Buspirone in levodopa-induced dyskinesias. *Clinical neuropharmacology*. 1994;17(1):73-82.
766. Hammerstad JP, Carter J, Nutt JG, Casten GC, Shrotriya RC, Alms DR, et al. Buspirone in Parkinson's disease. *Clinical neuropharmacology*. 1986;9(6):556-60.
767. Kleedorfer B, Lees AJ, Stern GM. Buspirone in the treatment of levodopa induced dyskinesias. *Journal of Neurology, Neurosurgery, and Psychiatry*. 1991;54(4):376-7.
768. Tronci E, Fidalgo C, Carta M. The Serotonergic System in Levodopa-Induced Dyskinesia. In: Fox SH, Brotchie JM, editors. *Levodopa-Induced Dyskinesia in Parkinson's Disease*. London: Springer London; 2014. p. 199-212.
769. Riahi G, Morissette M, Parent M, Di Paolo T. Brain 5-HT_{2A} receptors in MPTP monkeys and levodopa-induced dyskinesias. *Eur J Neurosci*. 2011;33(10):1823-31.
770. Oh JD, Bibbiani F, Chase TN. Quetiapine attenuates levodopa-induced motor complications in rodent and primate parkinsonian models. *Exp Neurol*. 2002;177(2):557-64.
771. Roberts C. ACP-103, a 5-HT_{2A} receptor inverse agonist. *Curr Opin Investig Drugs*. 2006;7(7):653-60.
772. Maertens de Noordhout A, Delwaide PJ. Open pilot trial of ritanserin in parkinsonism. *Clinical neuropharmacology*. 1986;9(5):480-4.
773. Taylor JL, Bishop C, Ullrich T, Rice KC, Walker PD. Serotonin 2A receptor antagonist treatment reduces dopamine D1 receptor-mediated rotational behavior but not l-DOPA-induced abnormal involuntary movements in the unilateral dopamine-depleted rat. *Neuropharmacology*. 2006;50(6):761-8.
774. Olanow CW, Damier P, Goetz CG, Mueller T, Nutt J, Rascol O, et al. Multicenter, open-label, trial of sarizotan in Parkinson disease patients with levodopa-induced dyskinesias (the SPLENDID Study). *Clinical neuropharmacology*. 2004;27(2).
775. Bartoszyk GD, van Amsterdam C, Greiner HE, Rautenberg W, Russ H, Seyfried CA. Sarizotan, a serotonin 5-HT_{1A} receptor agonist and dopamine receptor ligand. 1. Neurochemical profile. *Journal of Neural Transmission*. 2004;111(2):113-26.
776. Svenningsson P, Rosenblad C, Af Edholm Arvidsson K, Wictorin K, Keywood C, Shankar B, et al. Eltoprazine counteracts l-DOPA-induced dyskinesias in Parkinson's disease: a dose-finding study. *Brain*. 2015;138(Pt 4):963-73.
777. Politis M, Wu K, Loane C, Brooks DJ, Kiferle L, Turkheimer FE, et al. Serotonergic mechanisms responsible for levodopa-induced dyskinesias in Parkinson's disease patients. *J Clin Invest*. 2014;124(3):1340-9.

778. Rylander D, Parent M, O'Sullivan SS, Dovero S, Lees AJ, Bezard E, et al. Maladaptive Plasticity of Serotonin Axon Terminals in Levodopa-Induced Dyskinesia. *Ann Neurol*. 2010;68(5):619-28.
779. Roussakis AA, Politis M, Towey D, Piccini P. Serotonin-to-dopamine transporter ratios in Parkinson disease: Relevance for dyskinesias. *Neurology*. 2016;86(12):1152-8.
780. Jackson MJ, Al-Barghouthy G, Pearce RKB, Smith L, Hagan JJ, Jenner P. Effect of 5-HT_{1B/D} receptor agonist and antagonist administration on motor function in haloperidol and MPTP-treated common marmosets. *Pharmacology Biochemistry and Behavior*. 2004;79(3):391-400.
781. Iravani MM, Tayarani-Binazir K, Chu WB, Jackson MJ, Jenner P. In 1-Methyl-4-phenyl-1,2,3,6-tetrahydropyridine-Treated Primates, the Selective 5-Hydroxytryptamine 1a Agonist (R)-(+)-8-OHDPAT Inhibits Levodopa-Induced Dyskinesia but Only with Increased Motor Disability. *Journal of Pharmacology and Experimental Therapeutics*. 2006;319(3):1225-34.
782. Barnes NM, Hales TG, Lummis SC, Peters JA. The 5-HT₃ receptor--the relationship between structure and function. *Neuropharmacology*. 2009;56(1):273-84.
783. Gaddum JH, Picarelli ZP. Two kinds of tryptamine receptor. *Br J Pharmacol Chemother*. 1957;12(3):323-8.
784. Mengod G, Cortés R, Vilaró MT, Hoyer D. CHAPTER 1.6 - Distribution of 5-HT Receptors in the Central Nervous System. In: Müller CP, Jacobs BL, editors. *Handbook of Behavioral Neuroscience*. 21: Elsevier; 2010. p. 123-38.
785. Hoyer D, Neijt HC. Identification of serotonin 5-HT₃ recognition sites by radioligand binding in NG108-15 neuroblastoma-glioma cells. *European Journal of Pharmacology*. 1987;143(2):291-2.
786. Kilpatrick GJ, Jones BJ, Tyers MB. Identification and distribution of 5-HT₃ receptors in rat brain using radioligand binding. *Nature*. 1987;330(6150):746-8.
787. Waeber C, Dixon K, Hoyer D, Palacios JM. Localisation by autoradiography of neuronal 5-HT₃ receptors in the mouse CNS. *European Journal of Pharmacology*. 1988;151(2):351-2.
788. Parker RM, Barnes JM, Ge J, Barber PC, Barnes NM. Autoradiographic distribution of [³H]-(-S)-zacopride-labelled 5-HT₃ receptors in human brain. *J Neurol Sci*. 1996;144(1-2):119-27.
789. Browning KN. Role of central vagal 5-HT(3) receptors in gastrointestinal physiology and pathophysiology. *Frontiers in Neuroscience*. 2015;9:413.
790. Veelken R, Hilgers KF, Leonard M, Scrogin K, Ruhe J, Mann JF, et al. A highly selective cardiorenal serotonergic 5-HT₃-mediated reflex in rats. *American Journal of Physiology - Heart and Circulatory Physiology*. 1993;264(6):H1871-H7.
791. Sévoz-Couche C, Comet M-A, Hamon M, Laguzzi R. Role of Nucleus Tractus Solitarius 5-HT₃ Receptors in the Defense Reaction-Induced Inhibition of the Aortic Baroreflex in Rats. *Journal of Neurophysiology*. 2003;90(4):2521-30.
792. Galligan JJ. Ligand-gated ion channels in the enteric nervous system. *Neurogastroenterology and motility : the official journal of the European Gastrointestinal Motility Society*. 2002;14(6):611-23.
793. Alhaider A, Lei S, Wilcox G. Spinal 5-HT₃ receptor-mediated antinociception: possible release of GABA. *The Journal of Neuroscience*. 1991;11(7):1881-8.

794. Eide PK, Hole K. The role of 5-hydroxytryptamine (5-HT) receptor subtypes and plasticity in the 5-HT systems in the regulation of nociceptive sensitivity. *Cephalalgia : an international journal of headache*. 1993;13(2):75-85.
795. Zeitz KP, Guy N, Malmberg AB, Dirajlal S, Martin WJ, Sun L, et al. The 5-HT₃ Subtype of Serotonin Receptor Contributes to Nociceptive Processing via a Novel Subset of Myelinated and Unmyelinated Nociceptors. *The Journal of Neuroscience*. 2002;22(3):1010-9.
796. Hamon M, Gallissot MC, Menard F, Gozlan H, Bourgoin S, Verge D. 5-HT₃ receptor binding sites are on capsaicin-sensitive fibres in the rat spinal cord. *Eur J Pharmacol*. 1989;164(2):315-22.
797. Kidd EJ, Laporte AM, Langlois X, Fattaccini CM, Doyen C, Lombard MC, et al. 5-HT₃ receptors in the rat central nervous system are mainly located on nerve fibres and terminals. *Brain Res*. 1993;612(1-2):289-98.
798. Morales M, Wang S-D. Differential Composition of 5-Hydroxytryptamine₃ Receptors Synthesized in the Rat CNS and Peripheral Nervous System. *The Journal of Neuroscience*. 2002;22(15):6732-41.
799. Lecrubier Y, Puech AJ, Azcona A, Bailey PE, Lataste X. A randomized double-blind placebo-controlled study of tropisetron in the treatment of outpatients with generalized anxiety disorder. *Psychopharmacology (Berl)*. 1993;112(1):129-33.
800. Ricci LA, Knyshevski I, Melloni RH, Jr. Serotonin type 3 receptors stimulate offensive aggression in Syrian hamsters. *Behav Brain Res*. 2005;156(1):19-29.
801. Hoyer D, Clarke DE, Fozard JR, Hartig PR, Martin GR, Mylecharane EJ, et al. International Union of Pharmacology classification of receptors for 5-hydroxytryptamine (Serotonin). *Pharmacol Rev*. 1994;46(2):157-203.
802. Preston GC. 5-HT₃ antagonists and disorders of cognition. In: Racagni G, Brunello N, Langer SZ, editors. *Recent Advances in the Treatment of Neurodegenerative Disorders and Cognitive Dysfunction*. 7: Int. Acad. Biomed. Drug. Res.; 1994. p. 89-93.
803. Bétry C, Etiévant A, Oosterhof C, Ebert B, Sanchez C, Haddjeri N. Role of 5-HT₃ Receptors in the Antidepressant Response. *Pharmaceuticals*. 2011;4(12):603-29.
804. Morales M, Bloom FE. The 5-HT₃ receptor is present in different subpopulations of GABAergic neurons in the rat telencephalon. *J Neurosci*. 1997;17(9):3157-67.
805. Barnes NM, Sharp T. A review of central 5-HT receptors and their function. *Neuropharmacology*. 1999;38(8):1083-152.
806. Hoyer D, Hannon JP, Martin GR. Molecular, pharmacological and functional diversity of 5-HT receptors. *Pharmacol Biochem Behav*. 2002;71(4):533-54.
807. Pratt GD, Bowery NG, Kilpatrick GJ, Leslie RA, Barnes NM, Naylor RJ, et al. Consensus meeting agrees distribution of 5-HT₃ receptors in mammalian hindbrain. *Trends in pharmacological sciences*. 1990;11(4):135-7.
808. Kilpatrick GJ, Jones BJ, Tyers MB. Binding of the 5-HT₃ ligand, [3H]GR65630, to rat area postrema, vagus nerve and the brains of several species. *European Journal of Pharmacology*. 1989;159(2):157-64.
809. Abi-Dargham A, Laruelle M, Wong DT, Robertson DW, Weinberger DR, Kleinman JE. Pharmacological and regional characterization of [3H]LY278584 binding sites in human brain. *J Neurochem*. 1993;60(2):730-7.

