CROSS-CANADA DISEASE REPORT RAPPORT DES MALADIES DIAGNOSTIQUÉES AU CANADA Canine parvovirus type 2b is the most prevalent genomic variant strain found in parvovirus antigen positive diarrheic dog feces samples across Canada. Carl A. Gagnon^{a,b,c,*}, Véronique Allard^c, Guillaume Cloutier^d. ^aGroupe de recherche sur les maladies infectieuses du porc (GREMIP), ^bSwine and Poultry Infectious Diseases Research Center (CRIPA) and ^cMolecular diagnostic laboratory, Faculté de médecine vétérinaire, Université de Montréal, Saint-Hyacinthe, Quebec, Canada. ^dBoehringer Ingelheim (Canada) Ltd, Burlington, Ontario, Canada. *Corresponding author: Mailing address: Faculté de médecine vétérinaire, Université de

Montréal, 3200 rue Sicotte, Saint-Hyacinthe, Québec, Canada, J2S 7C6. Phone: 450-773-

8521 (8681). Fax: 450-778-8108. Email: carl.a.gagnon@umontreal.ca.

In 1978 an emerging virus was identified in dog population, the canine parvovirus (CPV) which was subsequently named CPV type 2 (CPV-2). The latter was distinct from known parvovirus; the canine minute virus (CnMV), formerly known as canine parvovirus type 1 (1). CPV-2 was inducing hemorrhagic enteritis, severe diarrhea, vomiting, leukopenia associated with a high mortality in infected dogs and had spread into non-immune dog populations worldwide causing a pandemic. Furthermore, CPV-2 was responsible for myocarditis in puppies. Today, the prevalence of CPV antibodies positive adult dogs is high due to vaccination and/or natural infection causing a high protective immunity status. Nevertheless, young non-immune animals of 6 weeks to 6 months old are susceptible to CPV infection when passive protection from maternal derived antibodies (MDA), which are able to protect puppies against myocarditis, have been lost. (2). The CPV-2 belongs to the virus family *Parvoviridae*, subfamilly *Parvovirinae* (which infect vertebrates) and genus *Parvovirus* (3). CPV-2 is a small non-enveloped single-stranded DNA virus of around 25 nm in size (1). The CPV-2 genome length is 5.2 kilo bases (kb) and possesses at least two major open reading frames (ORFs) (4). In the conventional orientation, the right-hand ORF encodes the viral capsid proteins VP1 and VP2, which are the main antigens that induce protective antibodies (5-7). The left-hand ORF encodes the non-structural proteins NS1 and NS2 (8). Since PCV-2 emergence, several viral genomic variants have emerged due to a surprisingly high substitution rate in CPV genome similar to those of RNA viruses (1, 9). At least four genomic variants have been described and officially recognized: the initial emerging CPV (named afterward CPV-2), followed shortly after by the CPV-2a (in 1979), the CPV-2b (in 1984) and CPV-2c (in 2000) (1). These CPV variants are differentiated through single nucleotide

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

polymorphism (SNP) located at the VP2 gene (10). The prevalence of CPV variants are geographically restricted and have evolved over time (1, 10). Unfortunately, to our knowledge, the prevalence of CPV genomic variants in Canada is not known. Thus, the main objective of the present study was to determine the prevalence of CPV genomic variants within Canadian diarrheic dog population.