810. Barnes JM, Barnes NM, Costall B, Ironside JW, Naylor RJ. Identification and characterisation of 5-hydroxytryptamine 3 recognition sites in human brain tissue. *J Neurochem.* 1989;53(6):1787-93.
811. Waeber C, Hoyer D, Palacios JM. 5-Hydroxytryptamine₃ receptors in the human brain: Autoradiographic visualization using [3H]ICS 205-930. *Neuroscience.* 1989;31(2):393-400.
812. Barnes JM, Barnes NM, Champaneria S, Costall B, Naylor RJ. Characterisation and autoradiographic localisation of 5-HT₃ receptor recognition sites identified with [3H]-(S)-zacopride in the forebrain of the rat. *Neuropharmacology.* 1990;29(11):1037-45.
813. Laporte AM, Koscielniak T, Ponchant M, Vergé D, Hamon M, Gozlan H. Quantitative autoradiographic mapping of 5-HT₃ receptors in the rat CNS using [125I]iodo-zacopride and [3H]zacopride as radioligands. *Synapse.* 1992;10(4):271-81.
814. Steward LJ, West KE, Kilpatrick GJ, Barnes NM. Labelling of 5-HT₃ receptor recognition sites in the rat brain using the agonist radioligand [3H]meta-chlorophenylbiguanide. *Eur J Pharmacol.* 1993;243(1):13-8.
815. Hewlett WA, Fridman S, Trivedi BL, Schmidt DE, De Paulis T, Ebert MH. Characterization of Desamino-5-[125I]Iodo-3-Methoxy-Zacopride ([125I]MIZAC) binding to 5-HT₃ receptors in the rat brain. *Progress in Neuro-Psychopharmacology and Biological Psychiatry.* 1998;22(2):397-410.
816. Akuzawa S, Miyata K, Fukutomi H. Characterization of [3H]YM060, a potent and selective 5-HT₃ receptor radioligand, in the cerebral cortex of rats. *Eur J Pharmacol.* 1995;281(1):37-42.
817. Carrillo M, Ricci LA, Schwartzer JJ, Melloni RH. Immunohistochemical characterization of 5-HT_{3A} receptors in the Syrian hamster forebrain. *Brain Res.* 2010;1329:67-81.
818. Farber L, Haus U, Spath M, Drechsler S. Physiology and pathophysiology of the 5-HT₃ receptor. *Scand J Rheumatol Suppl.* 2004;119:2-8.
819. Hannon J, Hoyer D. Molecular biology of 5-HT receptors. *Behav Brain Res.* 2008;195(1):198-213.
820. Tecott LH, Maricq AV, Julius D. Nervous system distribution of the serotonin 5-HT₃ receptor mRNA. *Proceedings of the National Academy of Sciences of the United States of America.* 1993;90(4):1430-4.
821. Puig MV, Santana N, Celada P, Mengod G, Artigas F. In vivo excitation of GABA interneurons in the medial prefrontal cortex through 5-HT₃ receptors. *Cereb Cortex.* 2004;14(12):1365-75.
822. Morales M, Battenberg E, de Lecea L, Bloom FE. The type 3 serotonin receptor is expressed in a subpopulation of GABAergic neurons in the rat neocortex and hippocampus. *Brain Res.* 1996;731(1-2):199-202.
823. Morales M, Battenberg E, de Lecea L, Sanna PP, Bloom FE. Cellular and subcellular immunolocalization of the type 3 serotonin receptor in the rat central nervous system. *Brain Res Mol Brain Res.* 1996;36(2):251-60.
824. Hoyer D. Serotonin 5-HT₃, 5-HT₄, and 5-HT-M receptors. *Neuropsychopharmacology.* 1990;3(5-6):371-83.
825. Cederholm JM, Schofield PR, Lewis TM. Gating mechanisms in Cys-loop receptors. *Eur Biophys J.* 2009;39(1):37-49.

826. Maricq AV, Peterson AS, Brake AJ, Myers RM, Julius D. Primary structure and functional expression of the 5HT3 receptor, a serotonin-gated ion channel. *Science*. 1991;254(5030):432-7.
827. Davies PA, Pistis M, Hanna MC, Peters JA, Lambert JJ, Hales TG, et al. The 5-HT3B subunit is a major determinant of serotonin-receptor function. *Nature*. 1999;397(6717):359-63.
828. Niesler B, Frank B, Kapeller J, Rappold GA. Cloning, physical mapping and expression analysis of the human 5-HT3 serotonin receptor-like genes HTR3C, HTR3D and HTR3E. *Gene*. 2003;310:101-11.
829. Karnovsky AM, Gotow LF, McKinley DD, Piechan JL, Ruble CL, Mills CJ, et al. A cluster of novel serotonin receptor 3-like genes on human chromosome 3. *Gene*. 2003;319:137-48.
830. Holbrook JD, Gill CH, Zebda N, Spencer JP, Leyland R, Rance KH, et al. Characterisation of 5-HT3C, 5-HT3D and 5-HT3E receptor subunits: evolution, distribution and function. *J Neurochem*. 2009;108(2):384-96.
831. Niesler B, Walstab J, Combrink S, Moller D, Kapeller J, Rietdorf J, et al. Characterization of the novel human serotonin receptor subunits 5-HT3C, 5-HT3D, and 5-HT3E. *Molecular pharmacology*. 2007;72(1):8-17.
832. Boyd GW, Low P, Dunlop JI, Robertson LA, Vardy A, Lambert JJ, et al. Assembly and cell surface expression of homomeric and heteromeric 5-HT3 receptors: the role of oligomerization and chaperone proteins. *Mol Cell Neurosci*. 2002;21(1):38-50.
833. Thompson AJ, Lummis SC. 5-HT3 receptors. *Curr Pharm Des*. 2006;12(28):3615-30.
834. Sudweeks SN, van Hooft JA, Yakel JL. Serotonin 5-HT(3) receptors in rat CA1 hippocampal interneurons: functional and molecular characterization. *The Journal of Physiology*. 2002;544(Pt 3):715-26.
835. Monk SA, Desai K, Brady CA, Williams JM, Lin L, Princiville A, et al. Generation of a selective 5-HT3B subunit-recognising polyclonal antibody; identification of immunoreactive cells in rat hippocampus. *Neuropharmacology*. 2001;41(8):1013-6.
836. Chameau P, van Hooft JA. Serotonin 5-HT3 receptors in the central nervous system. *Cell and Tissue Research*. 2006;326(2):573-81.
837. Thompson AJ, Lummis SC. The 5-HT3 receptor as a therapeutic target. *Expert Opin Ther Targets*. 2007;11(4):527-40.
838. Noam Y, Wadman WJ, van Hooft JA. On the voltage-dependent Ca²⁺ block of serotonin 5-HT3 receptors: a critical role of intracellular phosphates. *J Physiol*. 2008;586(15):3629-38.
839. Walstab J, Rappold G, Niesler B. 5-HT(3) receptors: role in disease and target of drugs. *Pharmacol Ther*. 2010;128(1):146-69.
840. Stewart A, Davies PA, Kirkness EF, Safa P, Hales TG. Introduction of the 5-HT3B subunit alters the functional properties of 5-HT3 receptors native to neuroblastoma cells. *Neuropharmacology*. 2003;44(2):214-23.
841. van Hooft JA, Yakel JL. 5-HT₃ receptors in the CNS: 3B or not 3B? *Trends in pharmacological sciences*. 2004;24(4):157-60.
842. Jensen AA, Davies PA, Brauner-Osborne H, Krzywkowski K. 3B but which 3B and that's just one of the questions: the heterogeneity of human 5-HT3 receptors. *Trends in pharmacological sciences*. 2008;29(9):437-44.
843. Engel M, Smidt M, Van Hooft J. The serotonin 5-HT3 receptor: a novel neurodevelopmental target. *Frontiers in Cellular Neuroscience*. 2013;7(76).