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

67

68

69

47

48

49

50

51

To fulfill this objective, Canadian veterinarian volunteers have submitted feces from diarrheic animals that were previously confirmed to be positive for CPV antigen after local testing by any commercially available capture antigen assay (such as SNAP® Parvo Test, Idexx Laboratories). The CPV vaccination status of diarrheic animals was requested and when they were known to be recently vaccinated, i.e. less than 30 days after the last vaccination, the feces samples of those animals were excluded from the study. It has been recently reported that animals that are vaccinated with a live attenuated vaccine may shed the vaccine strain for up to 21 days post-vaccination, as established by CPV real-time PCR positive results (11). Thus, a delay of 30 days post-vaccination for CPV genotyping was sufficient to circumvent the effects of vaccine strains on the results. Overall, samples from 49 cases were included in this study. These samples were submitted over a three year period (from September 27th 2012 until August 21st 2015) and originated from seven different provinces (see Table 1). Genotyping of CPV variants was performed by the concomitant analysis of three multiplex real-time PCR (qPCR) assays using minor groove binder (MGB) probes technology as previously described (12, 13) but with minor modifications. These MGB probes allow SNP discrimination of the ORF encoding the VP2 protein and therefore allow strains genotyping. Briefly, about 1

gram of feces was resuspended in 5 mL of phosphate buffer saline (PBS). Nucleic acid extraction was performed on supernatants or directly on vaccine formulation (Duramune®Max Pv and Nobivac®Canine 3-DAPv which contains the CPV-2b and CPV-2 variants, respectively, as positive controls) with either the QIAamp cador Pathogen Mini Kit (Qiagen) or the BioSprint 96 One-For-All vet kit (Qiagen) according to manufacturer's instructions. Thereafter, the three multiplex qPCR assays were carried out in a 25 µL volume containing 5 µL template or control DNA, 12.5 µL of QuantiTect Probe PCR buffer 2x (Qiagen), 3 pmol of TET-labeled probe, and 1.25 µL of a 20X mixture consisting of forward and reverse primer and the FAM-labeled probe. The qPCR were either performed on a SmartCycler (Cepheid) or RotorGene (Qiagen). The first conducted multiplex qPCR assay was targeting the change from adenine to guanine at position 4062 (SNP A4062G) in Genebank reference strain M38245 between CPV-2a and 2b variants, while the second multiplex qPCR assay was targeting the thymine to adenine change at position 4064 (SNP T4064A) between CPV-2b and 2c variants. As previously reported, the first multiplex qPCR assay could not properly discriminate CPV-2 and 2a variants. Thus, a third multiplex qPCR assay targeting SNP T3088C was developed to ensure discrimination between CPV-2 variant from all three other variants. Taken together, these three multiplex qPCR assays allow reliable and fast CPV genotyping results.

89

90

91

92

70

71

72

73

74

75

76

77

78

79

80

81

82

83

84

85

86

87

88

As shown in Table 1, the CPV-2b variant was the most prevalent in all the seven provinces from which the samples were obtained. The prevalence of CPV-2b between the different provinces was varying from 85.7 to 100%. Overall, the Canadian CPV-2b

prevalence was 89.8%. This finding was surprising because since 2007, CPV-2c was reported to be the most prevalent variant into the United States (US), the Canada neighboring country, varying between 48.1 to 73.5% (14, 15). In contrast, only one sample originating from Ontario was positive for CPV-2c variant in the current study (Table 1). Consequently, CPV-2c has an overall Canadian prevalence of 2%, which is much lower as compared to US prevalence. Four samples were positive for CPV-2a variant and they were originating from three provinces: Alberta, British Columbia and Quebec. Thus, CPV-2a has an overall Canadian prevalence of 8.2% (Table 1). As expected, no CPV-2 variant was detected as it is considered to be extinct (Table 1). Because vaccination could significantly affect the genotyping of CPV as attenuated vaccine strains could be shed in feces (11), information in regards to vaccination status was requested with the submitted samples. Unfortunately, the vaccination status of seven animals (i.e. 14.3% of the feces samples) was unknown (data not shown). Thus, even if those seven cases are excluded from our analyses, the CPV-2b variant remained the most prevalent variant found in diarrheic dogs with a value of 90.5% (data not shown). Interestingly, 19 diarrheic animals have been reported by veterinarians to be vaccinated and at least three different commercial vaccines have been used to vaccinate animals (data not shown). Noteworthy, only 1 of these animals (5.3%) seems to have been vaccinated according to the equivalent guidelines of the American Animal Hospital Association (AAHA) and the World Small Animal Veterinary Association (WSAVA). These guidelines are that: "Puppies with poor MDA may be vulnerable (and capable of responding to vaccination) at an earlier age, while others may possess MDA at such high titres that they are incapable of responding to vaccination until ≥ 12 weeks of age. No