844. Yang J. Ion permeation through 5-hydroxytryptamine-gated channels in neuroblastoma N18 cells. *The Journal of General Physiology*. 1990;96(6):1177-98.
845. Hargreaves AC, Lummis SC, Taylor CW. Ca²⁺ permeability of cloned and native 5-hydroxytryptamine type 3 receptors. *Molecular pharmacology*. 1994;46(6):1120-8.
846. Miquel MC, Emerit MB, Nosjean A, Simon A, Rumajogee P, Brisorgueil MJ, et al. Differential subcellular localization of the 5-HT₃-As receptor subunit in the rat central nervous system. *Eur J Neurosci*. 2002;15(3):449-57.
847. Huang J, Spier AD, Pickel VM. 5-HT₃A receptor subunits in the rat medial nucleus of the solitary tract: subcellular distribution and relation to the serotonin transporter. *Brain Res*. 2004;1028(2):156-69.
848. Faerber L, Drechsler S, Ladenburger S, Gschaidmeier H, Fischer W. The neuronal 5-HT₃ receptor network after 20 years of research--evolving concepts in management of pain and inflammation. *Eur J Pharmacol*. 2007;560(1):1-8.
849. Greenshaw AJ, Silverstone PH. The non-antiemetic uses of serotonin 5-HT₃ receptor antagonists. *Clinical pharmacology and therapeutic applications*. *Drugs*. 1997;53(1):20-39.
850. Funahashi M, Mitoh Y, Matsuo R. Activation of presynaptic 5-HT₃ receptors facilitates glutamatergic synaptic inputs to area postrema neurons in rat brain slices. *Methods and findings in experimental and clinical pharmacology*. 2004;26(8):615-22.
851. Ronde P, Nichols RA. High calcium permeability of serotonin 5-HT₃ receptors on presynaptic nerve terminals from rat striatum. *J Neurochem*. 1998;70(3):1094-103.
852. Sugita S, Shen KZ, North RA. 5-hydroxytryptamine is a fast excitatory transmitter at 5-HT₃ receptors in rat amygdala. *Neuron*. 1992;8(1):199-203.
853. Roerig B, Katz LC. Modulation of Intrinsic Circuits by Serotonin 5-HT₃ Receptors in Developing Ferret Visual Cortex. *The Journal of Neuroscience*. 1997;17(21):8324-38.
854. Barrera NP, Herbert P, Henderson RM, Martin IL, Edwardson JM. Atomic force microscopy reveals the stoichiometry and subunit arrangement of 5-HT₃ receptors. *Proceedings of the National Academy of Sciences of the United States of America*. 2005;102(35):12595-600.
855. Hapfelmeier G, Tredt C, Haseneder R, Zieglgansberger W, Eisensamer B, Rupprecht R, et al. Co-expression of the 5-HT_{3B} serotonin receptor subunit alters the biophysics of the 5-HT₃ receptor. *Biophysical journal*. 2003;84(3):1720-33.
856. Nichols RA, Mollard P. Direct observation of serotonin 5-HT₃ receptor-induced increases in calcium levels in individual brain nerve terminals. *J Neurochem*. 1996;67(2):581-92.
857. Nayak SV, Ronde P, Spier AD, Lummis SC, Nichols RA. Calcium changes induced by presynaptic 5-hydroxytryptamine-3 serotonin receptors on isolated terminals from various regions of the rat brain. *Neuroscience*. 1999;91(1):107-17.
858. Turner TJ, Mokler DJ, Luebke JI. Calcium influx through presynaptic 5-HT₃ receptors facilitates GABA release in the hippocampus: in vitro slice and synaptosome studies. *Neuroscience*. 2004;129(3):703-18.
859. Yakel JL, Shao XM, Jackson MB. The selectivity of the channel coupled to the 5-HT₃ receptor. *Brain Res*. 1990;533(1):46-52.
860. Wallis DI, North RA. The action of 5-hydroxytryptamine on single neurones of the rabbit superior cervical ganglion. *Neuropharmacology*. 1978;17(12):1023-8.

861. Neijt HC, Vijverberg HP, Van den Bercken J. The dopamine response in mouse neuroblastoma cells is mediated by serotonin 5HT₃ receptors. *Eur J Pharmacol.* 1986;127(3):271-4.
862. Richardson BP, Engel G, Donatsch P, Stadler PA. Identification of serotonin M-receptor subtypes and their specific blockade by a new class of drugs. *Nature.* 1985;316(6024):126-31.
863. Bachy A, Héaulme M, Giudice A, Michaud J-C, Lefevre IA, Souilhac J, et al. SR 57227A: a potent and selective agonist at central and peripheral 5-HT₃ receptors in vitro and in vivo. *European Journal of Pharmacology.* 1993;237(2):299-309.
864. Gozlan H. 5-HT₃ receptors. In: B O, I vW, W S, editors. *Pharmacochemistry Library.* 27: Elsevier; 1997. p. 221-58.
865. Schmidt CJ, Black CK. The putative 5-HT₃ agonist phenylbiguanide induces carrier-mediated release of [3H]dopamine. *European journal of pharmacology.* 1989;167(2):309-10.
866. Glennon RA, Young R, Dukat M. 5-HT₃ agonist 2-methylserotonin as a training drug in discrimination studies. *Pharmacol Biochem Behav.* 1992;41(2):361-4.
867. Smith HS, Cox LR, Smith EJ. 5-HT₃ receptor antagonists for the treatment of nausea/vomiting. *Annals of Palliative Medicine.* 2012;1(2):115-20.
868. Aapro M, Blower P. 5-hydroxytryptamine type-3 receptor antagonists for chemotherapy-induced and radiotherapy-induced nausea and emesis: can we safely reduce the dose of administered agents? *Cancer.* 2005;104(1):1-18.
869. Herrstedt J, Dombernowsky P. Anti-emetic therapy in cancer chemotherapy: current status. *Basic Clin Pharmacol Toxicol.* 2007;101(3):143-50.
870. Gregory RE, Ettinger DS. 5-HT₃ receptor antagonists for the prevention of chemotherapy-induced nausea and vomiting. A comparison of their pharmacology and clinical efficacy. *Drugs.* 1998;55(2):173-89.
871. Van Wijngaarden I, Tulp MTM, Soudijn W. The concept of selectivity in 5-HT receptor research. *European Journal of Pharmacology: Molecular Pharmacology.* 1990;188(6):301-12.
872. Butler A, Hill JM, Ireland SJ, Jordan CC, Tyers MB. Pharmacological properties of GR38032F, a novel antagonist at 5-HT₃ receptors. *Br J Pharmacol.* 1988;94(2):397-412.
873. Klein RL, Sanna E, McQuilkin SJ, Whiting PJ, Harris RA. Effects of 5-HT₃ receptor antagonists on binding and function of mouse and human GABAA receptors. *Eur J Pharmacol.* 1994;268(2):237-46.
874. Cunningham RS. 5-HT₃-receptor antagonists: a review of pharmacology and clinical efficacy. *Oncol Nurs Forum.* 1997;24(7 Suppl):33-40.
875. Ye JH, Schaefer R, Wu WH, Liu PL, Zbuzek VK, McArdle JJ. Inhibitory effect of ondansetron on glycine response of dissociated rat hippocampal neurons. *J Pharmacol Exp Ther.* 1999;290(1):104-11.
876. Mahesh R, Perumal RV, Pandi PV. Cancer chemotherapy-induced nausea and vomiting: role of mediators, development of drugs and treatment methods. *Pharmazie.* 2005;60(2):83-96.
877. Fayyaz M, Lackner JM. Serotonin receptor modulators in the treatment of irritable bowel syndrome. *Therapeutics and Clinical Risk Management.* 2008;4(1):41-8.
878. Clave P. Treatment of IBS-D with 5-HT₃ receptor antagonists vs spasmolytic agents: similar therapeutical effects from heterogeneous pharmacological targets. *Neurogastroenterology and motility : the official journal of the European Gastrointestinal Motility Society.* 2011;23(12):1051-5.
879. Olivier B, van Wijngaarden I, Soudijn W. 5-HT(3) receptor antagonists and anxiety; a preclinical and clinical review. *Eur Neuropsychopharmacol.* 2000;10(2):77-95.

880. Spath M, Welzel D, Farber L. Treatment of chronic fatigue syndrome with 5-HT₃ receptor antagonists--preliminary results. *Scand J Rheumatol Suppl.* 2000;113:72-7.
881. Engleman EA, Rodd ZA, Bell RL, Murphy JM. The Role of 5-HT₃ Receptors in Drug Abuse and as a Target for Pharmacotherapy. *CNS & neurological disorders drug targets.* 2008;7(5):454-67.
882. Spath M. Current experience with 5-HT₃ receptor antagonists in fibromyalgia. *Rheum Dis Clin North Am.* 2002;28(2):319-28.
883. Ferrari MD. 5-HT₃ receptor antagonists and migraine therapy. *J Neurol.* 1991;238 Suppl 1:S53-6.
884. Bhatnagar S, Nowak N, Babich L, Bok L. Deletion of the 5-HT₃ receptor differentially affects behavior of males and females in the Porsolt forced swim and defensive withdrawal tests. *Behav Brain Res.* 2004;153(2):527-35.
885. Lundblad M, Andersson M, Winkler C, Kirik D, Wierup N, Cenci MA. Pharmacological validation of behavioural measures of akinesia and dyskinesia in a rat model of Parkinson's disease. *Eur J Neurosci.* 2002;15(1):120-32.
886. Durif F, Vidailhet M, Assal F, Roche C, Bonnet AM, Agid Y. Low-dose clozapine improves dyskinesias in Parkinson's disease. *Neurology.* 1997;48(3):658-62.
887. Hamadjida A, Nuara SG, Veyres N, Frouni I, Kwan C, Sid-Otmane L, et al. The effect of mirtazapine on dopaminergic psychosis and dyskinesia in the parkinsonian marmoset. *Psychopharmacology (Berl).* 2017;234(6):905-11.
888. Di Paolo T, Grégoire L, Feuerbach D, Elbast W, Weiss M, Gomez-Mancilla B. AQW051, a novel and selective nicotinic acetylcholine receptor $\alpha 7$ partial agonist, reduces l-Dopa-induced dyskinesias and extends the duration of l-Dopa effects in parkinsonian monkeys. *Parkinsonism & Related Disorders.* 2014;20(11):1119-23.
889. Nousiainen S. Current and novel therapies for levodopa-induced dyskinesia [Master's thesis]: University of Helsinki, Finland; 2015.
890. Lieberman J, Johns C, Cooper T, Pollack S, Kane J. Clozapine pharmacology and tardive dyskinesia. *Psychopharmacology.* 1989;99(1):S54-S9.
891. Cappelli A, Anzini M, Vomero S, Mennuni L, Makovec F, Doucet E, et al. Novel Potent and Selective Central 5-HT₃ Receptor Ligands Provided with Different Intrinsic Efficacy. 1. Mapping the Central 5-HT₃ Receptor Binding Site by Arylpiperazine Derivatives. *Journal of Medicinal Chemistry.* 1998;41(5):728-41.
892. Feuerbach D, Pezous N, Weiss M, Shakeri-Nejad K, Lingenhoehl K, Hoyer D, et al. AQW051, a novel, potent and selective $\alpha 7$ nicotinic ACh receptor partial agonist: pharmacological characterization and phase I evaluation. *British Journal of Pharmacology.* 2015;172(5):1292-304.
893. Sydserff S, Sutton EJ, Song D, Quirk MC, Maciag C, Li C, et al. Selective $\alpha 7$ nicotinic receptor activation by AZD0328 enhances cortical dopamine release and improves learning and attentional processes. *Biochemical pharmacology.* 2009;78(7):880-8.
894. Ashby CR, Jr., Wang RY. Pharmacological actions of the atypical antipsychotic drug clozapine: a review. *Synapse.* 1996;24(4):349-94.
895. Anttila SA, Leinonen EV. A review of the pharmacological and clinical profile of mirtazapine. *CNS drug reviews.* 2001;7(3):249-64.
896. Bymaster FP, Calligaro DO, Falcone JF, Marsh RD, Moore NA, Tye NC, et al. Radioreceptor binding profile of the atypical antipsychotic olanzapine. *Neuropsychopharmacology.* 1996;14(2):87-96.

897. Koyama Y, Kondo M, Shimada S. Building a 5-HT_{3A} Receptor Expression Map in the Mouse Brain. *Scientific Reports*. 2017;7:42884.
898. Gehlert DR, Schober DA, Gackenhaimer SL, Mais DE, Ladouceur G, Robertson DW. Synthesis and evaluation of [¹²⁵I]-(S)-iodozacopride, a high affinity radioligand for 5HT₃ receptors. *Neurochemistry international*. 1993;23(4):373-83.
899. Gehlert DR, Gackenhaimer SL, Wong DT, Robertson DW. Localization of 5-HT₃ receptors in the rat brain using [³H]LY278584. *Brain Research*. 1991;553(1):149-54.
900. Bufton KE, Steward LJ, Barber PC, Barnes NM. Distribution and characterization of the [³H]granisetron-labelled 5-HT₃ receptor in the human forebrain. *Neuropharmacology*. 1993;32(12):1325-31.
901. Morales M, Battenberg E, Bloom FE. Distribution of neurons expressing immunoreactivity for the 5HT₃ receptor subtype in the rat brain and spinal cord. *The Journal of comparative neurology*. 1998;402(3):385-401.
902. Fletcher S, Barnes NM. Autoradiographic localization of the [³H]-(S)-zacopride labelled 5-HT₃ receptor in porcine brain. *Neurosci Lett*. 1999;269(2):91-4.
903. Marazziti D, Betti L, Giannaccini G, Rossi A, Masala I, Baroni S, et al. Distribution of [³H]GR65630 binding in human brain postmortem. *Neurochem Res*. 2001;26(3):187-90.
904. Steward LJ, Bufton KE, Hopkins PC, Davies WE, Barnes NM. Reduced levels of 5-HT₃ receptor recognition sites in the putamen of patients with Huntington's disease. *Eur J Pharmacol*. 1993;242(2):137-43.
905. Doucet E, Miquel MC, Nosjean A, Verge D, Hamon M, Emerit MB. Immunolabeling of the rat central nervous system with antibodies partially selective of the short form of the 5-HT₃ receptor. *Neuroscience*. 2000;95(3):881-92.
906. Blandina P, Goldfarb J, Green JP. Activation of a 5-HT₃ receptor releases dopamine from rat striatal slice. *Eur J Pharmacol*. 1988;155(3):349-50.
907. Yi SJ, Gifford AN, Johnson KM. Effect of cocaine and 5-HT₃ receptor antagonists on 5-HT-induced [³H]dopamine release from rat striatal synaptosomes. *Eur J Pharmacol*. 1991;199(2):185-9.
908. Benuck M, Reith ME. Dopamine releasing effect of phenylbiguanide in rat striatal slices. *Naunyn-Schmiedeberg's archives of pharmacology*. 1992;345(6):666-72.
909. Jacocks HM, 3rd, Cox BM. Serotonin-stimulated release of [³H]dopamine via reversal of the dopamine transporter in rat striatum and nucleus accumbens: a comparison with release elicited by potassium, N-methyl-D-aspartic acid, glutamic acid and D-amphetamine. *J Pharmacol Exp Ther*. 1992;262(1):356-64.
910. Santiago M, Machado A, Cano J. 5-HT₃ receptor agonist induced carrier-mediated release of dopamine in rat striatum in vivo. *British Journal of Pharmacology*. 1995;116(1):1545-50.
911. Jiang LH, Ashby CR, Jr., Kasser RJ, Wang RY. The effect of intraventricular administration of the 5-HT₃ receptor agonist 2-methylserotonin on the release of dopamine in the nucleus accumbens: an in vivo chronocoulometric study. *Brain Res*. 1990;513(1):156-60.
912. Chen JP, van Praag HM, Gardner EL. Activation of 5-HT₃ receptor by 1-phenylbiguanide increases dopamine release in the rat nucleus accumbens. *Brain Res*. 1991;543(2):354-7.
913. Palfreyman MG, Schmidt CJ, Sorensen SM, Dudley MW, Kehne JH, Moser P, et al. Electrophysiological, biochemical and behavioral evidence for 5-HT₂ and 5-HT₃ mediated control of dopaminergic function. *Psychopharmacology (Berl)*. 1993;112(1 Suppl):S60-7.

914. Shankar RP, Karan RS, Handu SS, Bhargava VK. Effect of the 5-HT₃ receptor antagonist ondansetron on amphetamine-induced hyperactivity and stereotypy in rats. *Indian journal of physiology and pharmacology*. 2000;44(3):355-8.
915. Naidu PS, Kulkarni SK. Reversal of neuroleptic-induced orofacial dyskinesia by 5-HT₃ receptor antagonists. *Eur J Pharmacol*. 2001;420(2-3):113-7.
916. Marty M, Pouillart P, Scholl S, Droz JP, Azab M, Brion N, et al. Comparison of the 5-Hydroxytryptamine₃ (Serotonin) Antagonist Ondansetron (Gr 38032F) with High-Dose Metoclopramide in the Control of Cisplatin-Induced Emesis. *New England Journal of Medicine*. 1990;322(12):816-21.
917. Koulou M, Lappalainen J, Hietala J, Sjöholm B. Effects of chronic administration of ondansetron (GR38032F), a selective 5-HT₃ receptor antagonist, on monoamine metabolism in mesolimbic and nigrostriatal dopaminergic neurons and on striatal D₂-receptor binding. *Psychopharmacology (Berl)*. 1990;101(2):168-71.
918. Costall B, Domeney AM, Naylor RJ, Tyers MB. Effects of the 5-HT₃ receptor antagonist, GR38032F, on raised dopaminergic activity in the mesolimbic system of the rat and marmoset brain. *Br J Pharmacol*. 1987;92(4):881-94.
919. Hagan RM, Butler A, Hill JM, Jordan CC, Ireland SJ, Tyers MB. Effect of the 5-HT₃ receptor antagonist, GR38032F, on responses to injection of a neurokinin agonist into the ventral tegmental area of the rat brain. *Eur J Pharmacol*. 1987;138(2):303-5.
920. Sirota P, Mosheva T, Shabtay H, Giladi N, Korczyn AD. Use of the selective serotonin 3 receptor antagonist ondansetron in the treatment of neuroleptic-induced tardive dyskinesia. *The American journal of psychiatry*. 2000;157(2):287-9.
921. Zoldan J, Friedberg G, Goldberg-Stern H, Melamed E. Ondansetron for hallucinosis in advanced Parkinson's disease. *The Lancet*. 1993;341(8844):562-3.
922. Zoldan J, Friedberg G, Livneh M, Melamed E. Psychosis in advanced Parkinson's disease: treatment with ondansetron, a 5-HT₃ receptor antagonist. *Neurology*. 1995;45(7):1305-8.
923. Blesa J, Pifl C, Sanchez-Gonzalez MA, Juri C, Garcia-Cabezas MA, Adanez R, et al. The nigrostriatal system in the presymptomatic and symptomatic stages in the MPTP monkey model: a PET, histological and biochemical study. *Neurobiol Dis*. 2012;48(1):79-91.
924. Blesa J, Przedborski S. Parkinson's disease: animal models and dopaminergic cell vulnerability. *Frontiers in Neuroanatomy*. 2014;8:155.
925. Porrás G, Li Q, Bezard E. Modeling Parkinson's disease in primates: The MPTP model. *Cold Spring Harb Perspect Med*. 2012;2(3):a009308.
926. Elliott PJ, Close SP, Walsh DM, Hayes AG, Marriott AS. Neuroleptic-induced catalepsy as a model of Parkinson's disease. I. Effect of dopaminergic agents. *J Neural Transm Park Dis Dement Sect*. 1990;2(2):79-89.
927. Konieczny J, Przegalinski E, Pokorski M. N-oleoyl-dopamine decreases muscle rigidity induced by reserpine in rats. *Int J Immunopathol Pharmacol*. 2009;22(1):21-8.
928. Duty S, Jenner P. Animal models of Parkinson's disease: a source of novel treatments and clues to the cause of the disease. *British Journal of Pharmacology*. 2011;164(4):1357-91.
929. Fox SH, Brotchie JM. The MPTP-lesioned non-human primate models of Parkinson's disease. Past, present, and future. *Recent Advances in Parkinson's Disease - Translational and Clinical Research*. Progress in Brain Research 2010. p. 133-57.
930. Meredith GE, Sonsalla PK, Chesselet MF. Animal models of Parkinson's disease progression. *Acta Neuropathol*. 2008;115(4):385-98.