93

94

95

96

97

98

99

100

101

102

103

104

105

106

107

108

109

110

111

112

113

114

single primary vaccination policy will therefore cover all possible situations. Thus, puppies should be vaccinated every 3-4 weeks between the ages of 6 and 16 weeks (e.g., at 6, 10, and 14 weeks, or 8, 12, and 16 weeks). To minimize the risk of MDA interference with vaccination, the final dose of the initial series should be administered between 14 and 16 weeks of age, regardless of the product used" (16, 17). Thus, at least 83.7% (n=41) of the diarrheic animals of this study were not or were improperly vaccinated (data not shown) because they received only one or two doses or the last puppy booster was given prior to 14 weeks of age. Noteworthy, three of the vaccinated animals received three vaccine shots as recommended by AAHA and WSAVA but surprisingly, the last booster shot was given prior to 14 weeks of age (data not shown). Consequently, Canadian veterinarians should encourage their pet owners to vaccinate their puppies following the AAHA and WSAVA guidelines. Several reports have demonstrated a very good efficacy of CPV vaccines in the context of homologous and heterologous challenges (10, 17). Our results may have indicated a vaccination failure in one vaccinated animal but it might be explain by errors included in the clinical data that were reported by the owner and/or the veterinarian. Moreover, other co-infecting microorganisms (bacteria, parasites, viruses) could have contributed to the diarrhea in the clinical cases that were submitted (18). Unfortunately, the intestinal microbiota of diarrheic animals was not part of this study and thus, it was not investigated. In conclusion, by far the most prevalent CPV variant found in feces of Canadian diarrheic dogs is CPV-2b.

136

137

135

116

117

118

119

120

121

122

123

124

125

126

127

128

129

130

131

132

133

134

Acknowledgements

- 138 This project was sponsored by Boehringer Ingelheim (Canada) Ltd. C.A. Gagnon was
- 139 financially supported by a Natural Sciences and Engineering Research Council of Canada
- 140 (NSERC) discovery grant. The authors wish to thank all the Canadian veterinarians who
- have voluntarily participated to this project by submitting diarrheic dog feces samples.
- Nucleic acids of CPV-2a and 2c variants were kindly provided by Dr Nicola Decaro
- 143 (University of Bari, Italy) and were used as positive controls to validate our qPCR
- 144 multiplex assays.

145

146

References

- 147 1. Hoelzer K Parrish CR. The emergence of parvoviruses of carnivores. Veterinary
- research 2010;41:39.
- 149 2. Thiry E. Virologie clinique du chien et du chat: Éditions du Point vétérinaire,
- 150 2002:203.
- 151 3. King AMQ, Adams MJ, Carstens EB, Lefkowitz EJ. Virus Taxonomy,
- 152 Classification and Nomenclature of Viruses, Ninth Report of the International
- 153 Committee on Taxonomy of Viruses (ICTV): Elsevier, Academic Press,
- 154 2012:1327.
- 155 4. Reed AP, Jones EV, Miller TJ. Nucleotide sequence and genome organization of
- canine parvovirus. Journal of virology 1988;62:266-276.
- 157 5. Nelson CD, Palermo LM, Hafenstein SL, Parrish CR. Different mechanisms of
- antibody-mediated neutralization of parvoviruses revealed using the Fab
- fragments of monoclonal antibodies. Virology 2007;361:283-293.
- 160 6. Langeveld JP, Casal JI, Cortes E, et al. Effective induction of neutralizing
- antibodies with the amino terminus of VP2 of canine parvovirus as a synthetic
- peptide. Vaccine 1994;12:1473-1480.
- 163 7. Casal JI, Langeveld JP, Cortes E, et al. Peptide vaccine against canine parvovirus:
- identification of two neutralization subsites in the N terminus of VP2 and
- optimization of the amino acid sequence. Journal of virology 1995;69:7274-7277.