931. Ungerstedt U. Postsynaptic supersensitivity after 6-hydroxy-dopamine induced degeneration of the nigro-striatal dopamine system. *Acta physiologica Scandinavica Supplementum*. 1971;367:69-93.
932. Dawson TM, Ko HS, Dawson VL. Genetic Animal Models of Parkinson's Disease. *Neuron*. 2010;66(5):646-61.
933. Sacino AN, Brooks M, McKinney AB, Thomas MA, Shaw G, Golde TE, et al. Brain Injection of α -Synuclein Induces Multiple Proteinopathies, Gliosis, and a Neuronal Injury Marker. *The Journal of Neuroscience*. 2014;34(37):12368-78.
934. Thakur P, Breger LS, Lundblad M, Wan OW, Mattsson B, Luk KC, et al. Modeling Parkinson's disease pathology by combination of fibril seeds and α -synuclein overexpression in the rat brain. *Proceedings of the National Academy of Sciences*. 2017;114(39):E8284-E93.
935. Niu Y, Guo X, Chen Y, Wang C-E, Gao J, Yang W, et al. Early Parkinson's disease symptoms in α -synuclein transgenic monkeys. *Human molecular genetics*. 2015;24(8):2308-17.
936. Visanji NP, Brotchie JM, Kalia LV, Koprach JB, Tandon A, Watts JC, et al. α -Synuclein-Based Animal Models of Parkinson's Disease: Challenges and Opportunities in a New Era. *TINS Trends in Neurosciences*. 2016;39(11):750-62.
937. Goldberg MS, Pisani A, Haburcak M, Vortherms TA, Kitada T, Costa C, et al. Nigrostriatal dopaminergic deficits and hypokinesia caused by inactivation of the familial Parkinsonism-linked gene DJ-1. *Neuron*. 2005;45(4):489-96.
938. Andres-Mateos E, Perier C, Zhang L, Blanchard-Fillion B, Greco TM, Thomas B, et al. DJ-1 gene deletion reveals that DJ-1 is an atypical peroxiredoxin-like peroxidase. *Proc Natl Acad Sci U S A*. 2007;104(37):14807-12.
939. Hinkle KM, Yue M, Behrouz B, Dachselt JC, Lincoln SJ, Bowles EE, et al. LRRK2 knockout mice have an intact dopaminergic system but display alterations in exploratory and motor co-ordination behaviors. *Molecular neurodegeneration*. 2012;7:25.
940. Sanchez G, Varaschin RK, Bueler H, Marcogliese PC, Park DS, Trudeau LE. Unaltered striatal dopamine release levels in young Parkin knockout, Pink1 knockout, DJ-1 knockout and LRRK2 R1441G transgenic mice. *PLoS One*. 2014;9(4):e94826.
941. Rousseaux MW, Marcogliese PC, Qu D, Hewitt SJ, Seang S, Kim RH, et al. Progressive dopaminergic cell loss with unilateral-to-bilateral progression in a genetic model of Parkinson disease. *Proc Natl Acad Sci U S A*. 2012;109(39):15918-23.
942. Hennis MR, Seamans KW, Marvin MA, Casey BH, Goldberg MS. Behavioral and Neurotransmitter Abnormalities in Mice Deficient for Parkin, DJ-1 and Superoxide Dismutase. *PLoS ONE*. 2013;8(12):e84894.
943. Torres EM, Dunnett SB. 6-OHDA Lesion Models of Parkinson's Disease in the Rat. In: Lane EL, Dunnett SB, editors. *Animal Models of Movement Disorders: Volume I*. Totowa, NJ: Humana Press; 2012. p. 267-79.
944. Deumens R, Blokland A, Prickaerts J. Modeling Parkinson's disease in rats: an evaluation of 6-OHDA lesions of the nigrostriatal pathway. *Exp Neurol*. 2002;175(2):303-17.
945. Glinka Y, Gassen M, Youdim MB. Mechanism of 6-hydroxydopamine neurotoxicity. *Journal of neural transmission Supplementum*. 1997;50:55-66.
946. Iderberg H, Francardo V, Pioli EY. Animal models of L-DOPA-induced dyskinesia: an update on the current options. *Neuroscience*. 2012;211:13-27.
947. Blum D, Torch S, Lambeng N, Nissou M, Benabid AL, Sadoul R, et al. Molecular pathways involved in the neurotoxicity of 6-OHDA, dopamine and MPTP: contribution to the apoptotic theory in Parkinson's disease. *Prog Neurobiol*. 2001;65(2):135-72.

948. Marin C, Rodriguez-Oroz MC, Obeso JA. Motor complications in Parkinson's disease and the clinical significance of rotational behavior in the rat: have we wasted our time? *Exp Neurol*. 2006;197(2):269-74.
949. Ungerstedt U. 6-Hydroxy-dopamine induced degeneration of central monoamine neurons. *Eur J Pharmacol*. 1968;5(1):107-10.
950. Winkler C, Kirik D, Björklund A, Cenci MA. 1-DOPA-Induced Dyskinesia in the Intrastratial 6-Hydroxydopamine Model of Parkinson's Disease: Relation to Motor and Cellular Parameters of Nigrostriatal Function. *Neurobiology of Disease*. 2002;10(2):165-86.
951. Blandini F, Armentero MT, Martignoni E. The 6-hydroxydopamine model: news from the past. *Parkinsonism Relat Disord*. 2008;14 Suppl 2:S124-9.
952. Sauer H, Oertel WH. Progressive degeneration of nigrostriatal dopamine neurons following intrastratial terminal lesions with 6-hydroxydopamine: a combined retrograde tracing and immunocytochemical study in the rat. *Neuroscience*. 1994;59(2):401-15.
953. Lee CS, Sauer H, Bjorklund A. Dopaminergic neuronal degeneration and motor impairments following axon terminal lesion by intrastratial 6-hydroxydopamine in the rat. *Neuroscience*. 1996;72(3):641-53.
954. Ma Y, Zhan M, OuYang L, Li Y, Chen S, Wu J, et al. The effects of unilateral 6-OHDA lesion in medial forebrain bundle on the motor, cognitive dysfunctions and vulnerability of different striatal interneuron types in rats. *Behav Brain Res*. 2014;266:37-45.
955. Moore AE, Cicchetti F, Hennen J, Isacson O. Parkinsonian motor deficits are reflected by proportional A9/A10 dopamine neuron degeneration in the rat. *Exp Neurol*. 2001;172(2):363-76.
956. Perese DA, Ulman J, Viola J, Ewing SE, Bankiewicz KS. A 6-hydroxydopamine-induced selective parkinsonian rat model. *Brain Res*. 1989;494(2):285-93.
957. Kirik D, Rosenblad C, Bjorklund A. Characterization of behavioral and neurodegenerative changes following partial lesions of the nigrostriatal dopamine system induced by intrastratial 6-hydroxydopamine in the rat. *Exp Neurol*. 1998;152(2):259-77.
958. Ungerstedt U. Adipsia and aphagia after 6-hydroxydopamine induced degeneration of the nigro-striatal dopamine system. *Acta physiologica Scandinavica Supplementum*. 1971;367:95-122.
959. Hefti F, Melamed E, Wurtman RJ. Partial lesions of the dopaminergic nigrostriatal system in rat brain: biochemical characterization. *Brain Res*. 1980;195(1):123-37.
960. Castaneda E, Whishaw I, Robinson T. Changes in striatal dopamine neurotransmission assessed with microdialysis following recovery from a bilateral 6-OHDA lesion: variation as a function of lesion size. *The Journal of Neuroscience*. 1990;10(6):1847-54.
961. Zigmond MJ, Abercrombie ED, Berger TW, Grace AA, Stricker EM. Compensations after lesions of central dopaminergic neurons: some clinical and basic implications. *Trends in neurosciences*. 1990;13(7):290-6.
962. Barneoud P, Parmentier S, Mazadier M, Miquet JM, Boireau A, Dubedat P, et al. Effects of complete and partial lesions of the dopaminergic mesotelencephalic system on skilled forelimb use in the rat. *Neuroscience*. 1995;67(4):837-48.
963. Blanchard V, Anglade P, Dziewczapolski G, Savasta M, Agid Y, Raisman-Vozari R. Dopaminergic sprouting in the rat striatum after partial lesion of the substantia nigra. *Brain Res*. 1996;709(2):319-25.
964. Fox SH, Chuang R, Brotchie JM. Serotonin and Parkinson's disease: On movement, mood, and madness. *Mov Disord*. 2009;24(9):1255-66.

965. Cenci MA, Lundblad M. CHAPTER B7 - Utility of 6-Hydroxydopamine Lesioned Rats in the Preclinical Screening of Novel Treatments for Parkinson Disease A2 - LeDoux, Mark. *Animal Models of Movement Disorders*. Burlington: Academic Press; 2005. p. 193-208.
966. Schallert T, Fleming SM, Leasure JL, Tillerson JL, Bland ST. CNS plasticity and assessment of forelimb sensorimotor outcome in unilateral rat models of stroke, cortical ablation, parkinsonism and spinal cord injury. *Neuropharmacology*. 2000;39(5):777-87.
967. Huot P, Johnston TH, Koprach JB, Espinosa MC, Reyes MG, Fox SH, et al. L-745,870 reduces the expression of abnormal involuntary movements in the 6-OHDA-lesioned rat. *Behav Pharmacol*. 2015;26(1-2):101-8.
968. Lane EL, Cheetham SC, Jenner P. Does contraversive circling in the 6-OHDA-lesioned rat indicate an ability to induce motor complications as well as therapeutic effects in Parkinson's disease? *Experimental Neurology*. 2006;197(2):284-90.
969. Siever L, Cohen R, Pert A. Assessing pharmacologically induced dopamine receptor sensitivity changes with the Ungerstedt turning model. *Psychopharmacology (Berl)*. 1981;75(2):212-3.
970. Papa SM, Engber TM, Kask AM, Chase TN. Motor fluctuations in levodopa treated parkinsonian rats: relation to lesion extent and treatment duration. *Brain Res*. 1994;662(1-2):69-74.
971. Henry B, Crossman AR, Brotchie JM. Characterization of enhanced behavioral responses to L-DOPA following repeated administration in the 6-hydroxydopamine-lesioned rat model of Parkinson's disease. *Experimental Neurology*. 1998;151(2):334-42.
972. Stefanova N, Lundblad M, Tison F, Poewe W, Cenci MA, Wenning GK. Effects of pulsatile L-DOPA treatment in the double lesion rat model of striatonigral degeneration (multiple system atrophy). *Neurobiol Dis*. 2004;15(3):630-9.
973. Dekundy A, Lundblad M, Danysz W, Cenci MA. Modulation of L-DOPA-induced abnormal involuntary movements by clinically tested compounds: further validation of the rat dyskinesia model. *Behav Brain Res*. 2007;179(1):76-89.
974. Andersson M, Konradi C, Cenci MA. cAMP response element-binding protein is required for dopamine-dependent gene expression in the intact but not the dopamine-denervated striatum. *J Neurosci*. 2001;21(24):9930-43.
975. Lundblad M, Usiello A, Carta M, Hakansson K, Fisone G, Cenci MA. Pharmacological validation of a mouse model of l-DOPA-induced dyskinesia. *Exp Neurol*. 2005;194(1):66-75.
976. Brotchie JM, Henry B, Hille CJ, Crossman AR. Opioid peptide precursor expression in animal models of dystonia secondary to dopamine-replacement therapy in Parkinson's disease. *Adv Neurol*. 1998;78:41-52.
977. Johansson PA, Andersson M, Andersson KE, Cenci MA. Alterations in cortical and basal ganglia levels of opioid receptor binding in a rat model of l-DOPA-induced dyskinesia. *Neurobiol Dis*. 2001;8(2):220-39.
978. Rylander D, Iderberg H, Li Q, Dekundy A, Zhang J, Li H, et al. A mGluR5 antagonist under clinical development improves L-DOPA-induced dyskinesia in parkinsonian rats and monkeys. *Neurobiol Dis*. 2010;39(3):352-61.
979. Ye JH, Ponnudurai R, Schaefer R. Ondansetron: a selective 5-HT(3) receptor antagonist and its applications in CNS-related disorders. *CNS drug reviews*. 2001;7(2):199-213.
980. Gaudette F, Hamadjida A, Bédard D, Nuara SG, Beaudry F, Huot P. Development and validation of a high-performance liquid chromatography-tandem mass spectrometry method to

- quantify LY-354,740 in rat and marmoset plasma. *Journal of Chromatography B*. 2017;1061-1062(Supplement C):392-8.
981. Paxinos G, Watson C. *The rat brain in stereotaxic coordinates*. Amsterdam; Boston: Elsevier; 2007.
982. Gage GJ, Kipke DR, Shain W. Whole Animal Perfusion Fixation for Rodents. *Journal of Visualized Experiments : JoVE*. 2012(65):3564.
983. Tareke E, Bowyer JF, Doerge DR. Quantification of rat brain neurotransmitters and metabolites using liquid chromatography/electrospray tandem mass spectrometry and comparison with liquid chromatography/electrochemical detection. *Rapid communications in mass spectrometry : RCM*. 2007;21(23):3898-904.
984. Palkovits M. Microdissection in Combination with Biochemical Microassays as a Tool in Tract Tracing. In: Heimer L, Záborszky L, editors. *Neuroanatomical Tract-Tracing Methods 2: Recent Progress*. Boston, MA: Springer US; 1989. p. 299-310.
985. Ling ZD, Collier TJ, Sortwell CE, Lipton JW, Vu TQ, Robie HC, et al. Striatal trophic activity is reduced in the aged rat brain. *Brain Research*. 2000;856(1):301-9.
986. Huot P, Johnston TH, Koprach JB, Fox SH, Brotchie JM. L-DOPA pharmacokinetics in the MPTP-lesioned macaque model of Parkinson's disease. *Neuropharmacology*. 2012;63(5):829-36.
987. Lakens D. Calculating and reporting effect sizes to facilitate cumulative science: a practical primer for t-tests and ANOVAs. *Frontiers in Psychology*. 2013;4:863.
988. Bland JM, Altman DG. Transforming data. *BMJ (Clinical research ed)*. 1996;312(7033):770.
989. Rylander D, Strome E, Mela F, Mercanti G, Cenci MA. The severity of L-DOPA-induced dyskinesia in the rat is positively correlated with the density of striatal serotonin afferents. *Parkinsonism & Related Disorders*. 2007;13:S90.
990. Rylander D, Parent M, O'Sullivan SS, Dovero S, Lees AJ, Bezard E, et al. Maladaptive plasticity of serotonin axon terminals in levodopa-induced dyskinesia. *Ann Neurol*. 2010;68(5):619-28.
991. Gagnon D, Gregoire L, Di Paolo T, Parent M. Serotonin hyperinnervation of the striatum with high synaptic incidence in parkinsonian monkeys. *Brain Struct Funct*. 2016;221(7):3675-91.
992. Roila F, Del Favero A. Ondansetron clinical pharmacokinetics. *Clin Pharmacokinet*. 1995;29(2):95-109.
993. Simpson KH, Hicks FM. Clinical pharmacokinetics of ondansetron. A review. *The Journal of pharmacy and pharmacology*. 1996;48(8):774-81.
994. Steece-Collier K, Collier TJ, Danielson PD, Kurlan R, Yurek DM, Sladek JR, Jr. Embryonic mesencephalic grafts increase levodopa-induced forelimb hyperkinesia in parkinsonian rats. *Mov Disord*. 2003;18(12):1442-54.
995. Goudie AJ, Leathley MJ. Effects of the 5-HT₃ antagonist GR38032F (ondansetron) on benzodiazepine withdrawal in rats. *Eur J Pharmacol*. 1990;185(2-3):179-86.
996. Martin P, Gozlan H, Puech AJ. 5-HT₃ receptor antagonists reverse helpless behaviour in rats. *Eur J Pharmacol*. 1992;212(1):73-8.
997. Ramamoorthy R, Radhakrishnan M, Borah M. Antidepressant-like effects of serotonin type-3 antagonist, ondansetron: an investigation in behaviour-based rodent models. *Behav Pharmacol*. 2008;19(1):29-40.

998. King GR, Xiong Z, Ellinwood EH, Jr. Blockade of cocaine sensitization and tolerance by the co-administration of ondansetron, a 5-HT₃ receptor antagonist, and cocaine. *Psychopharmacology (Berl)*. 1997;130(2):159-65.
999. Bonhaus DW, Stefanich E, Loury DN, Hsu SAO, Eglen RM, Wong EHF. Allosteric Interactions Among Agonists and Antagonists at 5-Hydroxytryptamine₃ Receptors. *Journal of Neurochemistry*. 1995;65(1):104-10.
1000. Eisensamer B, Rammes G, Gimpl G, Shapa M, Ferrari U, Hapfelmeier G, et al. Antidepressants are functional antagonists at the serotonin type 3 (5-HT₃) receptor. *Molecular Psychiatry*. 2003;8:994.
1001. Andrews P. 20 - THE ROLE OF FUNCTIONAL GROUPS IN DRUG-RECEPTOR INTERACTIONS A2 - Wermuth, Camille G. *The Practice of Medicinal Chemistry (Second Edition)*. London: Academic Press; 2003. p. 327-39.
1002. van Wijngaarden I, Tulp MT, Soudijn W. The concept of selectivity in 5-HT receptor research. *Eur J Pharmacol*. 1990;188(6):301-12.
1003. Tepper JM, Tecuapetla F, Koós T, Ibáñez-Sandoval O. Heterogeneity and Diversity of Striatal GABAergic Interneurons. *Frontiers in Neuroanatomy*. 2010;4:150.
1004. Gittis AH, Nelson AB, Thwin MT, Palop JJ, Kreitzer AC. Distinct roles of GABAergic interneurons in the regulation of striatal output pathways. *J Neurosci*. 2010;30(6):2223-34.
1005. Kubota Y, Kawaguchi Y. Dependence of GABAergic synaptic areas on the interneuron type and target size. *J Neurosci*. 2000;20(1):375-86.
1006. Dehorter N, Guigoni C, Lopez C, Hirsch J, Eusebio A, Ben-Ari Y, et al. Dopamine-deprived striatal GABAergic interneurons burst and generate repetitive gigantic IPSCs in medium spiny neurons. *J Neurosci*. 2009;29(24):7776-87.
1007. Steinert JR, Kopp-Scheinflug C, Baker C, Challiss RA, Mistry R, Haustein MD, et al. Nitric oxide is a volume transmitter regulating postsynaptic excitability at a glutamatergic synapse. *Neuron*. 2008;60(4):642-56.
1008. West AR, Galloway MP, Grace AA. Regulation of striatal dopamine neurotransmission by nitric oxide: effector pathways and signaling mechanisms. *Synapse*. 2002;44(4):227-45.
1009. Galarraga E, Vilchis C, Tkatch T, Salgado H, Tecuapetla F, Perez-Rosello T, et al. Somatostatinergic modulation of firing pattern and calcium-activated potassium currents in medium spiny neostriatal neurons. *Neuroscience*. 2007;146(2):537-54.
1010. Lopez-Huerta VG, Tecuapetla F, Guzman JN, Vargas J, Galarraga E. Presynaptic modulation by somatostatin in the neostriatum. *Neurochem Res*. 2008;33(8):1452-8.
1011. Kang UJ, Park DH, Wessel T, Baker H, Joh TH. Dopa-decarboxylation in the striata of rats with unilateral substantia nigra lesions. *Neurosci Lett*. 1992;147(1):53-7.
1012. Nakamura K, Ahmed M, Barr E, Leiden JM, Kang UJ. The localization and functional contribution of striatal aromatic L-amino acid decarboxylase to L-3,4-dihydroxyphenylalanine decarboxylation in rodent parkinsonian models. *Cell transplantation*. 2000;9(5):567-76.
1013. Tashiro Y, Kaneko T, Sugimoto T, Nagatsu I, Kikuchi H, Mizuno N. Striatal neurons with aromatic L-amino acid decarboxylase-like immunoreactivity in the rat. *Neurosci Lett*. 1989;100(1-3):29-34.
1014. Lopez-Real A, Rodriguez-Pallares J, Guerra MJ, Labandeira-Garcia JL. Localization and functional significance of striatal neurons immunoreactive to aromatic L-amino acid decarboxylase or tyrosine hydroxylase in rat Parkinsonian models. *Brain Res*. 2003;969(1-2):135-46.