- Wang D, Yuan W, Davis I,Parrish CR. Nonstructural protein-2 and the replication of canine parvovirus. Virology 1998;240:273-281.
- 168 9. Shackelton LA, Parrish CR, Truyen U,Holmes EC. High rate of viral evolution
- associated with the emergence of carnivore parvovirus. Proceedings of the
- National Academy of Sciences of the United States of America 2005;102:379-
- 171 384.
- 172 10. Decaro N Buonavoglia C. Canine parvovirus--a review of epidemiological and
- diagnostic aspects, with emphasis on type 2c. Veterinary microbiology
- 174 2012;155:1-12.
- 175 11. Decaro N, Crescenzo G, Desario C, et al. Long-term viremia and fecal shedding
- in pups after modified-live canine parvovirus vaccination. Vaccine 2014;32:3850-
- 177 3853.
- 178 12. Decaro N, Elia G, Desario C, et al. A minor groove binder probe real-time PCR
- assay for discrimination between type 2-based vaccines and field strains of canine
- parvovirus. Journal of virological methods 2006;136:65-70.
- 181 13. Decaro N, Elia G, Martella V, et al. Characterisation of the canine parvovirus type
- 2 variants using minor groove binder probe technology. Journal of virological
- methods 2006;133:92-99.
- 184 14. Kapil S, Cooper E, Lamm C, et al. Canine parvovirus types 2c and 2b circulating
- in North American dogs in 2006 and 2007. Journal of clinical microbiology
- 186 2007;45:4044-4047.
- 187 15. Markovich JE, Stucker KM, Carr AH, Harbison CE, Scarlett JM, Parrish CR.
- Effects of canine parvovirus strain variations on diagnostic test results and clinical
- management of enteritis in dogs. Journal of the American Veterinary Medical
- 190 Association 2012;241:66-72.
- 191 16. Day MJ, Horzinek MC, Schultz RD. WSAVA guidelines for the vaccination of
- dogs and cats. The Journal of small animal practice 2010;51:1-32.
- 193 17. Welborn LV, DeVries JG, Ford R, et al. 2011 AAHA canine vaccination
- guidelines. Journal of the American Animal Hospital Association 2011;47:1-42.

18. Gizzi AB, Oliveira ST, Leutenegger CM, et al. Presence of infectious agents and co-infections in diarrheic dogs determined with a real-time polymerase chain reaction-based panel. BMC veterinary research 2014;10:23.

Table 1. Prevalence of canine parvovirus type 2 genomic variants in feces of diarrheic dogs

Province Nova Scotia	Total of submitted feces samples ^a	Genomic variants % (n)							
		CPV-2		CPV-2a		CPV-2b		CPV-2c	
		0,0	(0)	0,0	(0)	100,0	(3)	0,0	(0)
New Brunswick	1	0,0	(0)	0,0	(0)	100,0	(1)	0,0	(0)
Quebec	10	0,0	(0)	10,0	(1)	90,0	(9)	0,0	(0)
Ontario	13	0,0	(0)	0,0	(0)	92,3	(12)	7,7	(1)
Saskatchewan	1	0,0	(0)	0,0	(0)	100,0	(1)	0,0	(0)
Alberta	7	0,0	(0)	14,3	(1)	85,7	(6)	0,0	(0)
British Columbia	14	0,0	(0)	14,3	(2)	85,7	(12)	0,0	(0)
Total:	49	0,0	(0)	8,2	(4)	89,8	(44)	2,0	(1)

^aSamples were submitted from CPV antigen positive diarrheic animals starting 2012/09/27 until 2015/08/21.