1015. Blandina P, Goldfarb J, Green JP. Activation of a 5-HT₃ receptor releases dopamine from rat striatal slice. *European Journal of Pharmacology*. 1988;155(3):349-50.
1016. Benloucif S, Galloway MP. Facilitation of dopamine release in vivo by serotonin agonists: studies with microdialysis. *Eur J Pharmacol*. 1991;200(1):1-8.
1017. Benloucif S, Keegan MJ, Galloway MP. Serotonin-facilitated dopamine release in vivo: pharmacological characterization. *J Pharmacol Exp Ther*. 1993;265(1):373-7.
1018. Porrás G, De Deurwaerdere P, Moison D, Spampinato U. Conditional involvement of striatal serotonin₃ receptors in the control of in vivo dopamine outflow in the rat striatum. *Eur J Neurosci*. 2003;17(4):771-81.
1019. Invernizzi R, Pozzi L, Samanin R. Selective reduction of extracellular dopamine in the rat nucleus accumbens following chronic treatment with DAU 6215, a 5-HT₃ receptor antagonist. *Neuropharmacology*. 1995;34(2):211-5.
1020. Di Matteo V, Di Giovanni G, Pierucci M, Esposito E. Serotonin control of central dopaminergic function: focus on in vivo microdialysis studies. In: Di Giovanni G, Di Matteo V, Esposito E, editors. *Progress in Brain Research*. 172: Elsevier; 2008. p. 7-44.
1021. Di Matteo V, Pierucci M, Esposito E, Crescimanno G, Benigno A, Di Giovanni G. Serotonin modulation of the basal ganglia circuitry: therapeutic implication for Parkinson's disease and other motor disorders. *Prog Brain Res*. 2008;172:423-63.
1022. Zazpe A, Artaiz I, Del Río J. Role of 5-HT₃ receptors in basal and K(+)-evoked dopamine release from rat olfactory tubercle and striatal slices. *British Journal of Pharmacology*. 1994;113(3):968-72.
1023. Blandina P, Goldfarb J, Craddock-Royal B, Green JP. Release of endogenous dopamine by stimulation of 5-hydroxytryptamine₃ receptors in rat striatum. *J Pharmacol Exp Ther*. 1989;251(3):803-9.
1024. Sorensen SM, Humphreys TM, Palfreyman MG. Effect of acute and chronic MDL 73,147EF, a 5-HT₃ receptor antagonist, on A9 and A10 dopamine neurons. *Eur J Pharmacol*. 1989;163(1):115-8.
1025. Navailles S, Lagiere M, Contini A, De Deurwaerdere P. Multisite intracerebral microdialysis to study the mechanism of L-DOPA induced dopamine and serotonin release in the parkinsonian brain. *ACS Chem Neurosci*. 2013;4(5):680-92.
1026. Zoli M, Jansson A, Sykova E, Agnati LF, Fuxe K. Volume transmission in the CNS and its relevance for neuropsychopharmacology. *Trends in pharmacological sciences*. 1999;20(4):142-50.
1027. Venton BJ, Zhang H, Garris PA, Phillips PE, Sulzer D, Wightman RM. Real-time decoding of dopamine concentration changes in the caudate-putamen during tonic and phasic firing. *J Neurochem*. 2003;87(5):1284-95.
1028. Utter AA, Basso MA. The basal ganglia: an overview of circuits and function. *Neurosci Biobehav Rev*. 2008;32(3):333-42.
1029. Belforte JE, Pazo JH. Turning behaviour induced by stimulation of the 5-HT receptors in the subthalamic nucleus. *Eur J Neurosci*. 2004;19(2):346-55.
1030. Iderberg H, Maslava N, Thompson AD, Bubser M, Niswender CM, Hopkins CR, et al. Pharmacological stimulation of metabotropic glutamate receptor type 4 in a rat model of Parkinson's disease and L-DOPA-induced dyskinesia: Comparison between a positive allosteric modulator and an orthosteric agonist. *Neuropharmacology*. 2015;95:121-9.

1031. Putterman DB, Munhall AC, Kozell LB, Belknap JK, Johnson SW. Evaluation of levodopa dose and magnitude of dopamine depletion as risk factors for levodopa-induced dyskinesia in a rat model of Parkinson's disease. *J Pharmacol Exp Ther.* 2007;323(1):277-84.
1032. Bastide MF, Dovero S, Charron G, Porras G, Gross CE, Fernagut P-O, et al. Immediate-early gene expression in structures outside the basal ganglia is associated to l-DOPA-induced dyskinesia. *Neurobiology of Disease.* 2014;62(Supplement C):179-92.
1033. Monville C, Torres EM, Dunnett SB. Comparison of incremental and accelerating protocols of the rotarod test for the assessment of motor deficits in the 6-OHDA model. *Journal of neuroscience methods.* 2006;158(2):219-23.
1034. Leite-Almeida H, Almeida-Torres L, Mesquita AR, Pertovaara A, Sousa N, Cerqueira JJ, et al. The impact of age on emotional and cognitive behaviours triggered by experimental neuropathy in rats. *Pain.* 2009;144(1-2):57-65.
1035. Olsson M, Nikkhah G, Bentlage C, Bjorklund A. Forelimb akinesia in the rat Parkinson model: differential effects of dopamine agonists and nigral transplants as assessed by a new stepping test. *J Neurosci.* 1995;15(5 Pt 2):3863-75.
1036. Carta M, Lindgren HS, Lundblad M, Stancampiano R, Fadda F, Cenci MA. Role of striatal L-DOPA in the production of dyskinesia in 6-hydroxydopamine lesioned rats. *J Neurochem.* 2006;96(6):1718-27.
1037. Gerfen CR. Molecular effects of dopamine on striatal-projection pathways. *Trends in neurosciences.* 2000;23(10 Suppl):S64-70.
1038. Andersson M, Hilbertson A, Cenci MA. Striatal fosB expression is causally linked with l-DOPA-induced abnormal involuntary movements and the associated upregulation of striatal prodynorphin mRNA in a rat model of Parkinson's disease. *Neurobiol Dis.* 1999;6(6):461-74.
1039. Pavon N, Martin AB, Mendiola A, Moratalla R. ERK phosphorylation and FosB expression are associated with L-DOPA-induced dyskinesia in hemiparkinsonian mice. *Biol Psychiatry.* 2006;59(1):64-74.
1040. Cenci MA, Ohlin KE, Odin P. Current options and future possibilities for the treatment of dyskinesia and motor fluctuations in Parkinson's disease. *CNS Neurol Disord Drug Targets.* 2011;10(6):670-84.
1041. Bezard E, Munoz A, Tronci E, Pioli EY, Li Q, Porras G, et al. Anti-dyskinetic effect of anpirtoline in animal models of L-DOPA-induced dyskinesia. *Neurosci Res.* 2013;77(4):242-6.
1042. Huot P, Sgambato-Faure V, Fox SH, McCreary AC. Serotonergic Approaches in Parkinson's Disease: Translational Perspectives, an Update. *ACS Chem Neurosci.* 2017;8(5):973-86.

VII. Appendix

Table III: Duration rating scale of ALO AIMs in the 6-OHDA-lesioned rat

Parameter	Score
axial	0: no dyskinesia 1: occasional signs of dyskinesia, present < 50% of observation time 2: frequent signs of dyskinesia, present > 50% of the observation time 3: dyskinesia present during the entire observation period, but suppressible by external stimuli 4: continuous dyskinesia not suppressible by external stimuli
limbs	0: no dyskinesia 1: occasional signs of dyskinesia, present < 50% of observation time 2: frequent signs of dyskinesia, present > 50% of the observation time 3: dyskinesia present during the entire observation period, but suppressible by external stimuli 4: continuous dyskinesia not suppressible by external stimuli
orolingual	0: no dyskinesia 1: occasional signs of dyskinesia, present < 50% of observation time 2: frequent signs of dyskinesia, present > 50% of the observation time 3: dyskinesia present during the entire observation period, but suppressible by external stimuli 4: continuous dyskinesia not suppressible by external stimuli

Table IV: Amplitude rating scale of ALO AIMs in the 6-OHDA-lesioned rat

Parameter	Score
axial	<p>0: no dyskinesia</p> <p>1: sustained deviation of the head and neck at about a 30° angle</p> <p>2: sustained deviation of the head and neck between an angle of 30° and 60°</p> <p>3: sustained twisting of the head, neck and upper trunk, at an angle between 60° and 90°</p> <p>4: sustained twisting of the head, neck and trunk at maximal amplitude, causing the rat to lose balance from a bipedal position</p>
limbs	<p>0: no dyskinesia</p> <p>1: tiny movements of the paw around a fixed position</p> <p>2: displacement of the whole limb (horizontal or up-and-down)</p> <p>3: large displacement of the limb with visible contraction of shoulder muscles</p> <p>4: vigorous limb displacement of maximal amplitude, with contraction of both shoulder groups and extensor muscles</p>
orolingual	<p>0: no dyskinesia</p> <p>1: twitching of facial muscles accompanied by small masticatory movements without jaw opening</p> <p>2: twitching of facial muscles accompanied by masticatory movements, occasional jaw opening</p> <p>3: movements involving facial muscles and masticatory muscles, frequent jaw opening and occasional tongue protrusion</p> <p>4: involvement of all of the above muscles to the maximal possible degree</p>

Table V: Glass' delta of right forepaw use across ondansetron treatments

dose of ondansetron (mg/mL)	lesion	veh	0.0001	0.001	0.01	0.1	1
mean	82.31	50.79	52.07	43.17	46.03	34	43.98
standard deviation	11.57	27.67	24.09	28.38	24.64	25.76	21.15
Glass' delta	-	2.72	2.61	3.38	3.14	4.18	3.31

Table VI: Glass' delta of both forepaw use across ondansetron treatments

dose of ondansetron (mg/mL)	lesion	veh	0.0001	0.001	0.01	0.1	1
mean	17.09	41.03	34.83	35.28	34.88	45.95	34.56
standard deviation	11.26	22.49	19.93	26.59	21.54	24.85	15.15
Glass' delta	-	2.13	1.58	1.62	1.58	2.56	1.55

Figure 11: Equation to calculate Glass' delta

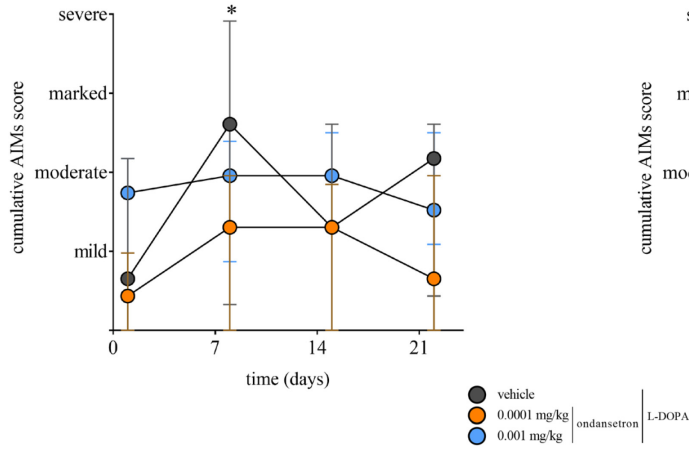
$$\text{Glass' delta} = \frac{|\bar{X}_2 - \bar{X}_1|}{\sqrt{s^2_{\text{control}}}}$$

\bar{X}_1 = mean of control group

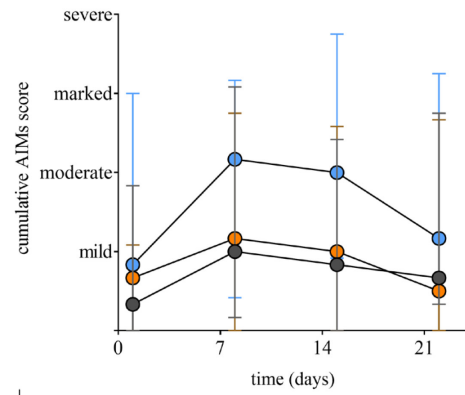
\bar{X}_2 = mean of experimental group

s_{control} = standard deviation of control group

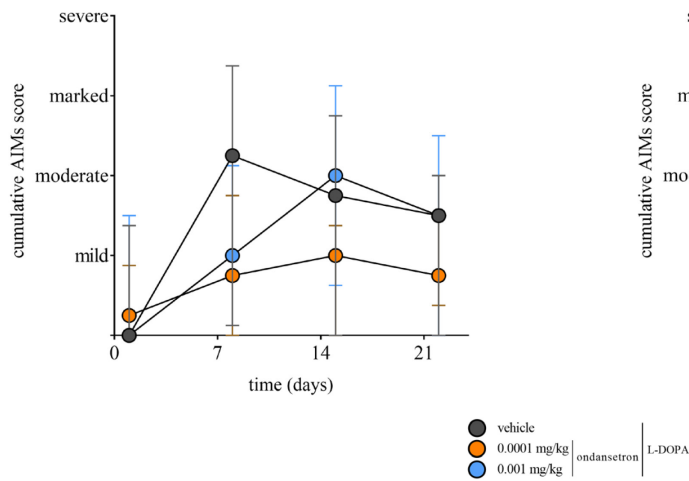
A axial AIMS duration development timecourse



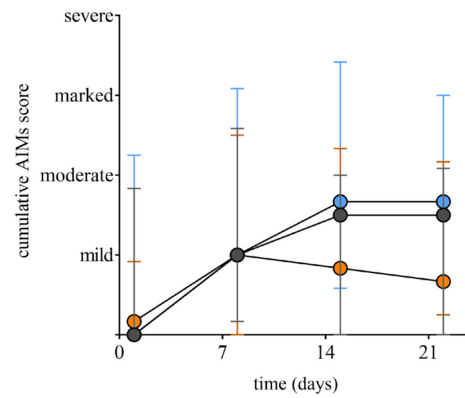
B axial AIMS amplitude development timecourse



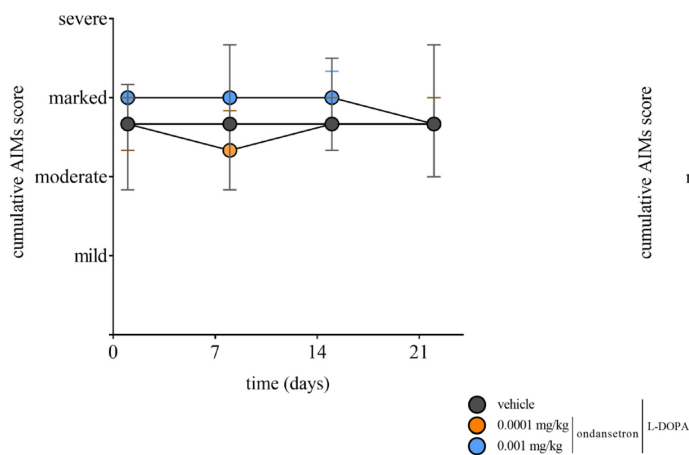
C limbs AIMS duration development timecourse



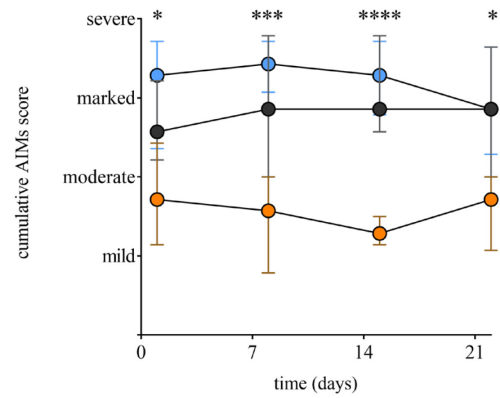
D limbs AIMS amplitude development timecourse



E orolingual AIMS duration development timecourse



F orolingual AIMS amplitude development timecourse



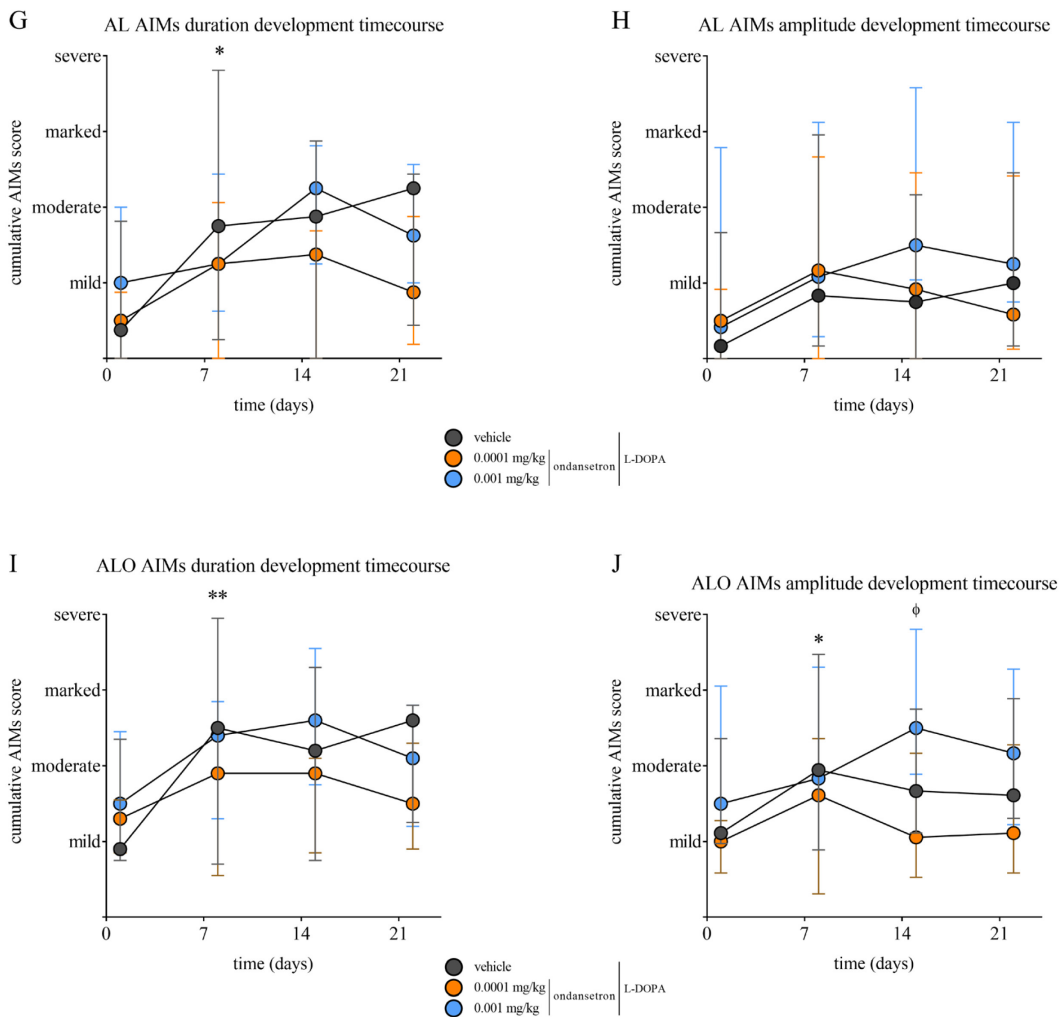


Figure 12: Time course of the development of AIMs during the 22-day priming phase of the *de novo* ondansetron study. On day 8 of priming, **A.** the duration of axial AIMs was significantly reduced in animals previously treated with L-DOPA/0.0001 mg/kg ondansetron ($n = 9$), by 53%, when compared with L-DOPA/vehicle ($n = 7$) ($P < 0.05$, Tukey's *post hoc* test). **B.** In contrast, the amplitude of axial AIMs was not significantly reduced in animals that were previously treated with L-DOPA/ondansetron 0.0001 mg/kg ($n = 9$) compared to animals that received L-DOPA/vehicle. **C.** The duration and **D.** amplitude of limbs AIMs was comparable between animals administered ondansetron 0.0001 mg/kg and vehicle. **E.** The duration of orolingual AIMs was not affected by ondansetron treatment. **F.** Administration of L-DOPA/ondansetron 0.0001 mg/kg resulted in a significant decrease in the amplitude of orolingual AIMs on days 1, 8, 5 and 22 by 21%, 32%, 37% and 24%, respectively, when

compared with vehicle ($P < 0.05$, $P < 0.001$, $P < 0.0001$ and $P < 0.05$, Tukey's *post hoc* test). **G.** On day 8, when administered with L-DOPA, 0.0001 mg/kg ondansetron significantly reduced the duration of AL AIMs, by 53%, when compared with L-DOPA/vehicle ($P < 0.05$, Tukey's *post hoc* test). **H.** The amplitude of AL AIMs was comparable between L-DOPA/0.0001 mg/kg ondansetron and L-DOPA/vehicle. **I.** Administration of ondansetron 0.0001 mg/kg resulted in a significant decrease in the duration of ALO AIMs on day 15, by 33%, when compared with L-DOPA/vehicle ($P < 0.01$, Tukey's *post hoc* test). **J.** The amplitude of ALO AIMs was reduced in animals that were treated with L-DOPA/ondansetron 0.0001 mg/kg, by 31%, compared to animals that received L-DOPA/vehicle ($P < 0.05$, Tukey's *post hoc* test). Data are graphed as the median with interquartile range. *: $P < 0.05$; **: $P < 0.01$, ***: $P < 0.001$, ****: $P < 0.0001$ vehicle versus ondansetron 0.0001 mg/kg; ϕ : $P < 0.01$ ondansetron 0.0001 mg/kg versus ondansetron 0.001 mg/kg.